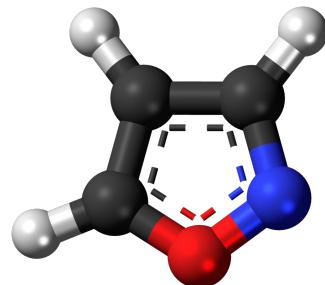


# **Application of Isoxazole in Chemoproteomics as Novel Photo-cross-linker**



230408

Junhao Fu

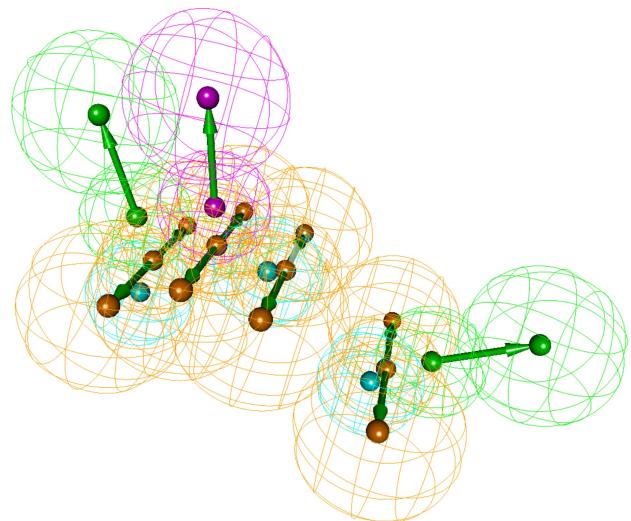
# **Contents**

## **1. introduction**

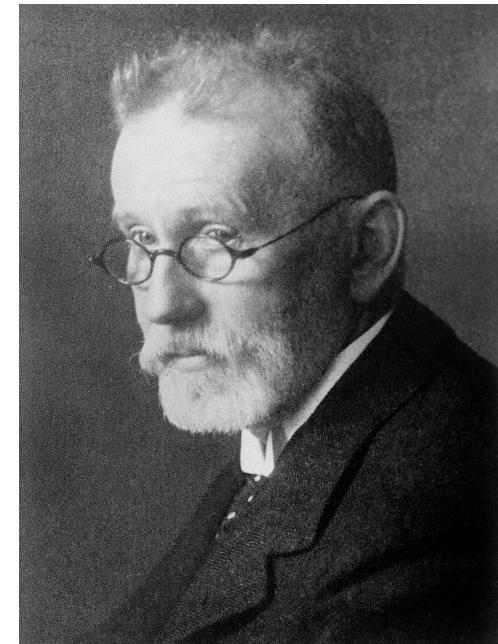
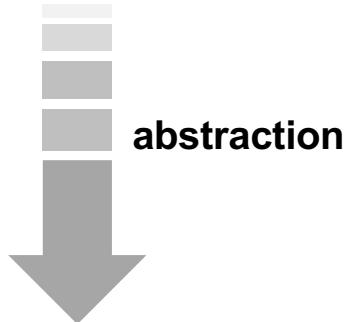
## **2. development of isoxazole-based novel photo-cross-linker — main**

**Cheng, K.; Qi, J.; Ren, X.; Zhang, J.; Li, H.; Xiao, H.; Wang, R.; Liu, Z.; Meng, L.; Ma, N.; Sun. H. *Angew. Chem., Int. Ed.* 2022, 61, e202209947.**

# Pharmacophore & Molecular Interaction



a molecular framework that carries the essential features for a drug's biological activity (1909)



Paul Ehrlich (1854~1915)  
Nobel Prize in physiology or medicine

an ensemble of **steric and electronic features** that is necessary to ensure the optimal supramolecular interactions with a specific biological target and to trigger (or block) its biological response (1997, IUPAC)

# Pharmacophoric Features

| feature type                 | complementary feature type | interaction type               | structural examples   |
|------------------------------|----------------------------|--------------------------------|---|
| hydrogen bond acceptor (HBA) | HBD                        | hydrogen bonding               | amines, carboxylates, ketones, alcohols, fluorine substituents, ...               |
| hydrogen bond donor (HBD)    | HBA                        | hydrogen bonding               | amines, amides, alcohols, ...   |
| aromatic (AR)                | AR, PI                     | $\pi$ -stacking, cation- $\pi$ | aromatic rings  |
| positive ionizable (PI)      | AR, NI                     | ionic, cation- $\pi$           | ammonium ions, metal cations, ...   |
| negative ionizable (NI)      | PI                         | ionic                          | carboxylates  |
| hydrophobic (H)              | H                          | hydrophobic interaction        | halogen substituents, alkyl groups, alicyclic rings, weak or non-polar atoms, ... |

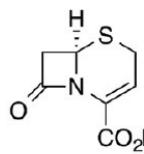
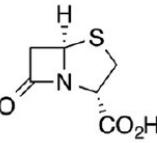
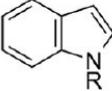
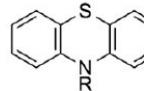
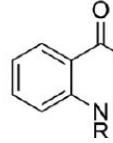
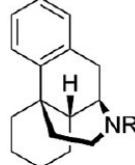
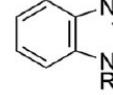
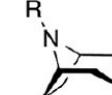
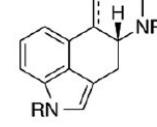
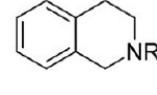
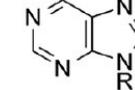
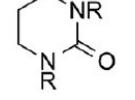


**potential ligands of biomolecules** generally posses these features

frequent involvement of **N atom**, especially **N-heterocycles**

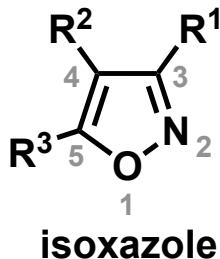
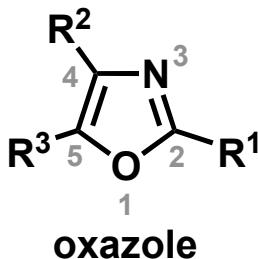
# N-Heterocycles in FDA-Proved Drugs

among FDA-proved unique small-molecule drugs: N-atom: 84%, N-heterocycle: 59%

|   |  |   |   |  |   |  |
|---|--|---|---|--|---|--|
| #1<br><br>piperidine             | #2<br><br>pyridine  | #3<br><br>piperazine         | #4<br><br>cephem         | #5<br><br>pyrrolidine     | #6<br><br>thiazole                     | #7<br><br>imidazole     |
| #8<br><br>penam                  | #9<br><br>indole    | #10<br><br>tetrazole         | #10<br><br>phenothiazine | #10<br><br>pyrimidine     | #13<br><br>4-quinolinone               | #13<br><br>morphinan    |
| #15<br><br>benzimidazole        | #15<br><br>tropane | #17<br><br>morpholine       | #17<br><br>ergoline     | #19<br><br>imidazolidine | #19<br><br>tetrahydroisoquinoline     | #21<br><br>imidazoline |
| #21<br><br>1,4-dihydropyridine | #21<br><br>purine | #24<br><br>1,2,4-triazole | #24<br><br>isoxazole   | #24<br><br>quinazoline  | #24<br><br>tetrahydro-2-pyrimidinone |  |

# Isoxazole and Its Occurrence

core structure:



- electron rich aromatic ring with hydrogen bond acceptors
- interaction with biomolecules:
  - hydrogen bonding
  - pi-pi stacking
- good solubility
- bio-compatibility



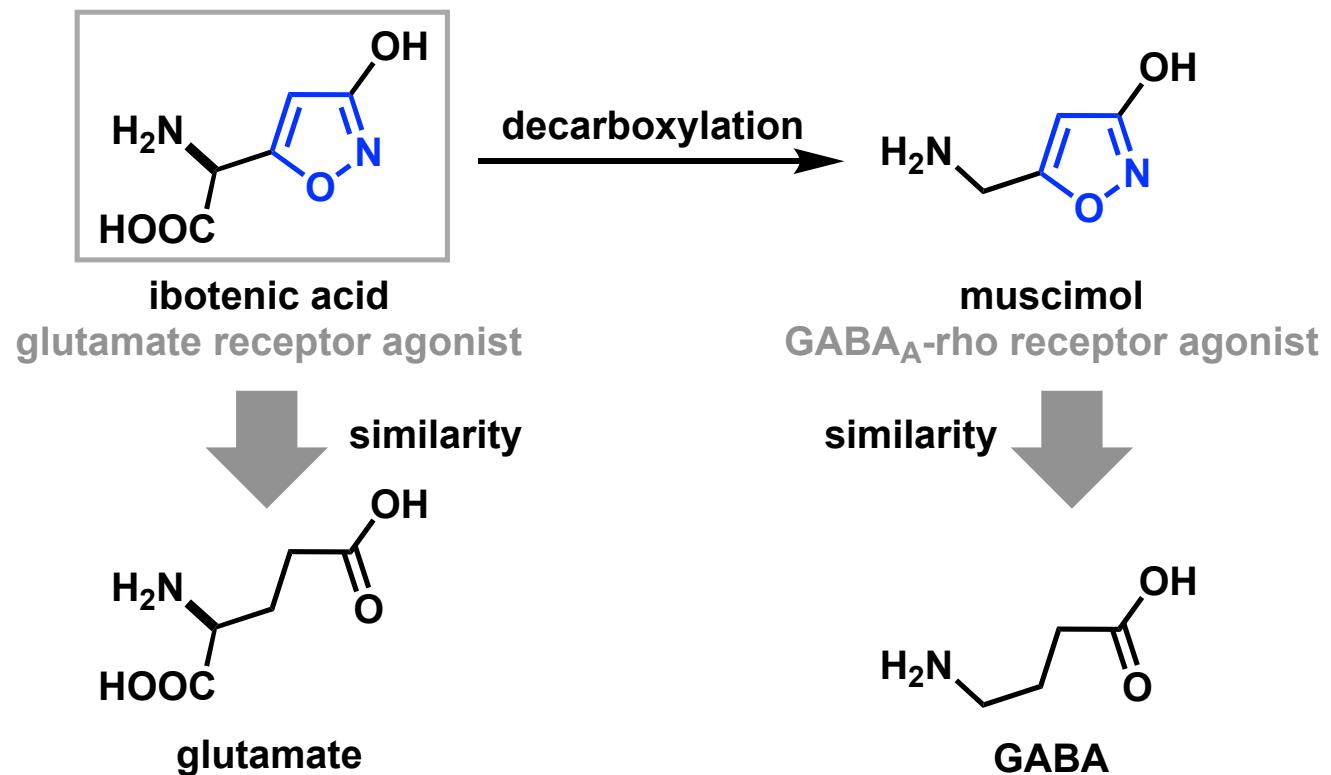
potential in drug discovery

occurrence in nature: ibotenic acid

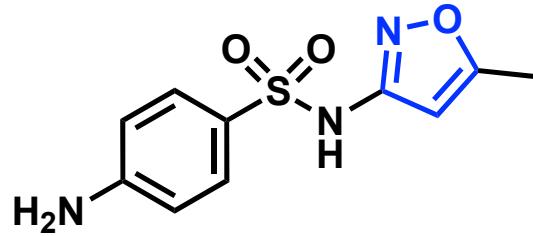


*Amanita muscaria*

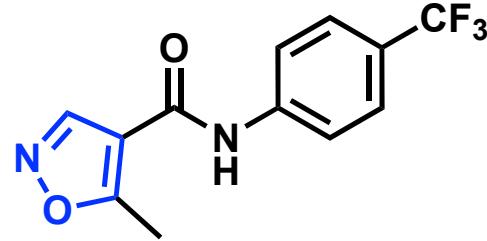
- isolated by Takemoto *et al.* in 1964
- psychoactive
- powerful neurotoxin
- 10-fold stronger ‘umami’ than glutamic acid (味の素) ...



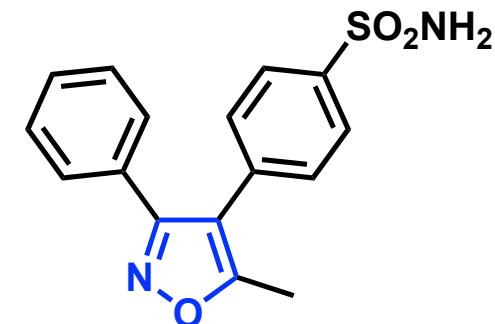
# Isoxazole in Bioactive Molecules



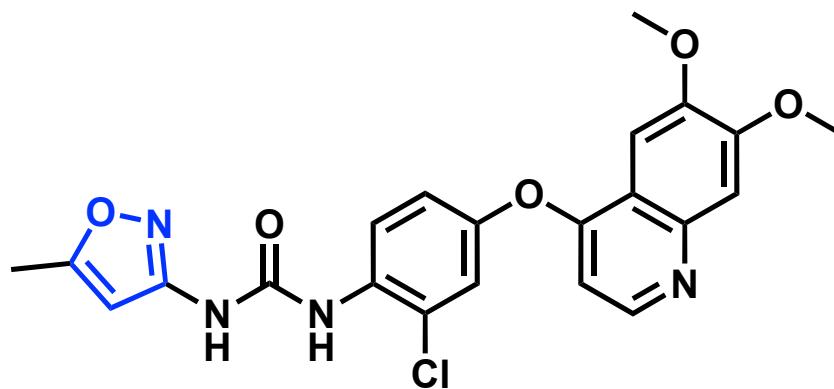
**Sulfamethoxazole (1968)**  
antibacterial



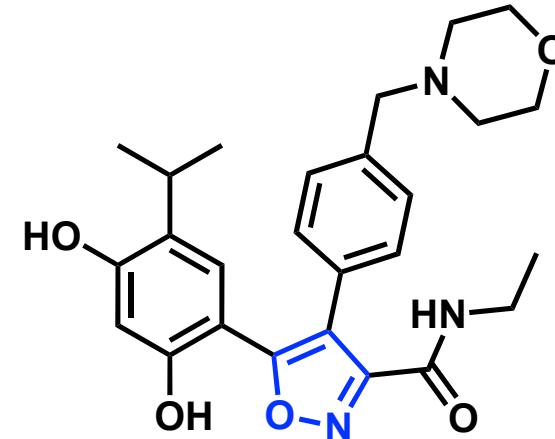
**Leflunomide (1998)**  
anti-rheumatic



**Valdecoxib (2001)**  
anti-inflammatory



**Tivozanib (2021)**  
anti-cancer



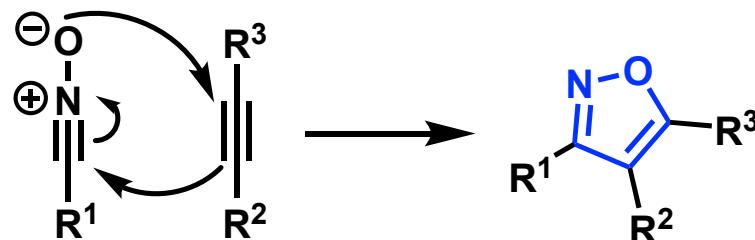
**Luminespib (phase II)**  
anti-cancer

wide variety of bioactivities in isoxazole-containing molecules:  
antibacterial, antiviral, anti-rheumatic, anti-inflammatory, anti-cancer, anti-diabetic, ...

