



MicroRNA Therapeutics

-Overview and Cutting Edge of Small Molecule-based Approach-

Literature Seminar on 13th, May, 2017

D3 Kai Kitamura

Outline

- Introduction of miRNA

- Overview of miRNA therapeutics

- Main topic:

Rational Design of Small Molecule Targeting Noncoding RNAs

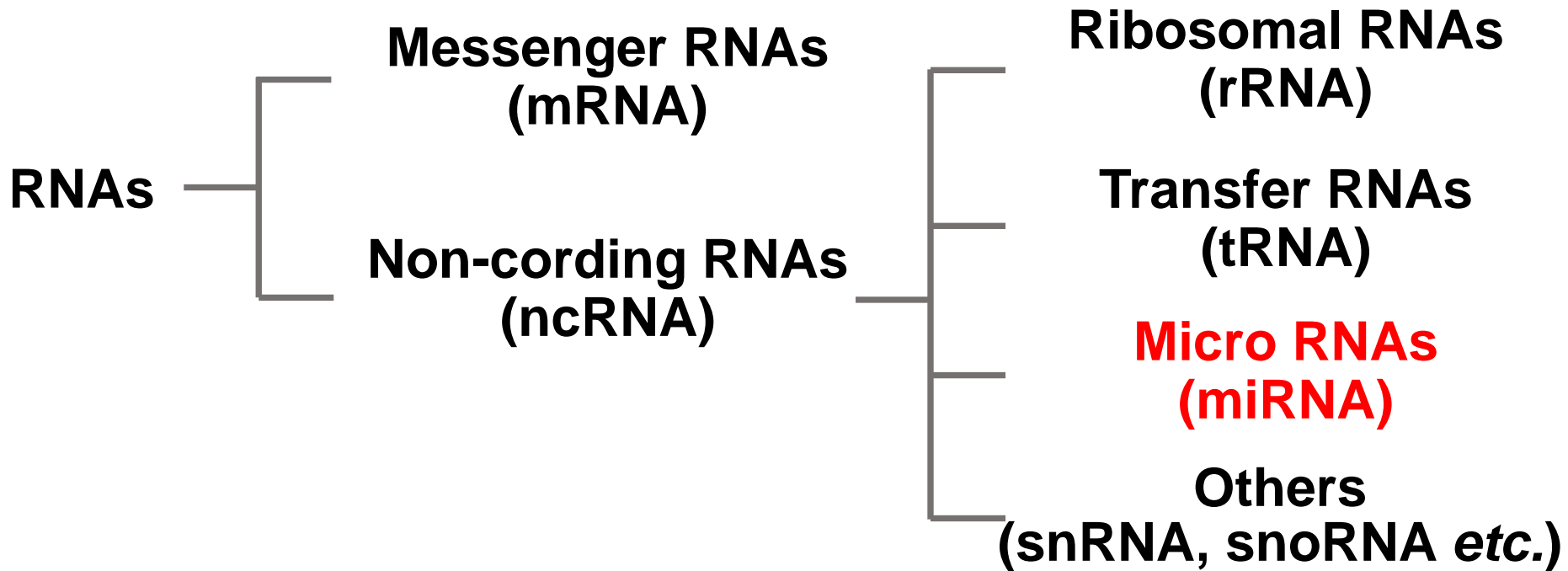
(Velagapudi, S. P. *et al. ACS Cent. Sci.* **2017**, 3, 205)

- Subtopic:

Regulating miRNA by Bifunctional Small Molecule

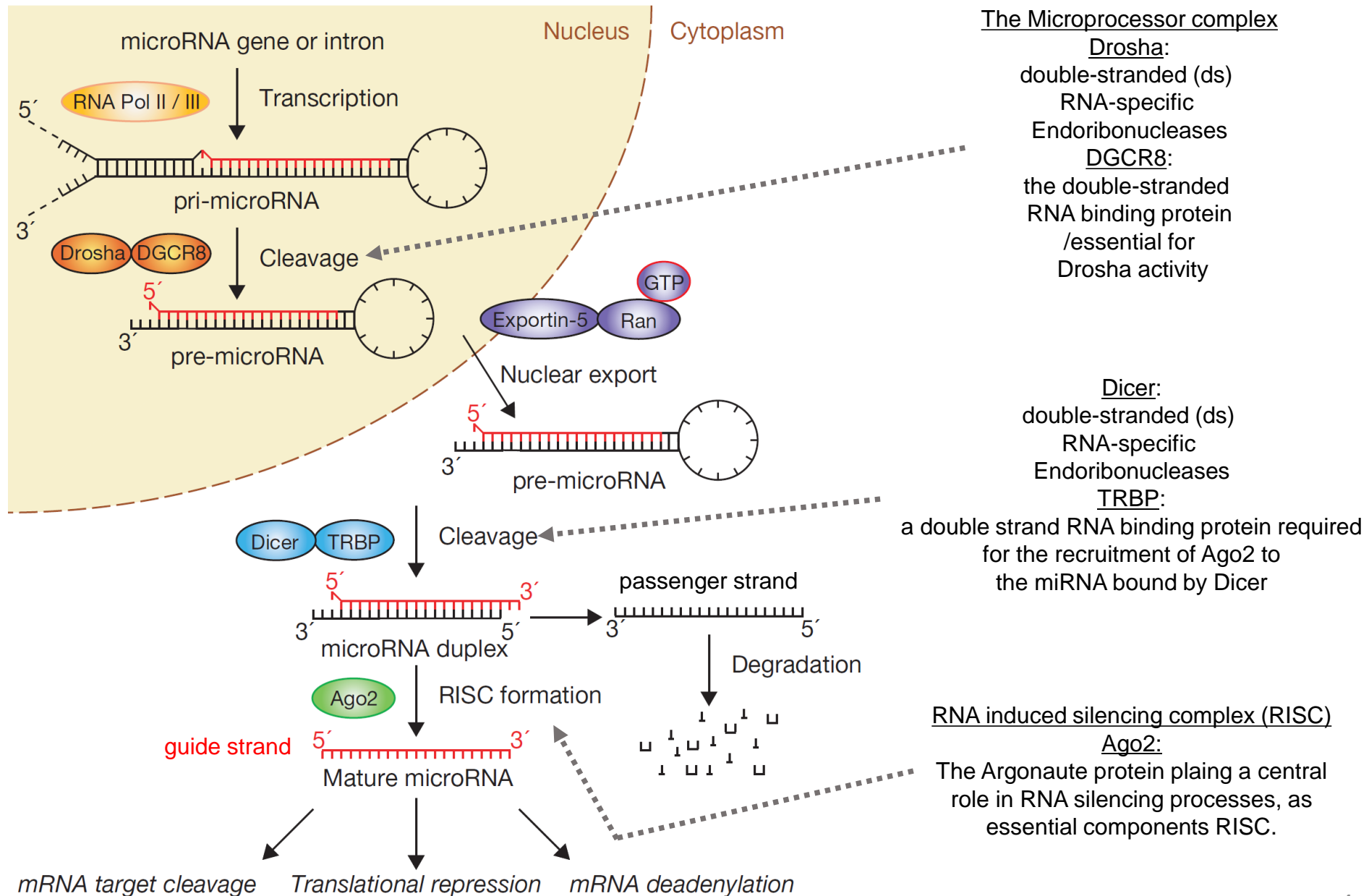
(Yan, H. *et al. J. Am. Chem. Soc.* **2017**, 139, 4987)

MicroRNAs (miRNAs)



- ✓ *Small non-coding RNAs consist of 21-25 nucleotides*
- ✓ *Encoded in the genomes of plants and animals, and highly conserved*
- ✓ *Regulating the expression of genes by binding to mainly the 3'-untranslated regions (3'-UTR) of specific mRNAs*