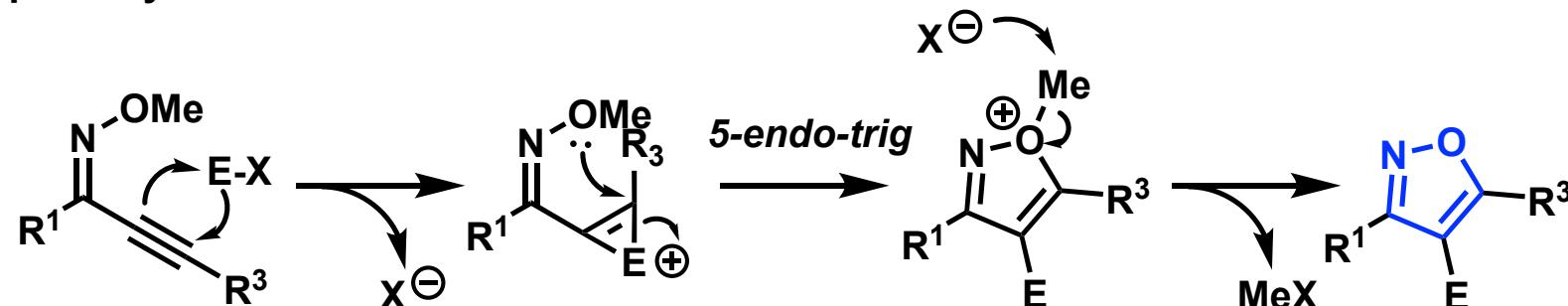
1. Arya, G.C.; Kaur, K.; Jaitak, V. *Eur. J. Med. Chem.* **2021**, 221, 113511;
2. Sysak, A.; Obminska-Mrukowicz, B. *Eur. J. Med. Chem.* **2017**, 137, 292–309.

# Synthetic Availability of Isoxazole

A. [3+2] cycloaddition: Quilico *et al.* (1950)

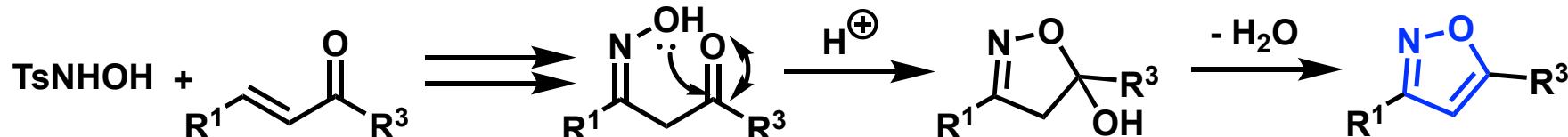


B. electrophilic cyclization: Larock *et al.* 2005



E-X = I<sub>2</sub>, ICl, Br<sub>2</sub>, PhSeBr

C. condensation: She *et al.* (2009)

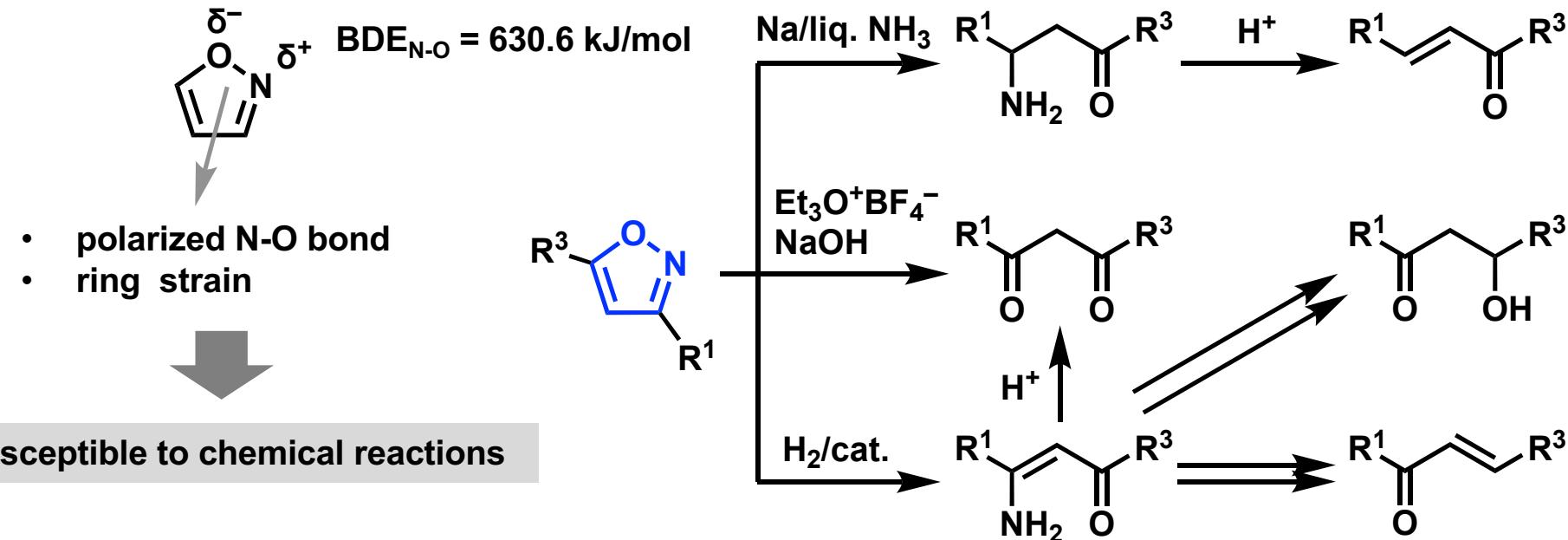


Numerous other routes have also been developed, making it accessible for simple construction of the isoxazole structure.

1. Quilico, A.; Stagno d'Alcontres, G.; Grünanger, P. *Nature* **1950**, *166*, 226–227.
2. Waldo, J.P.; Larock, R.C. *Org. Lett.* **2005**, *7*, 5203–5205.
3. Tang, S.; He, J.; Sun, Y.; He, L.; She, X. *Org. Lett.* **2009**, *11*, 3982–3985.

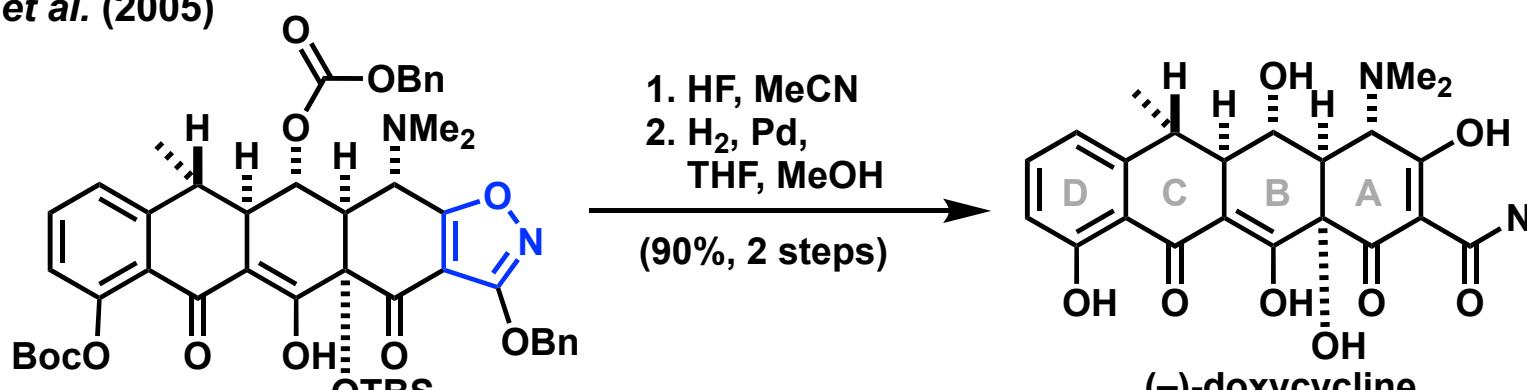
# Synthetic Intermediate Role of Isoxazole

reductive **ring opening** reaction — key reaction as synthetic intermediate:



application in the total synthesis of natural product:

Myers et al. (2005)

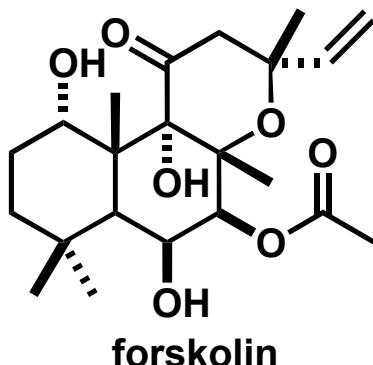


antibacterial activity against Gram-positive strains

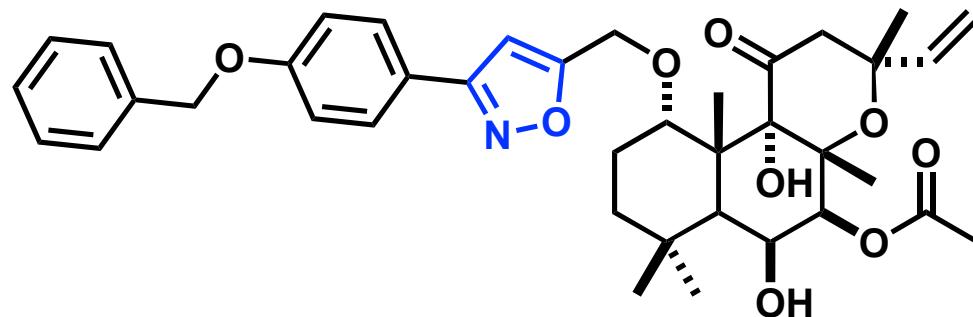
1. Baraldi, P.G.; Barco, A.; Benetti, S.; Pollini, G.P. Simoni, D. *Synthesis* **1987**, 10, 857–869.
2. Li, Z.; Carmichael, I.; Ptasińska, S. *Phys. Chem. Chem. Phys.* **2018**, 20, 18271–18278.
3. Charest, M.G.; Lerner, C.D.; Brubaker, J.D.; Siegel, D.R.; Myers, A.G. *Science* **2005**, 308, 395–398.

# Modification of Natural Products with Isoxazoles

in anticancer natural product:



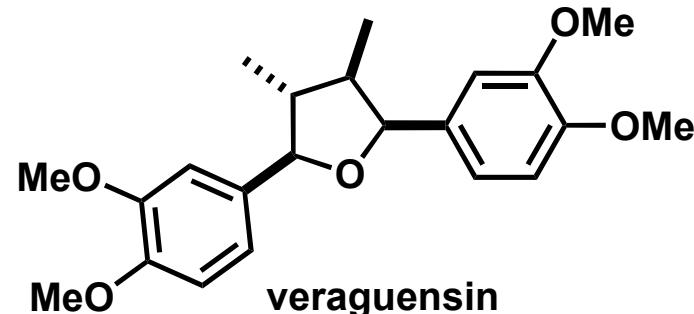
C1-OH  
substitution



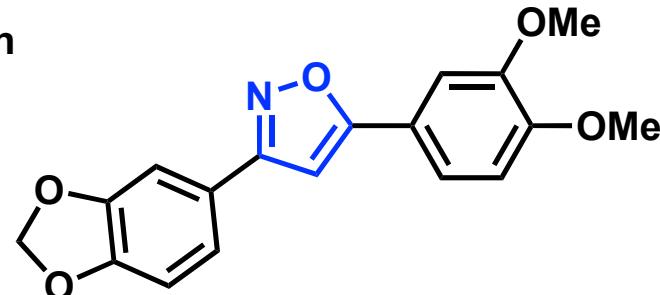
$IC_{50} = 63.3 \mu M$  against MCF-7,  
 $IC_{50} > 100 \mu M$  against BT-474 cells

$IC_{50} = 0.5 \mu M$  against MCF-7,  
 $IC_{50} = 0.5 \mu M$  against BT-474 cells

in antimicrobial natural product:



alternation of  
tetrahydrofuran  
core



antileishmanial activity:  $IC_{50} = 48.8 \mu M$   
(against *L. donovani*)

antileishmanial activity:  $IC_{50} = 2.0 \mu M$   
(against *L. amazonensis*)

introduction of isoxazole to natural products could possibly **change their biological properties**

1. Burra, S.; Voora, V.; Rao, C.P.; Kumar, P. V.; Kancha, R.K.; Krupadanam, G.L.D. *Bioorg. Med. Chem. Lett.* **2017**, 27, 4314–4318.
2. Baroni, A.C.M. et al. *Chem. Biol. Drug Des.* **2019**, 93, 313–324.

# **Contents**

1. introduction
2. development of isoxazole-based novel photo-cross-linker — main  
Cheng, K.; Qi, J.; Ren, X.; Zhang, J.; Li, H.; Xiao, H.; Wang, R.; Liu, Z.; Meng, L.;  
Ma, N.; Sun. H. *Angew. Chem., Int. Ed.* 2022, 61, e202209947.

# Author Information



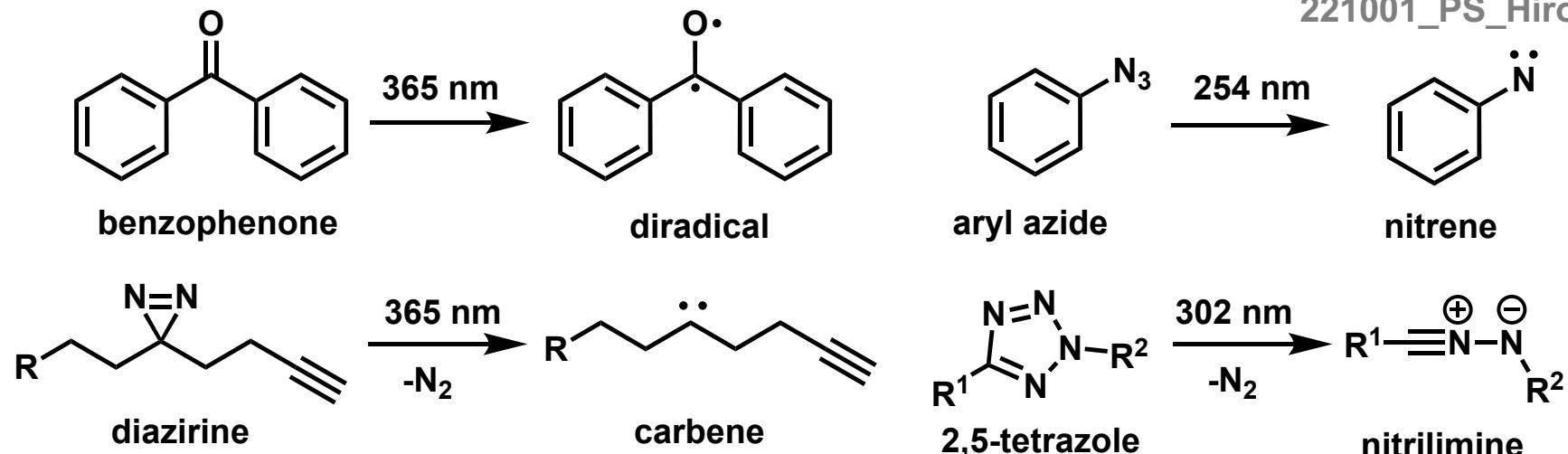
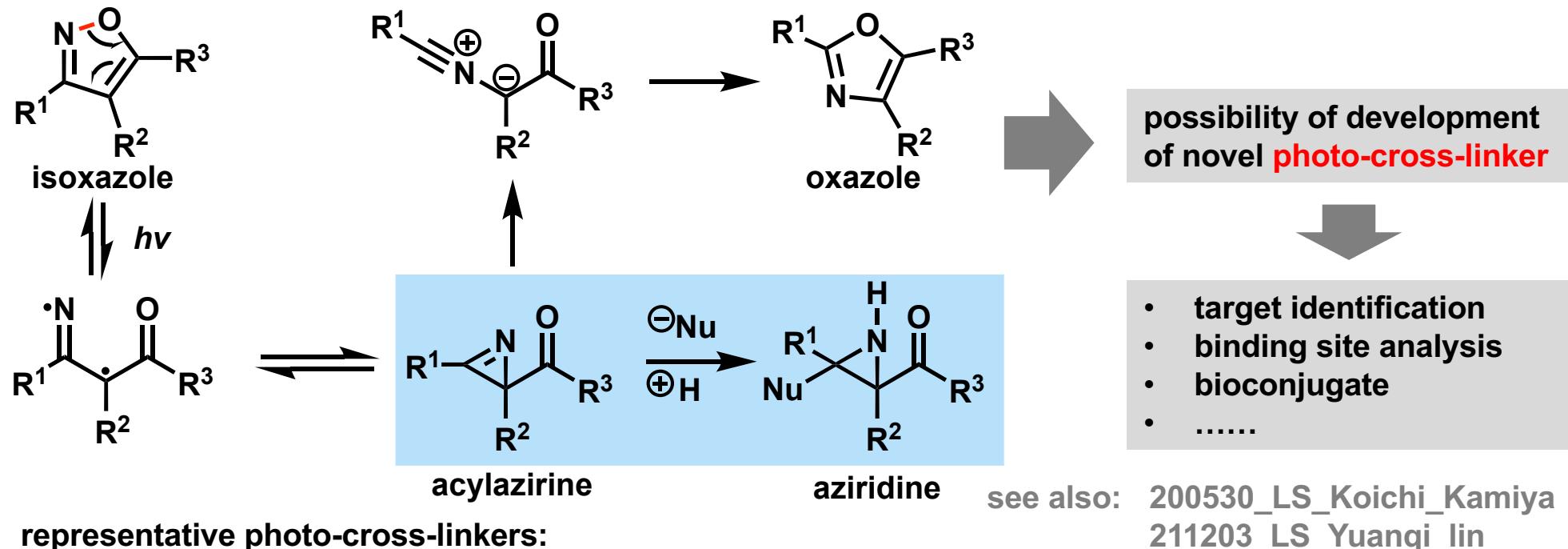
**Dr. Hongyan Sun**

- 2003-2008**    Ph.D. in chemical biology @ National University of Singapore (Prof. Shao Qin Yao)
- 2008-2011**    postdoctoral fellow @ Max Planck Institute of Molecular Physiology, Germany (Prof. Herbert Waldmann)
- 2011-**    associate professor @ City University of Hongkong, department of chemistry

**current research topics:**

- 1. Development of microarray-based screening platform**
- 2. Development of novel fluorescent probes for live cell imaging studies**
- 3. Development of novel biomaterials for biological application**

# Photochemistry of Isoxazole

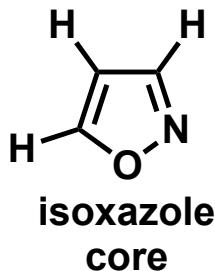


1. Singh *et al.* *J. Am. Chem. Soc.* **1966**, *88*, 1844–1845.
2. Meijler *et al.* *Bioorg. Med. Chem.* **2012**, *20*, 554–570.
3. Brandon *et al.* *ACS Chem. Biol.* **2014**, *9*, 2823–2832.

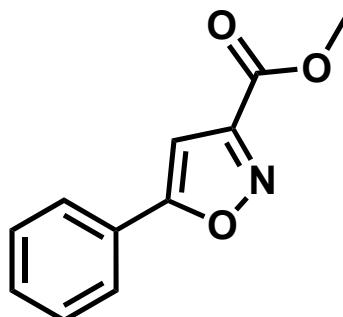
4. Schobert *et al.* *J. Org. Chem.* **2013**, *78*, 2455–2461.
5. Yao *et al.* *Angew. Chem., Int. Ed.* **2013**, *52*, 8551–8556.
6. Yao *et al.* *Angew. Chem., Int. Ed.* **2016**, *55*, 2002–2006.

# Verification of Photo-Reactivity – Model Study (1)

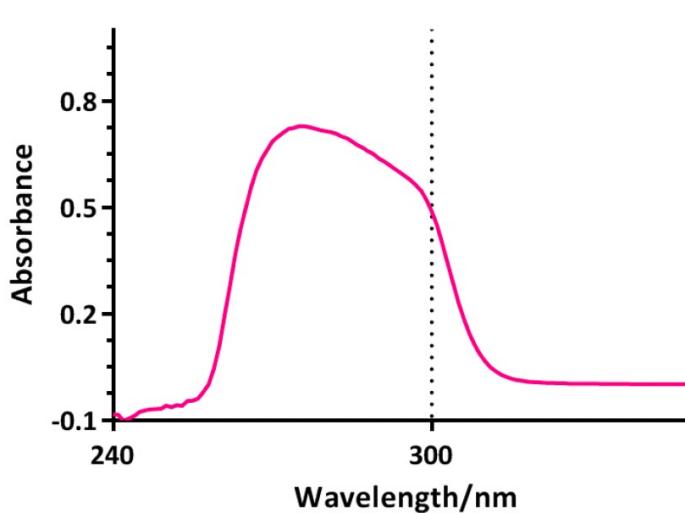
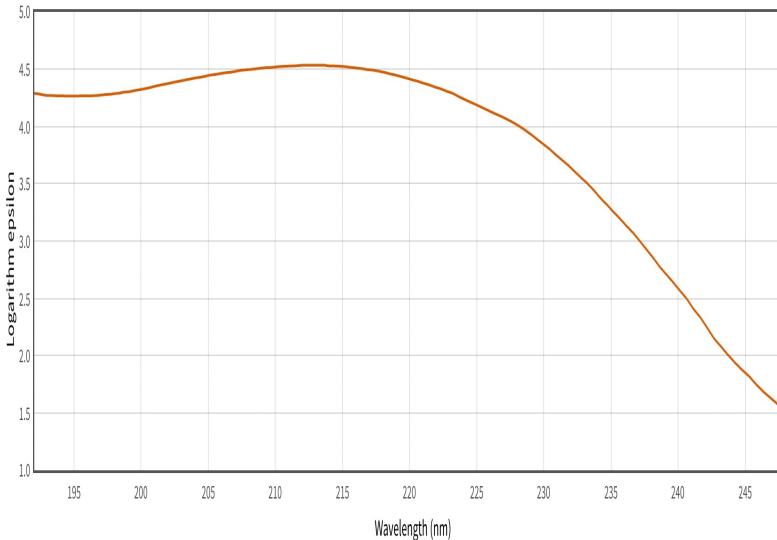
design of model compound:



introduction of  
aryl substitution



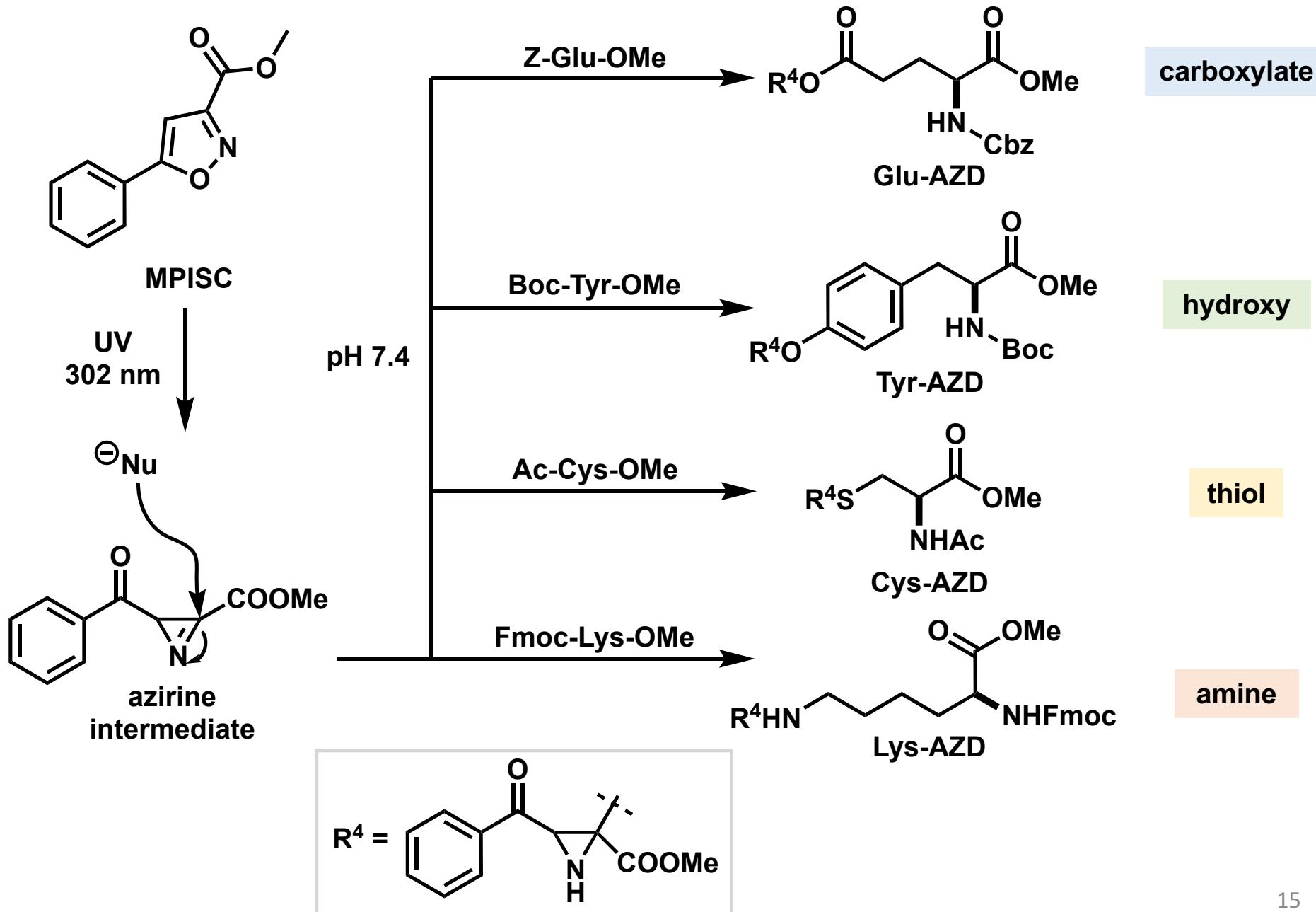
methyl 5-phenylisoxazole-  
3-carboxylate  
(MPISC)



302 nm for the irradiation:  
avoiding damage on cell and  
proteins by short-wave UV

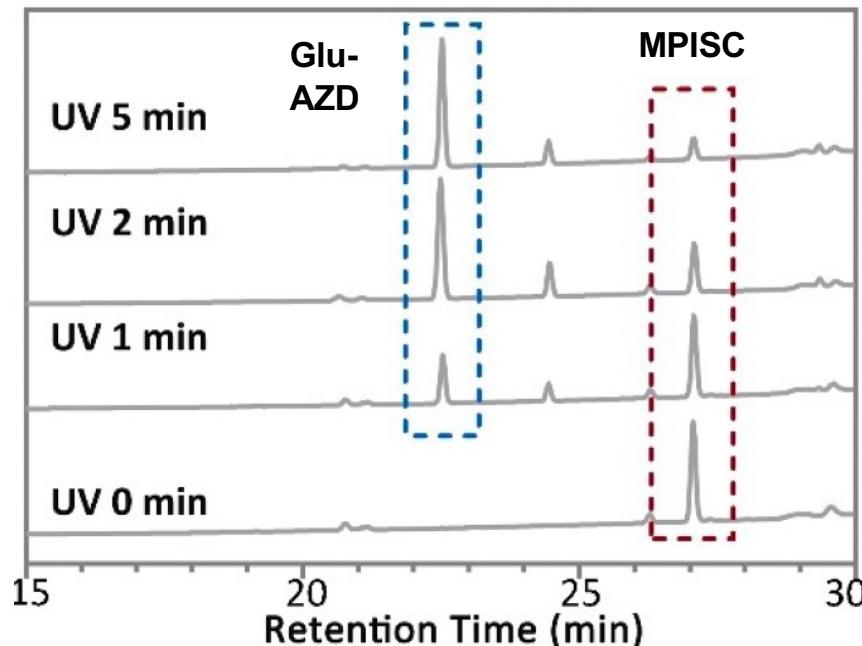
# Verification of Photo-Reactivity – Model Study (2)

photo cross-link with nucleophilic amino acids:



# Verification of Photo-Reactivity – Model Study (3)

for acidic amino acid (Glu):

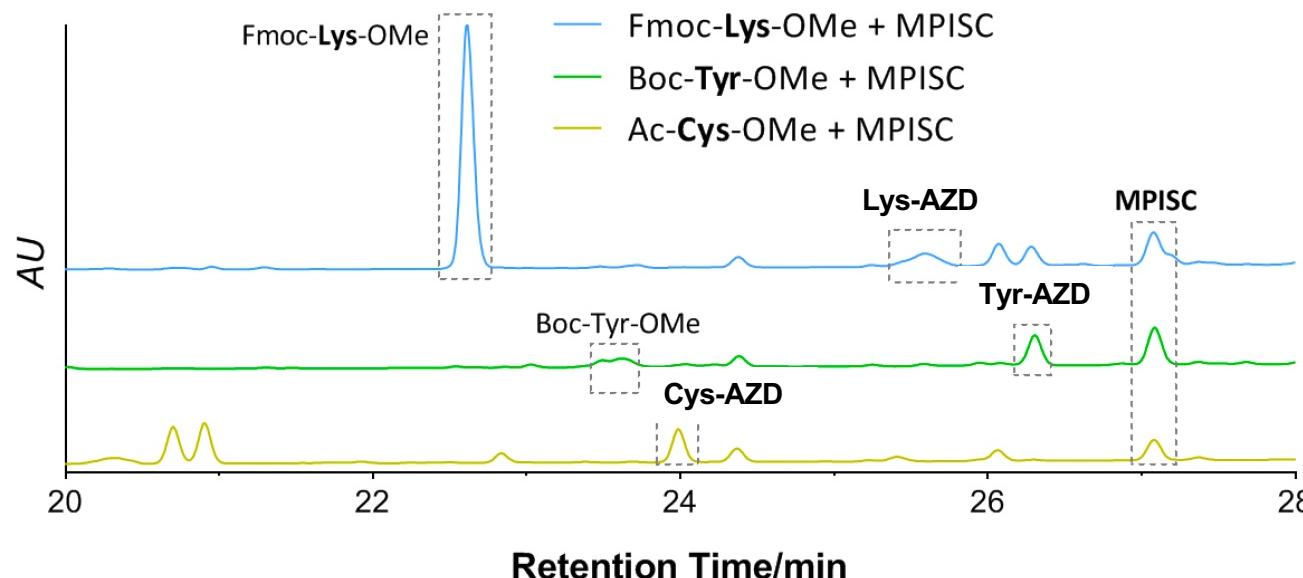


- rapid consumption of MPISC within 5 min
- high yield of Glu-AZD
- no observation of azirine intermediate



preference on **acidic amino acid** at physiological pH

for other nucleophilic amino acids:



- relatively lower yields
- possibly influenced by protonation/deprotonation status at pH7.4

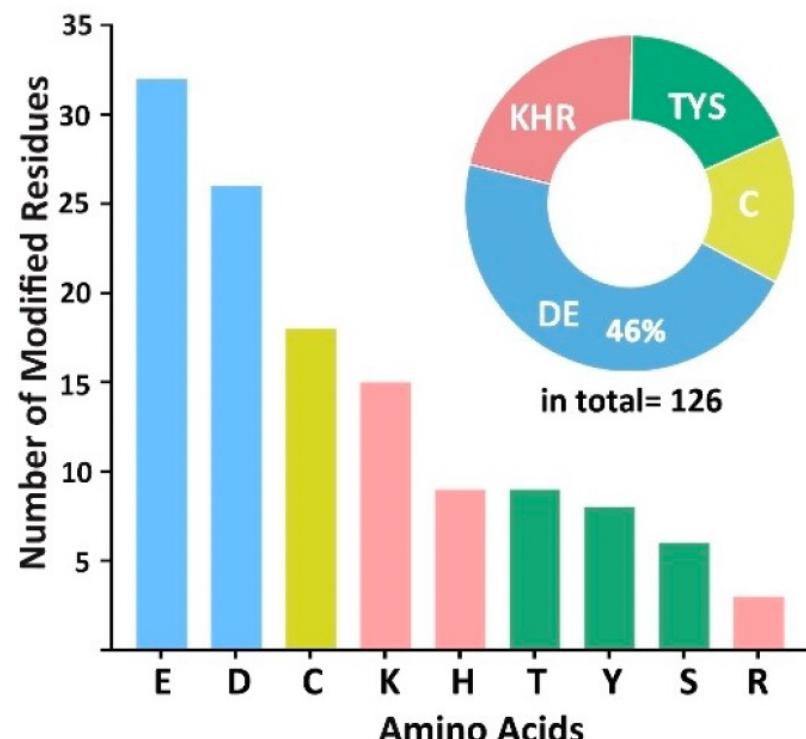
# Verification of Photo-Reactivity – Model Study (4)

photo-cross-link site analysis on bovine serum albumin (BSA):

workflow:



results: 126 cross-link sites(in total )



\* These results only reflected 126 possible labeling sites, instead of a stoichiometry of MPISC:BSA 126:1

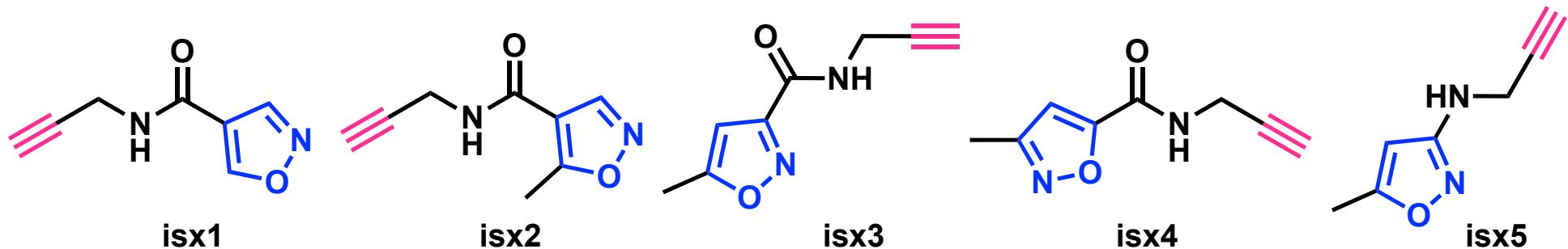
the cross-link preference of MPISC:

- acidic residues: Glu, Asp (main)
- basic residues: Lys, His, Arg
- hydroxylic residues: Thr, Tyr, Ser
- thiol side chain residues: Cys

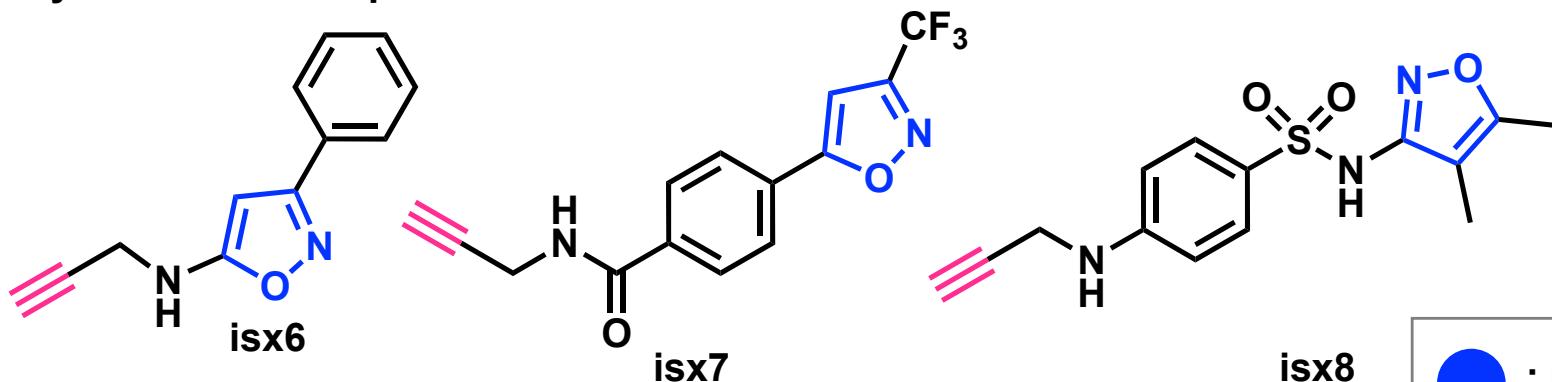
# Functionalization of Isoxazole Photo-Cross-Linker

design and synthesis of isoxazole photoaffinity labeling (PAL) probes:

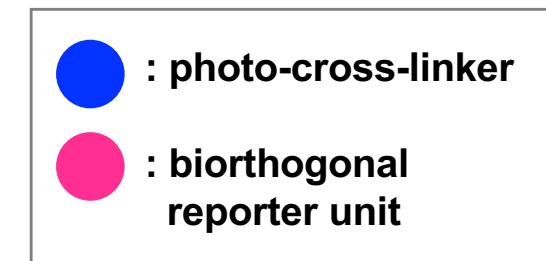
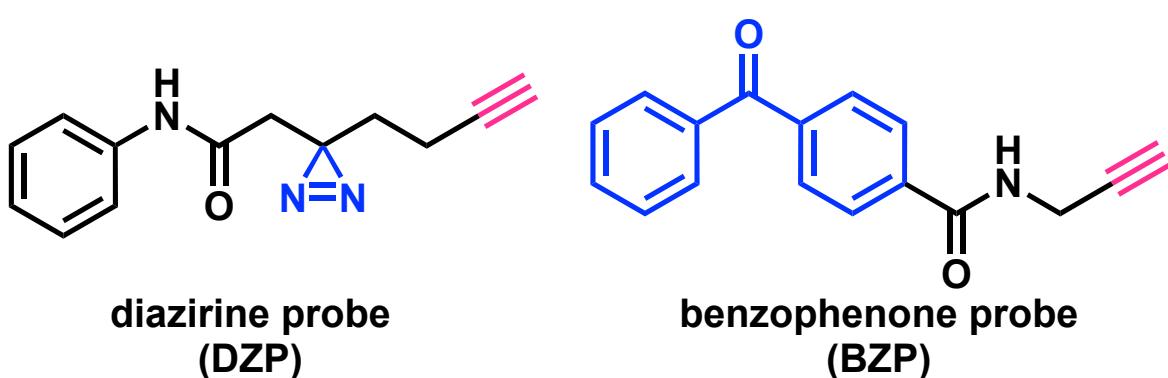
alkyl isoxazole PAL probes:



aryl isoxazole PAL probes:



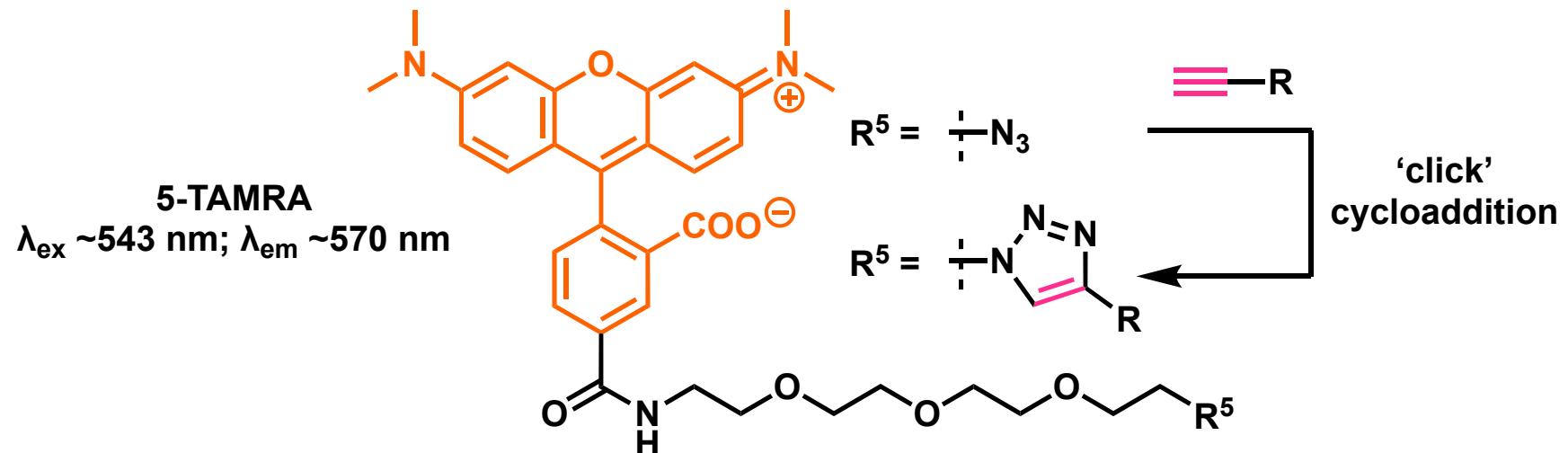
non-isoxazole PAL probes:



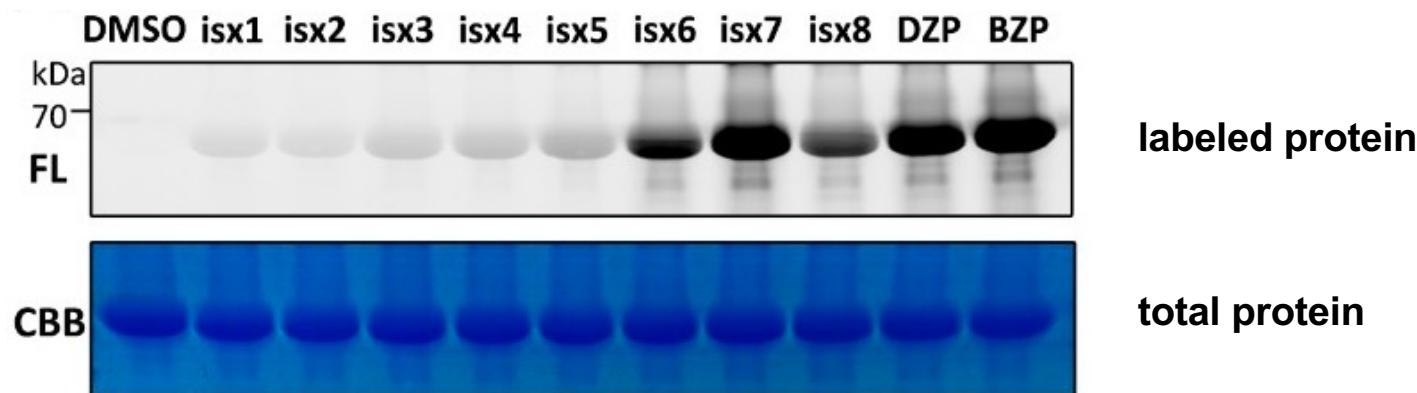
construction could be achieved by condensation or substitution reactions using known compounds

# Photoaffinity Labeling Using Isoxazole Probes (1)

detection of labeled proteins: gel fluorescence



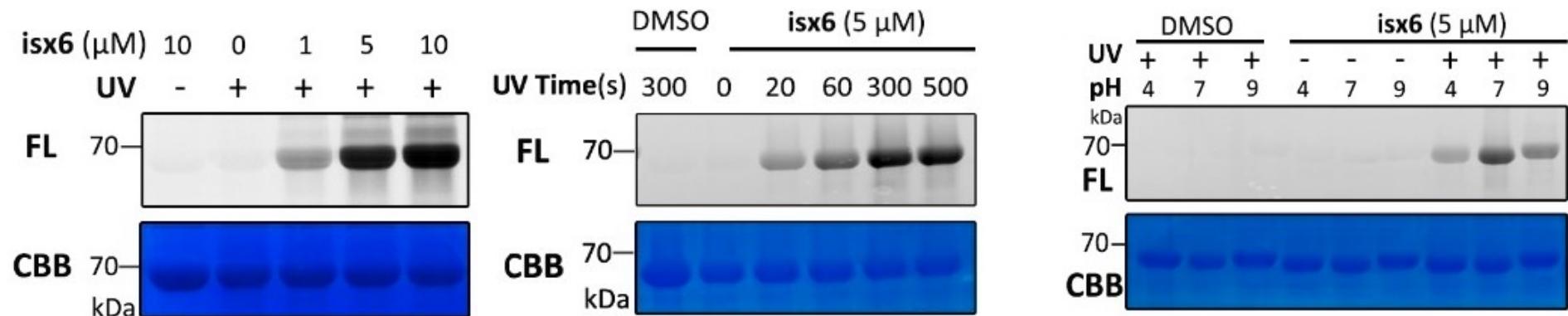
labeling of recombinant BSA:



- alkyl probes (isx1–5) showed moderate labeling, which is a result of its weak absorbance at 302 nm
- aryl probes (isx6–8) showed equipotent labeling efficiency with non-isoxazole PAL probes

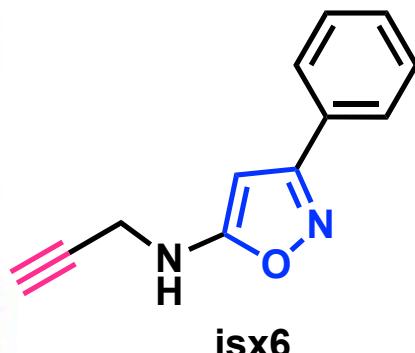
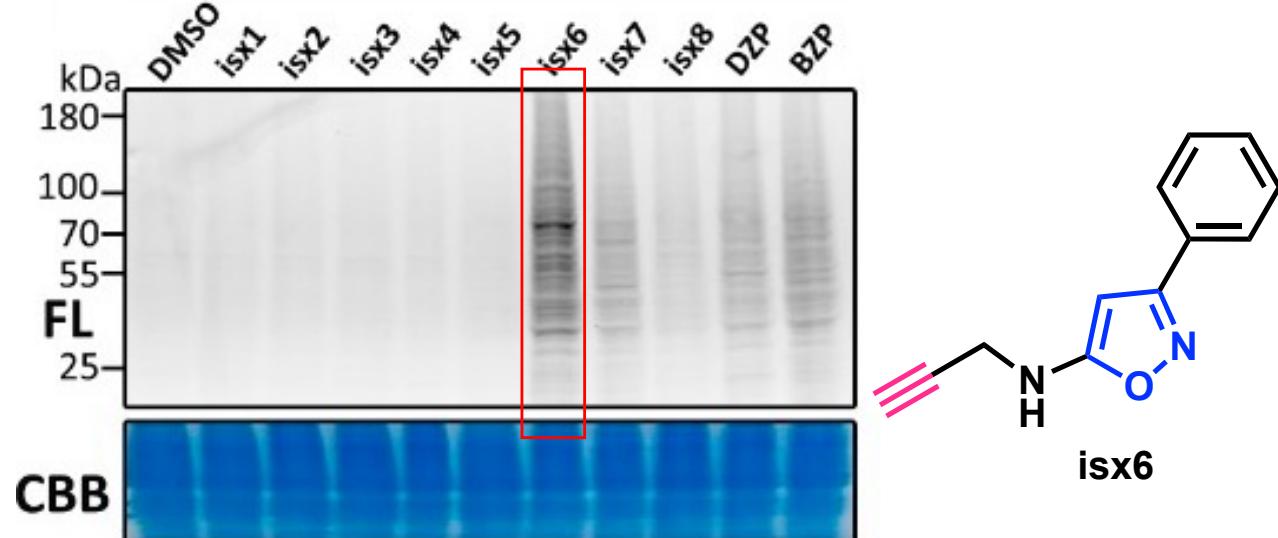
# Photoaffinity Labeling Using Isoxazole Probes (2)

investigation of influence of conditions on labeling efficiency:



- probe concentration-dependent
- quick reaction within 5 min
- pH-dependent: **physiological pH is favored**

in situ photoaffinity labeling of live HeLa cell:

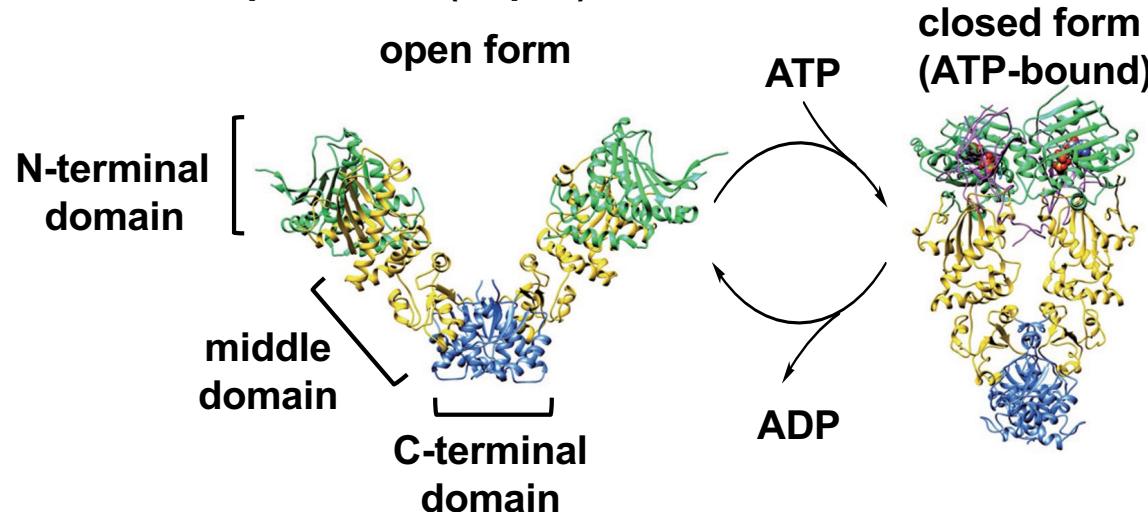


- labeling of **interaction proteins**
- isx6 labeled the most proteins even more than diazirine-type probe (DZP)

potential of isoxazole as  
**effective photo-cross-linker**

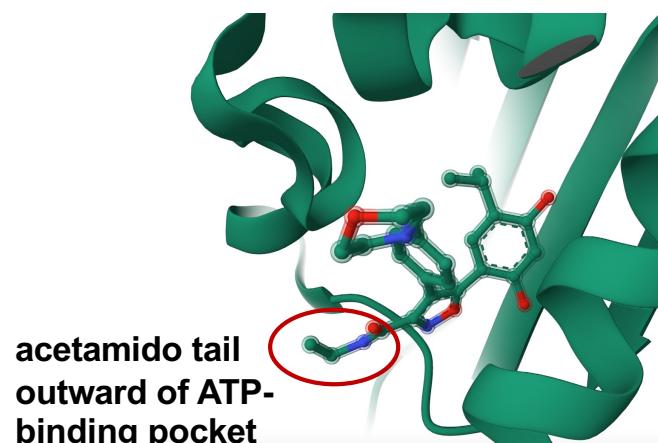
# Heat Shock Protein 90 (Hsp90) and Luminespib

heat shock protein 90 (Hsp90):



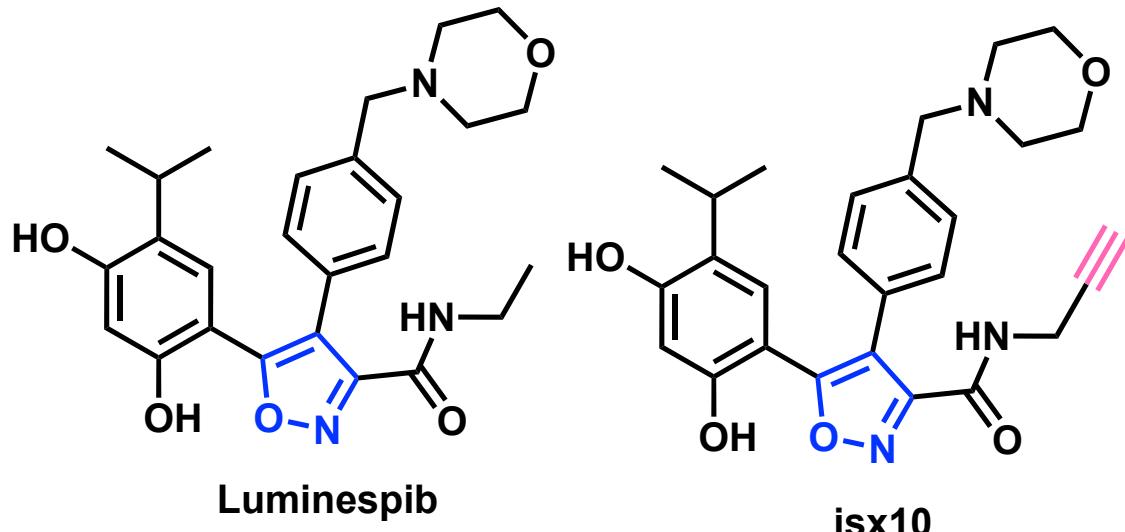
- assistance in protein folding (chaperone function)
- stabilization of proteins against cell **heat stress** (including those for tumor growth)

docking of luminespib with Hsp90:  
(PDB ID: 2VCI)



acetamido tail  
outward of ATP-  
binding pocket

Hsp90 inhibitor luminespib and PAL probe design:



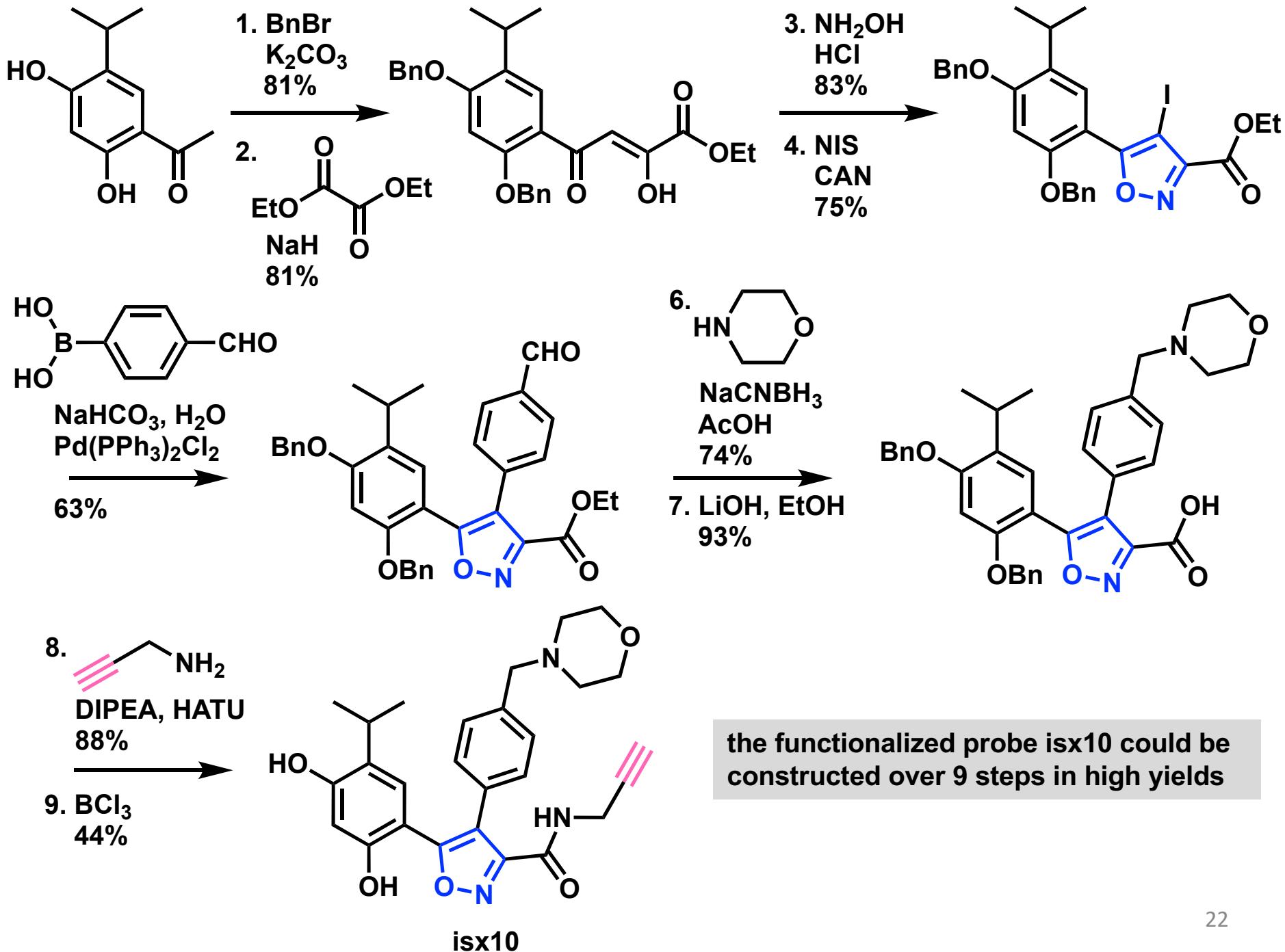
- adverse effects: diarrhea, visual disturbances, and nausea
- necessity of **identification of off-targets**
- photo-cross-linker embedded