Biogenesis of miRNAs



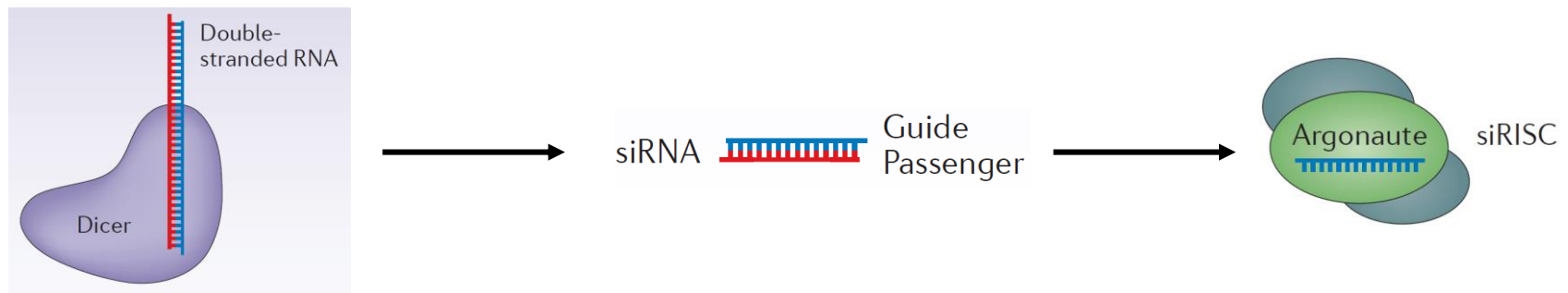
Difference between miRNAs and siRNAs

Similarities

- ✓ *Processed by Dicer*
- ✓ *Formation of RISC (RNA-induced silencing complex)*
- ✓ *degrade mRNAs bearing fully complementary sequences*

Differences

- ✓ *miRNAs are endogenously encoded small noncoding RNAs, derived by processing of short RNA hairpins*
- ✓ *siRNAs are derived by processing of long double-stranded RNAs and are often of exogenous origin*
- ✓ *miRNA can inhibit the translation of mRNAs bearing partially complementary target sequences*



The role of miRNA in diseases

■ The role of miRNAs in cancer

- ✓ Dysregulation of miRNA biogenesis enzymes
(mutations, transcriptional changes *etc.* in Drosha, Dicer, *etc.*)
- ✓ Dysregulation of tumor-suppressive miRNAs
(miR-34 family, let-7 family, miR-200 family, miR-15/16 *etc.*)
- ✓ Dysregulation of miRNAs with oncogenic function
(miR-21 = anti-apoptotic role, miR-155, miR-210, *etc.*)

■ Other diseases relevant to miRNAs

- ✓ Hepatitis C infection (miR-122: upregulating the replication)
- ✓ Cardiovascular disease (miR-21, miR143/145, miR-1 *etc.*)
- ✓ Atherosclerosis (miR-33: downregulating)
- ✓ Diabetes (miR-200, miR192, miR-29 family)
- ✓ Scleroderma (miR-29)

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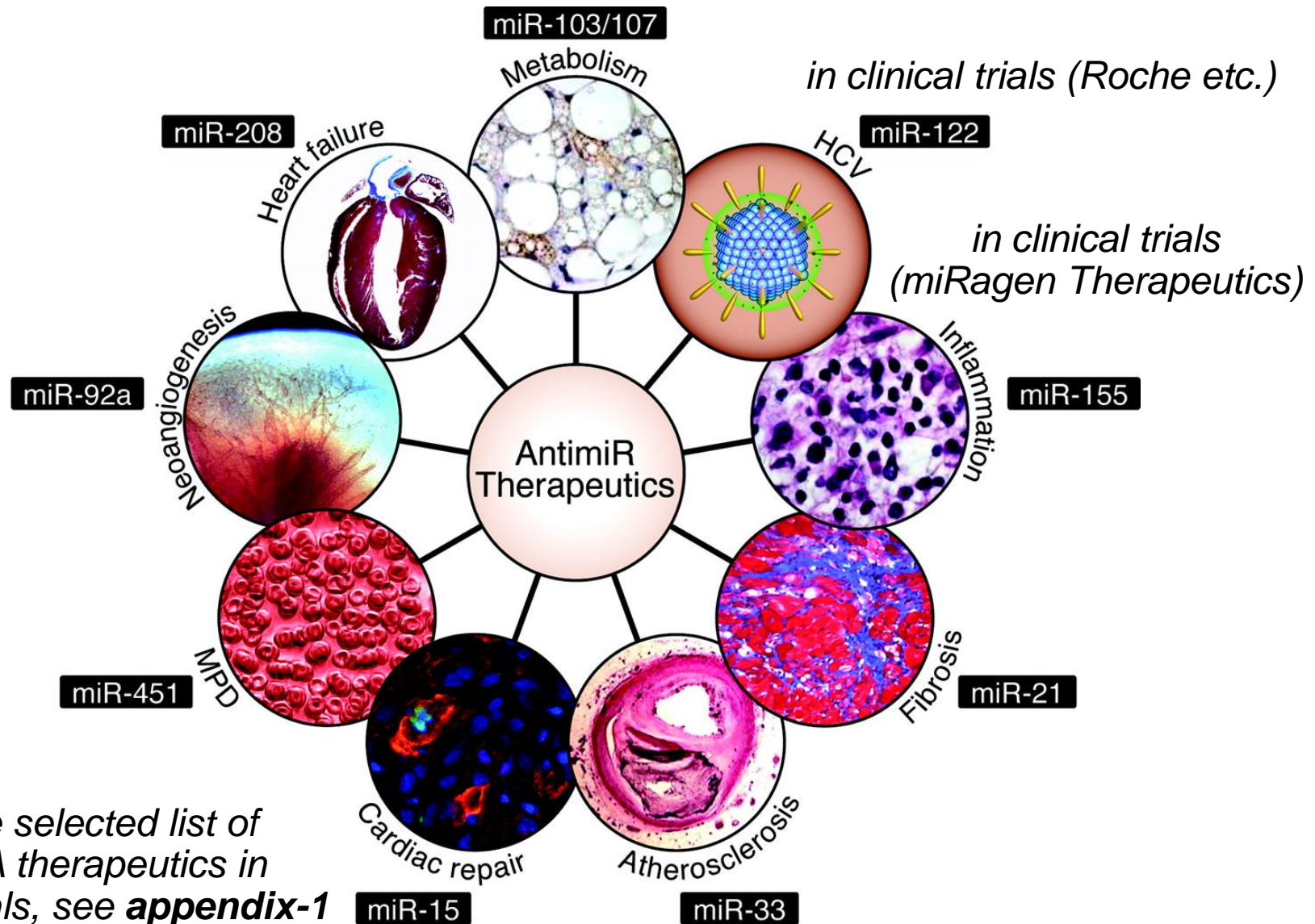
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(Yan, H. *et al. J. Am. Chem. Soc.* **2017**, 139, 4987)

miRNA therapeutics

in clinical trials (Regulus Therapeutics)



Strategies in miRNA therapeutics

■ Micro RNA therapeutics:

1. Compensation for miRNA dysfunction
 2. Inhibition of miRNA activity
- RNAs themselves seem to be promising drug candidates

Problems in using RNAs as drug candidates

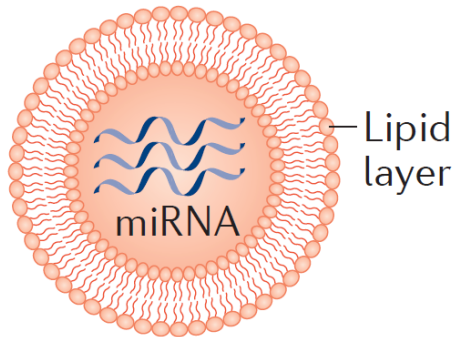
- ✓ Instability in bloodstream
- ✓ Poor delivery to the target site (poor cell permeability)

Strategies

- ✓ Delivery vehicle to encapsulate RNAs
- ✓ Chemical modifications to the nucleotide backbone
- ✓ Small molecules to control biogenesis of RNAs (Topic)