minimal structure modification



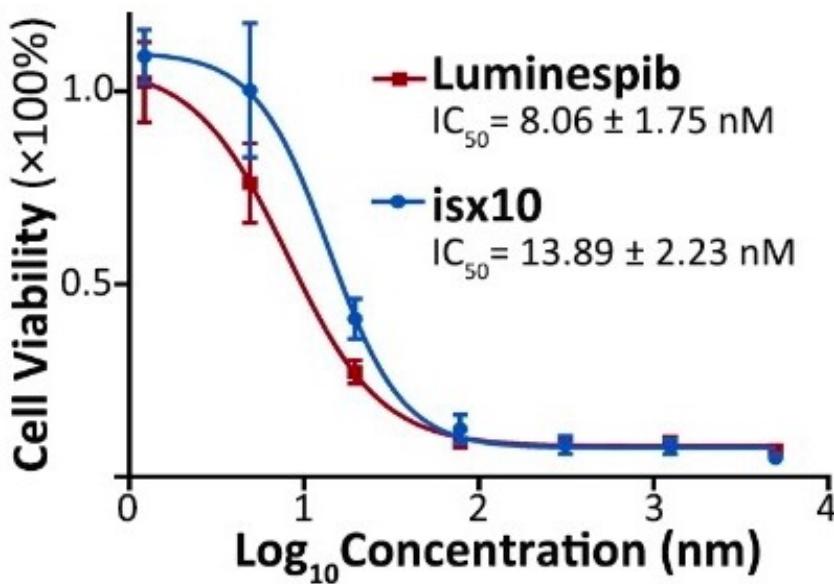
- retaining of biological properties
- profiling of interaction proteins

# Synthesis of isx10

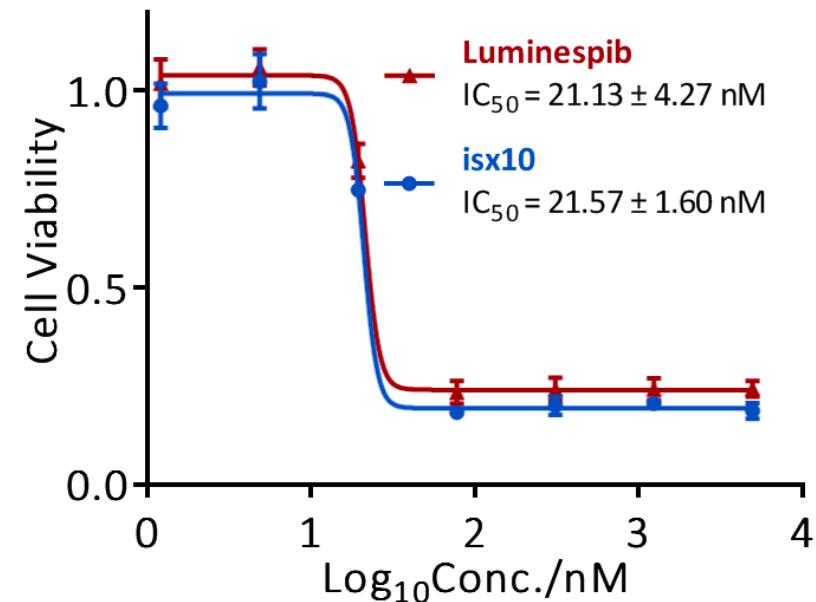


# Bioactivities of isx10

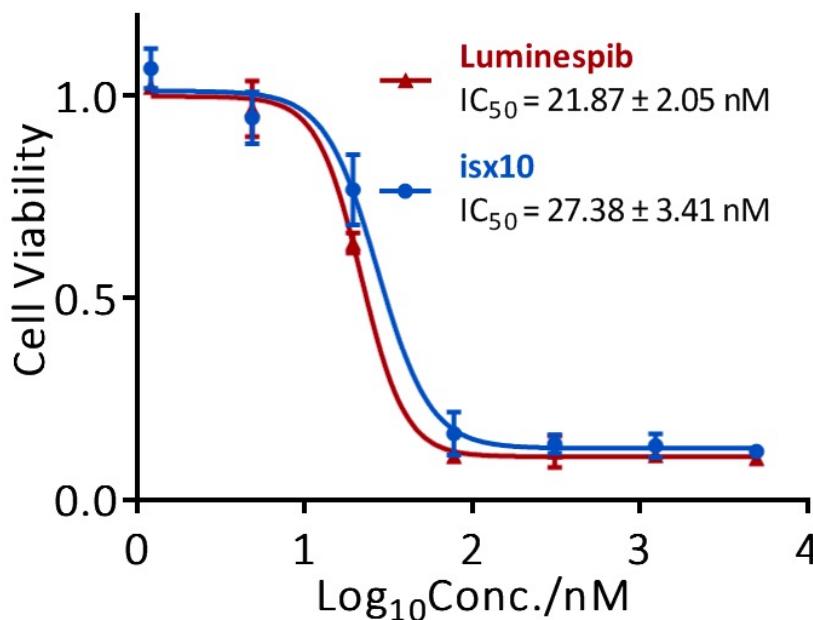
HeLa: cervical cancer cell



HEK293T: human embryonic kidney cell



HCT116: human colon cancer cell



|            | $\text{IC}_{50} (\text{nM})$ |         |        |
|------------|------------------------------|---------|--------|
|            | HeLa                         | HEK293T | HCT116 |
| Luminespib | 8.06                         | 21.1    | 21.9   |
| isx10      | 13.89                        | 21.6    | 27.4   |

- equipotent cytotoxicity
- similar cell-line selectivity

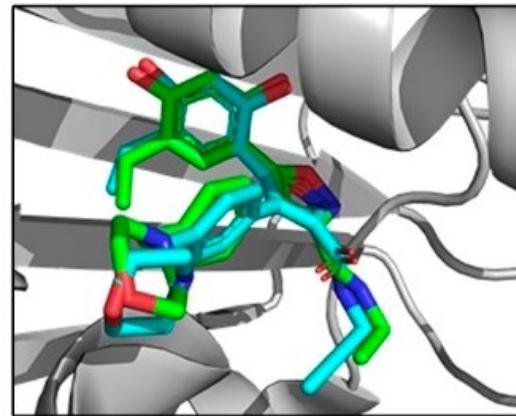


preservation of bioactivities

# Interaction of isx10 with Hsp90

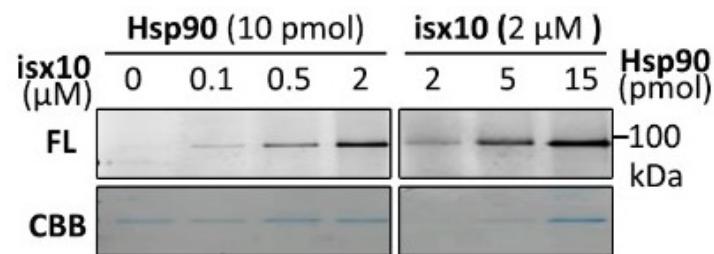
docking analysis of compounds with Hsp90:

- similar atomic coordinates
- similar Hsp90-binding patterns



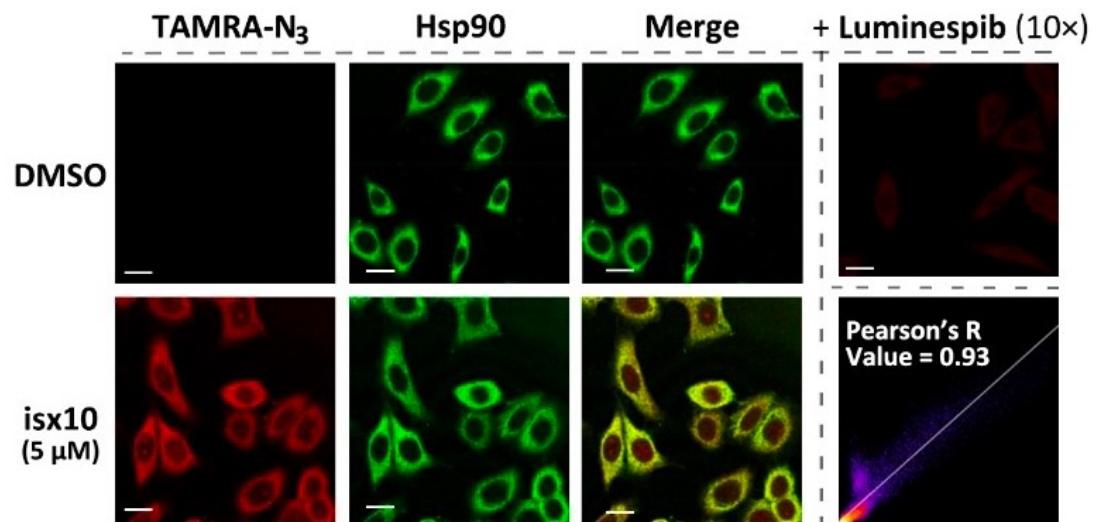
- : Hsp90
- : Luminespib
- : isx10

photoaffinity labeling of Hsp90 with isx10:  
using recombinant Hsp90:



- sensitive labeling of recombinant Hsp90 even at low concentrations
- good efficiency and specificity

in live HeLa cell:



- : fluorescence of TAMRA

- : fluorescence of Alexa Fluor 488 conjugated on secondary antibody

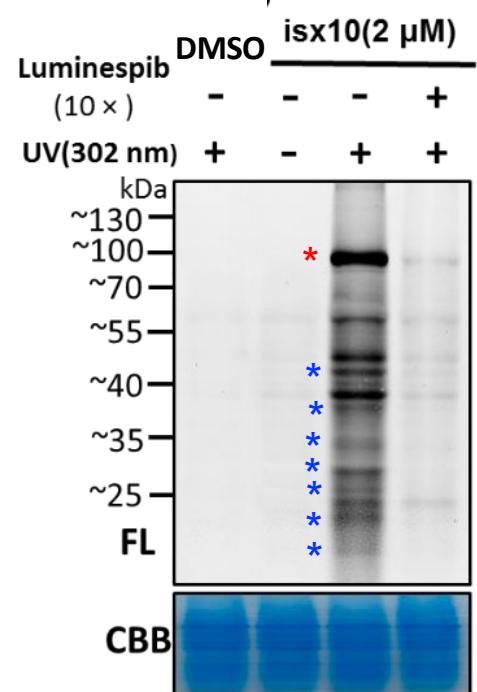
functionalization of luminespib didn't affect the molecular interaction with Hsp90

# Photoaffinity Labeling of Interaction Proteins

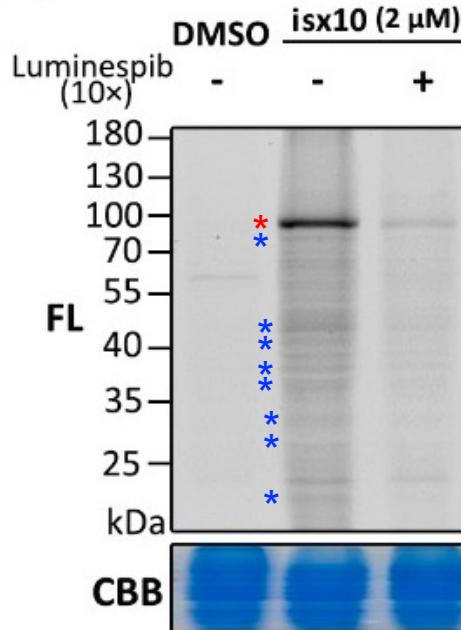
competitive photoaffinity labeling: exclusion of non-specific interactions

parent luminespib used as competitor

using HeLa cell lysate:



in live HeLa cell:



\*: Hsp90

\*: specifically labeled proteins

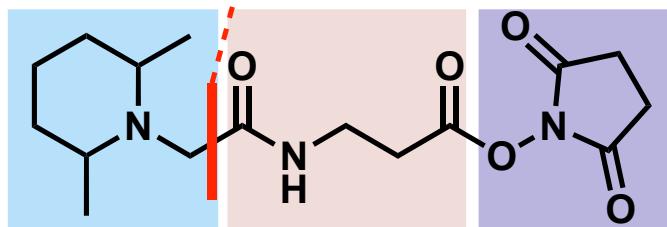
- several unknown proteins specifically labeled
- existence of possible interaction proteins confirmed



identification of possible interaction proteins –  
tandem mass tag (TMT)-based chemoproteomic experiments

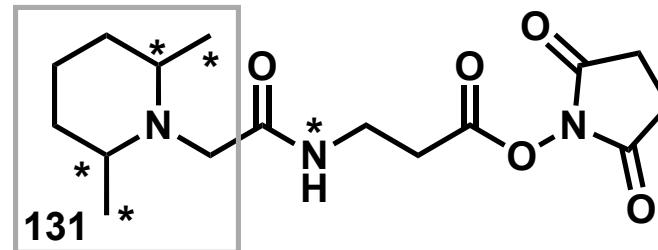
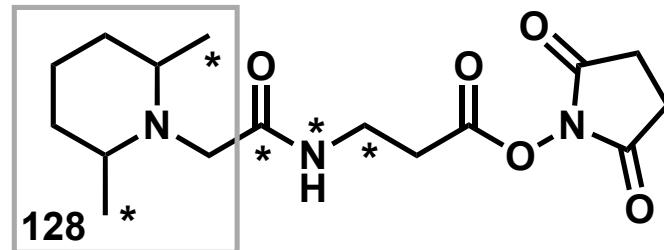
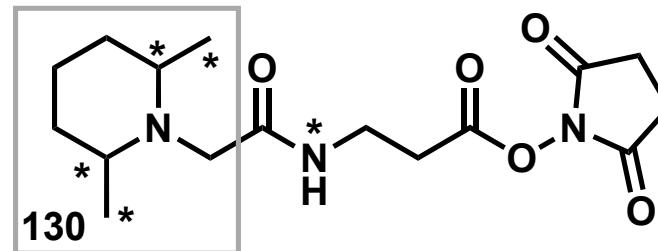
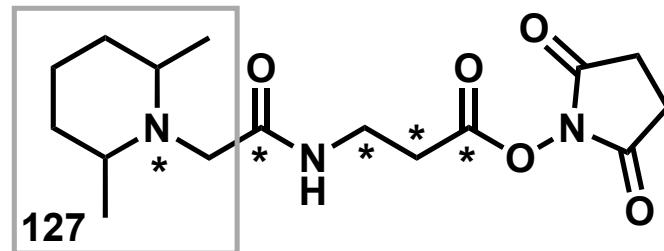
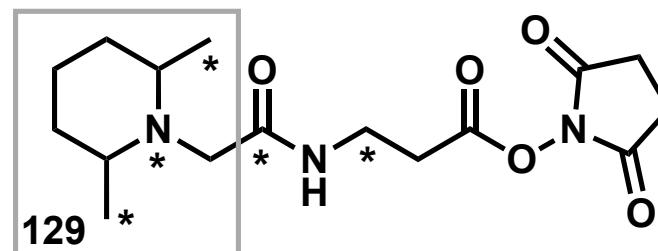
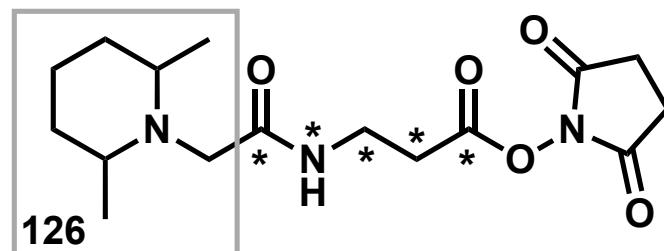
# TMT-based Peptide Quantification (1)

cleavable by higher energy collisional dissociation (HCD)



mass reporter    mass normalizer    NH<sub>2</sub>-reactive group

\*: heavy atom

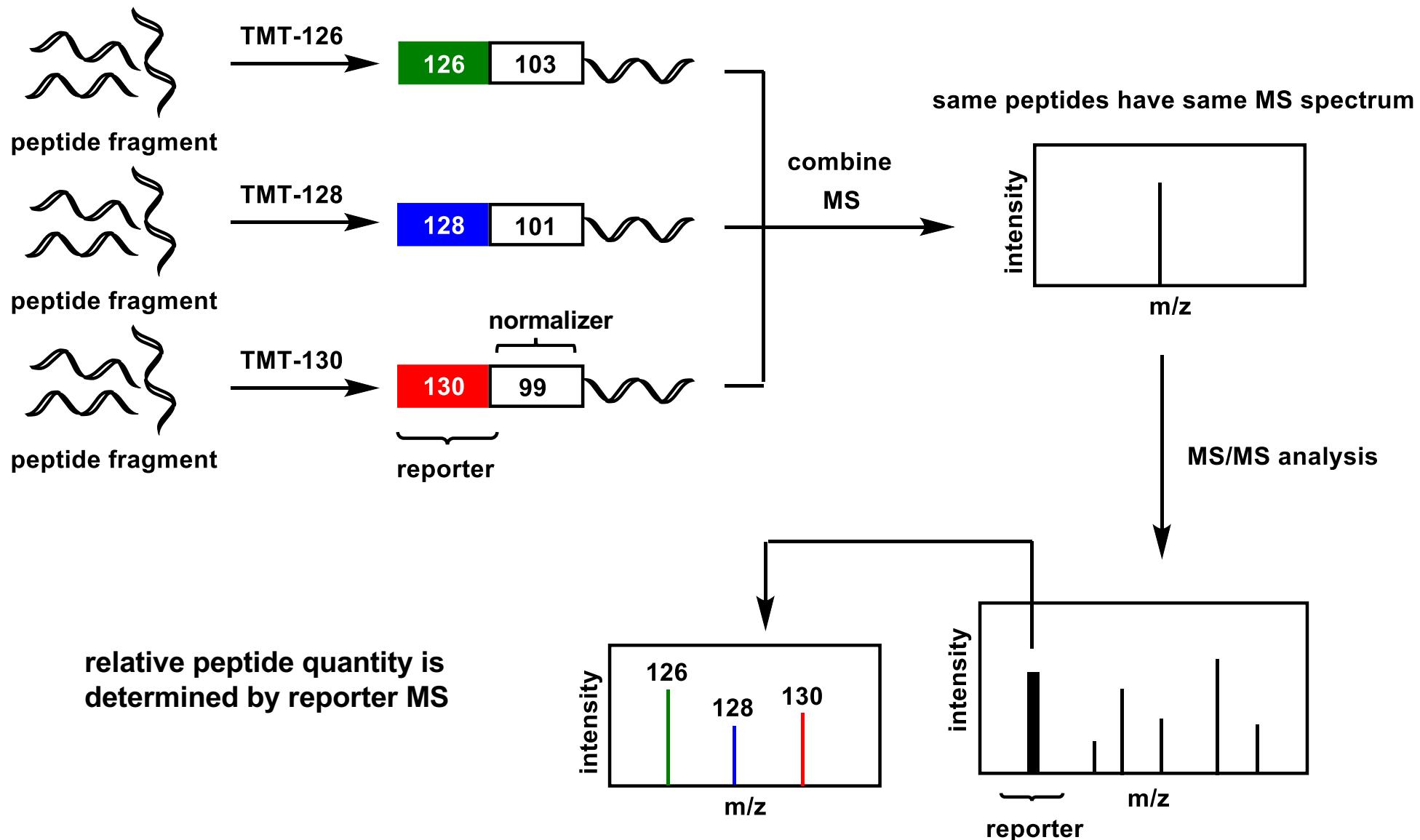


peptides can be tagged through nucleophilic addition of N-terminal NH<sub>2</sub> or Lys side chain NH<sub>2</sub> to TMT

1. Dayon, L.; Hainard, A.; Licker, V.; Turck, N.; Kuhn, K.; Hochstrasser, D. F.; Burkard, P. R.; Sanchez, J. *Anal. Chem.* **2008**, *80*, 1895.
2. see also 200530\_LS\_Koichi\_Kamiya

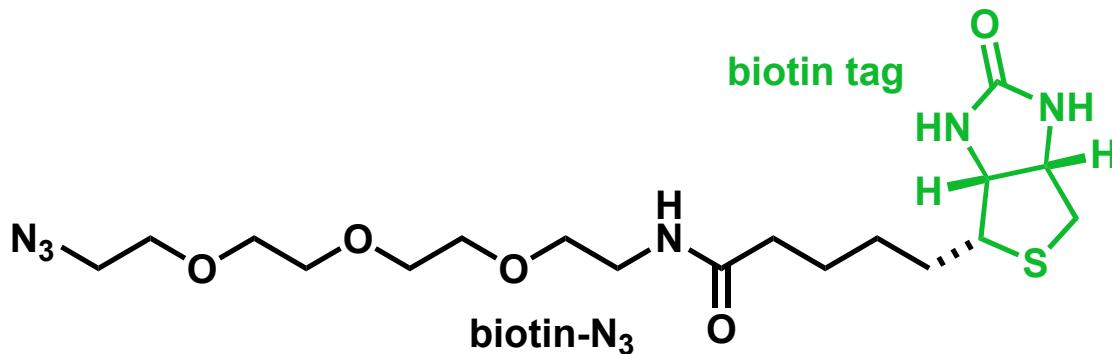
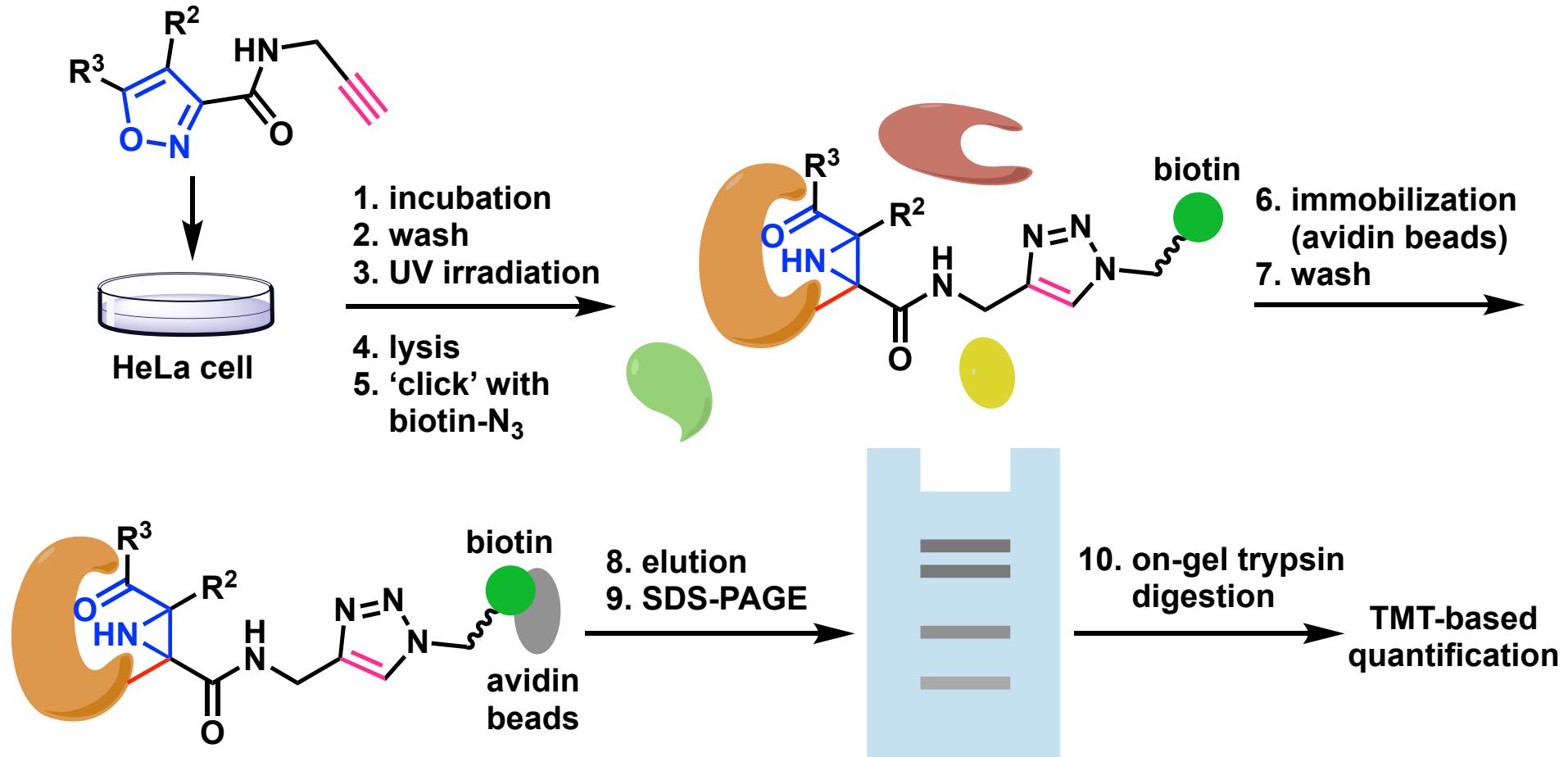
# TMT-based Peptide Quantification (2)

a total  $m/z$  number of 229 to keep the mass of TMT constant



- Thompson, A.; Schäfer, J.; Kuhn, K.; Kienle, S.; Schwarz, J.; Schmidt, G.; Neumann, T.; Hamon, C. *Anal. Chem.* **2003**, 75, 4942–4942.

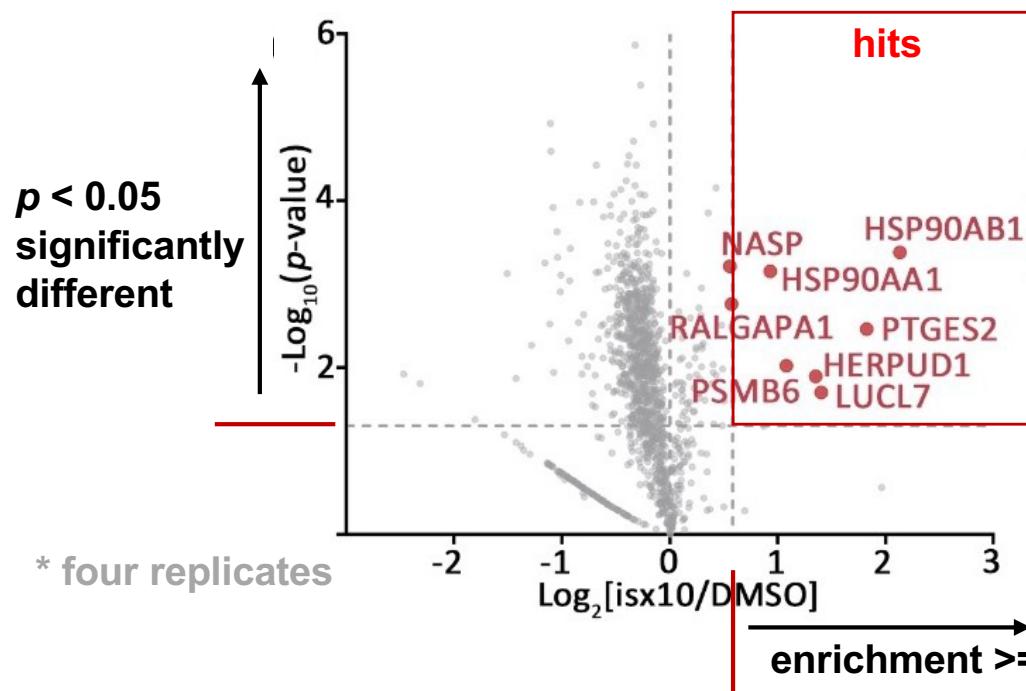
# PAL Pull-Down — Enrichment of Interaction Proteins



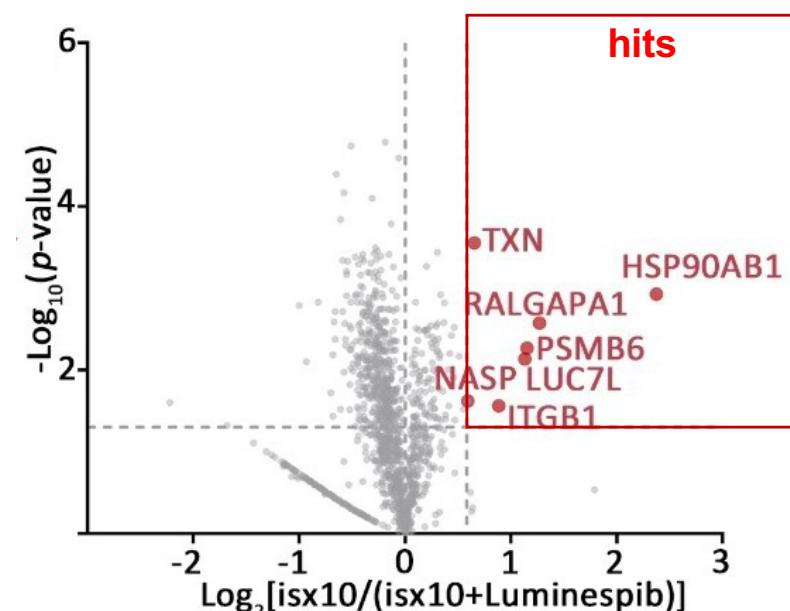
immobilization of labeled proteins:  
strong biotin-avidin interaction  
( $K_D \sim 10^3$  pM)

# Identification of Interaction Proteins

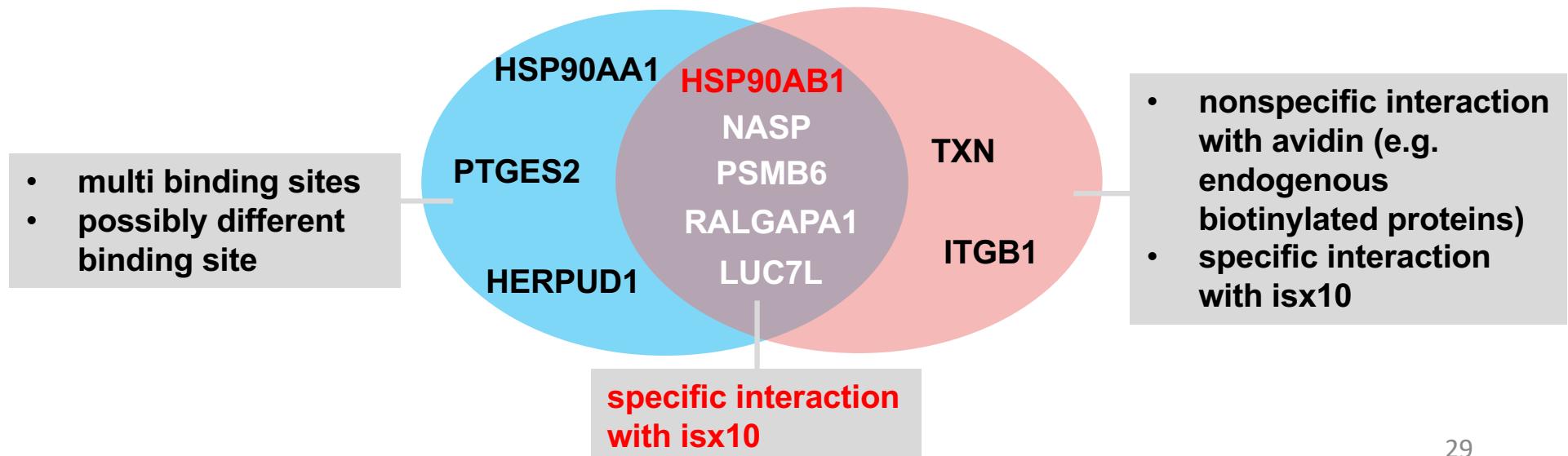
normal:



competitive:

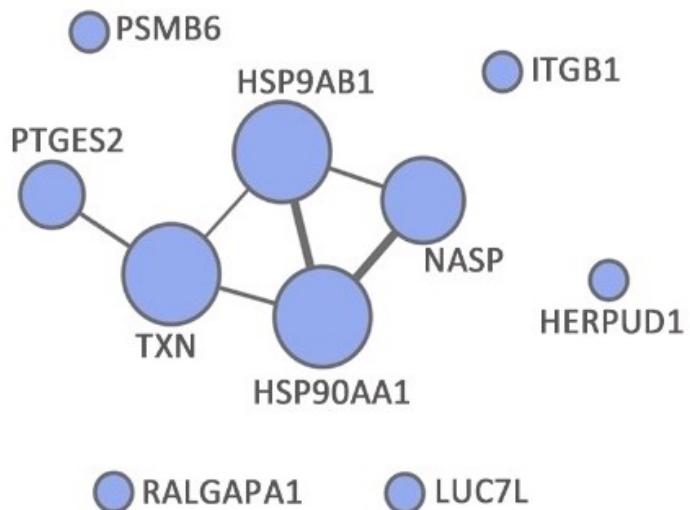


10 enriched proteins:



# Investigation on Enriched Proteins

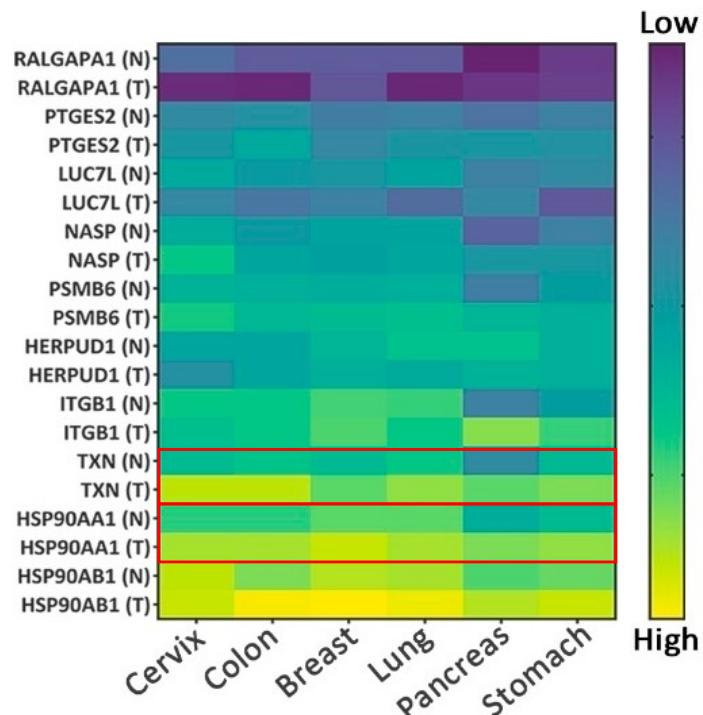
protein-protein interactions (PPIs):



- HSP90AB1: Hsp90 $\beta$  isoform  
HSP90AA1: Hsp90 $\alpha$  isoform  
NASP: nuclear autoantigenic sperm protein  
(histone chaperone and Hsp90 binding protein)  
TXN: thioredoxin: reductive Cys residue  
(anti oxidative stress)  
PTGES2: prostaglandin E2 synthase  
(potential COVID-90 therapeutic target)

three interaction proteins in Hsp90 PPIs network:  
possible involvement in anti-cancer activity

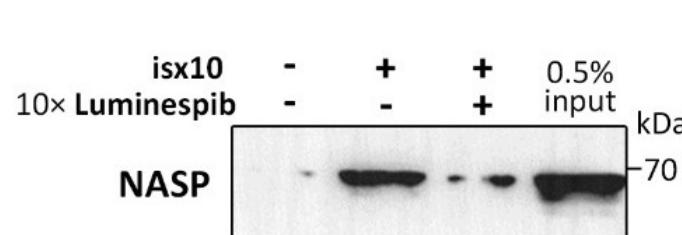
gene expression in normal and tumor tissues:



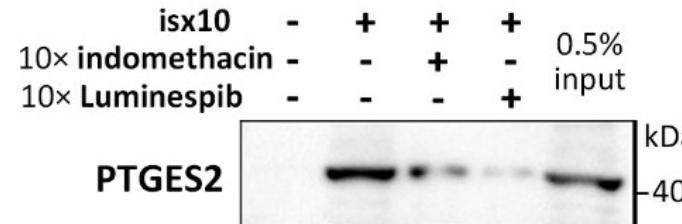
- Hsp90 and TXN showed higher expression in cancer tissues — cancer therapeutic specificity
- other interaction proteins have minor changes in expression level — potential toxicity

# Validation of Protein Binding

competitive PAL pull-down and Western blotting:



indomethacin: known inhibitor of PTGES2

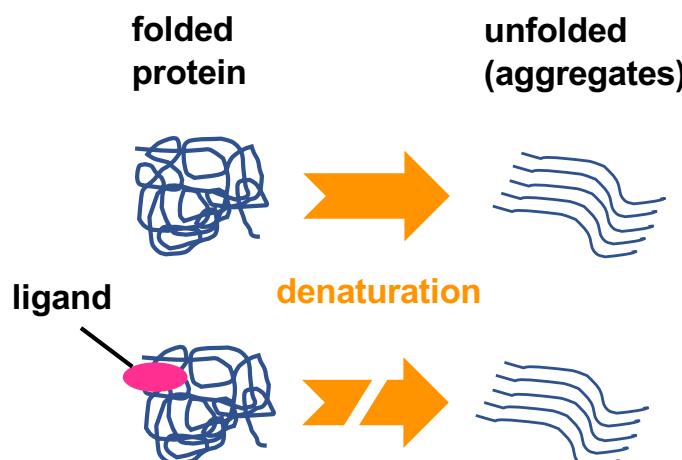


chemiluminescence from  
enzyme activity of antibody-  
conjugated HRP

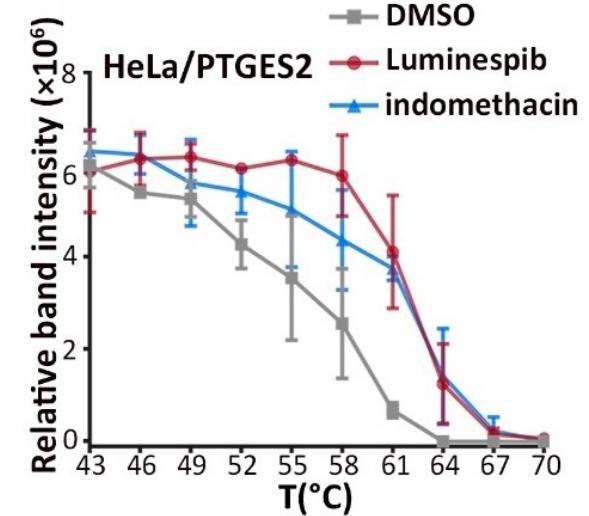
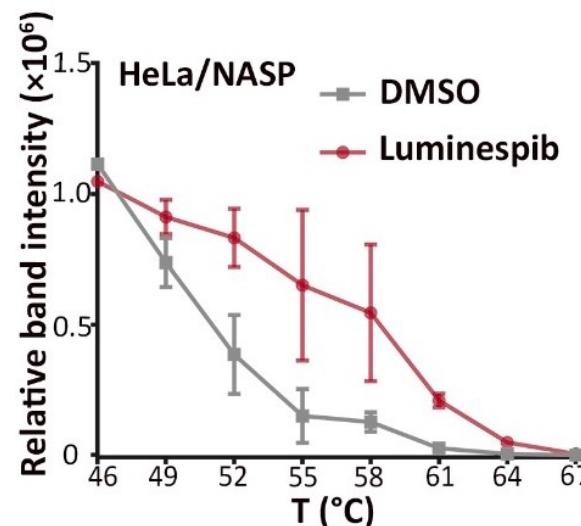
**luminespib specifically interacts with NASP and PTGES2**

cellular thermal shift assay (CETSA):

influence of binding complex formation on thermostability



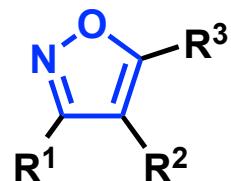
melting curve: plot of temperature to soluble folded protein



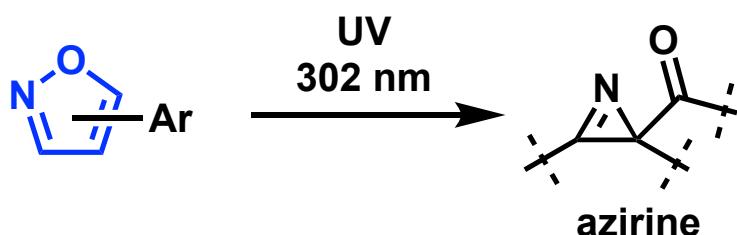
**remarkable stabilization of luminespib on NASP and PTGES2 validated the binding affinity**

# Summary

isoxazole: significant skeleton for bioactive compounds



- **pharmacophoric features**
- easily synthesized
- important synthetic intermediate



- efficient labeling of amino acids and proteins
- application of luminespib-derived probe in chemoproteomics

capitalize on  
photochemistry features

future  
perspectives

novel **photo-cross-linker** (embedded/extracellular):

- **chemoproteomics**: identification of interaction proteins
- bioconjugate: cross-link with protein/antibody
- covalent inhibition: photo therapeutics