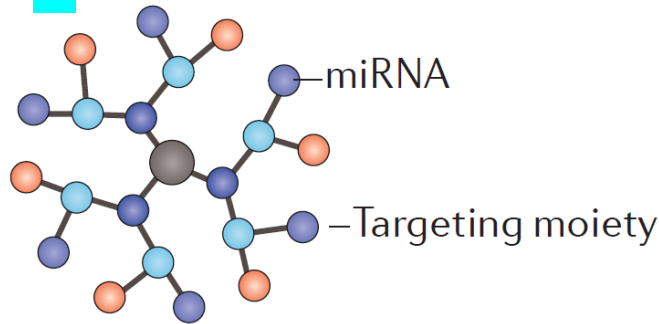
RNA delivery technologies

A



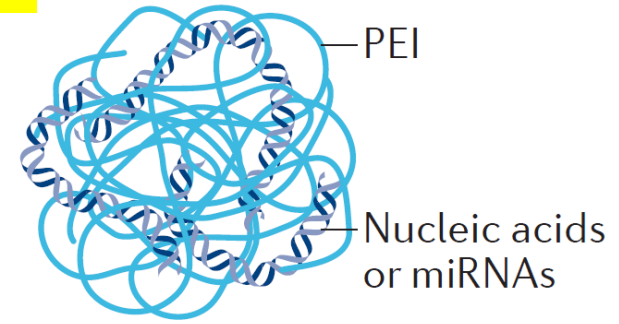
Liposomes (DOPC or NLE)

B



Dendrimers

C



PEI particles

A: Neutral lipid emulsions (NLEs) consist of DOPC (Dioleoyl-phosphatidylcholine)

Neutral charge but low efficiency of delivery to tumor

B: Dendrimers consist of poly(amideamine)- or poly(propyleneimine)-nucleic acid conjugate

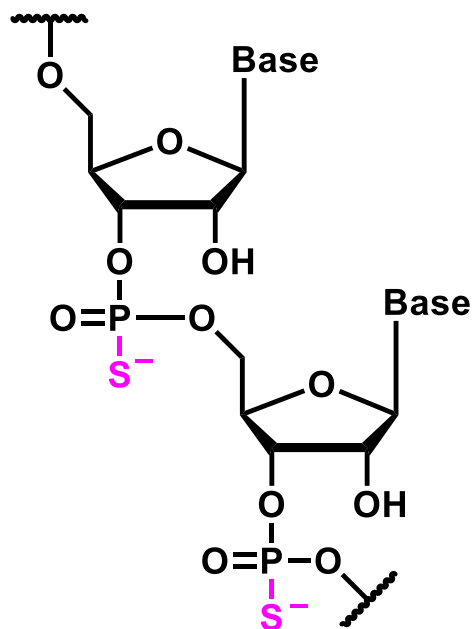
C: Proton-sponge with polyethylenimine (PEI)

High efficiency of delivery but often toxic due to cationic property

D: Poly(ethyleneglycol) (PEG)-RNA conjugate via disulfide bond

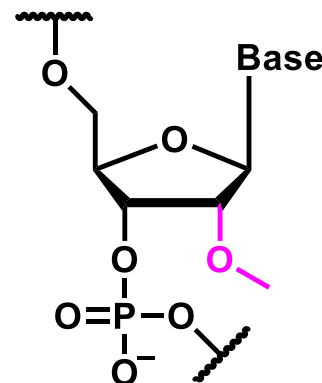
One of the most advanced system currently in clinical trials

1st and 2nd generation chemical modifications of RNA (phosphorothioate, 2'-*O*-alkylation)

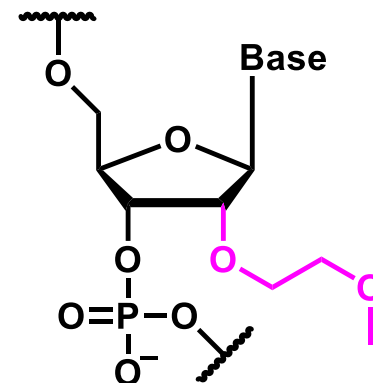


phosphorothioate

(the oxidation step of phosphoramidite method is replaced by sulfur transfer reaction)



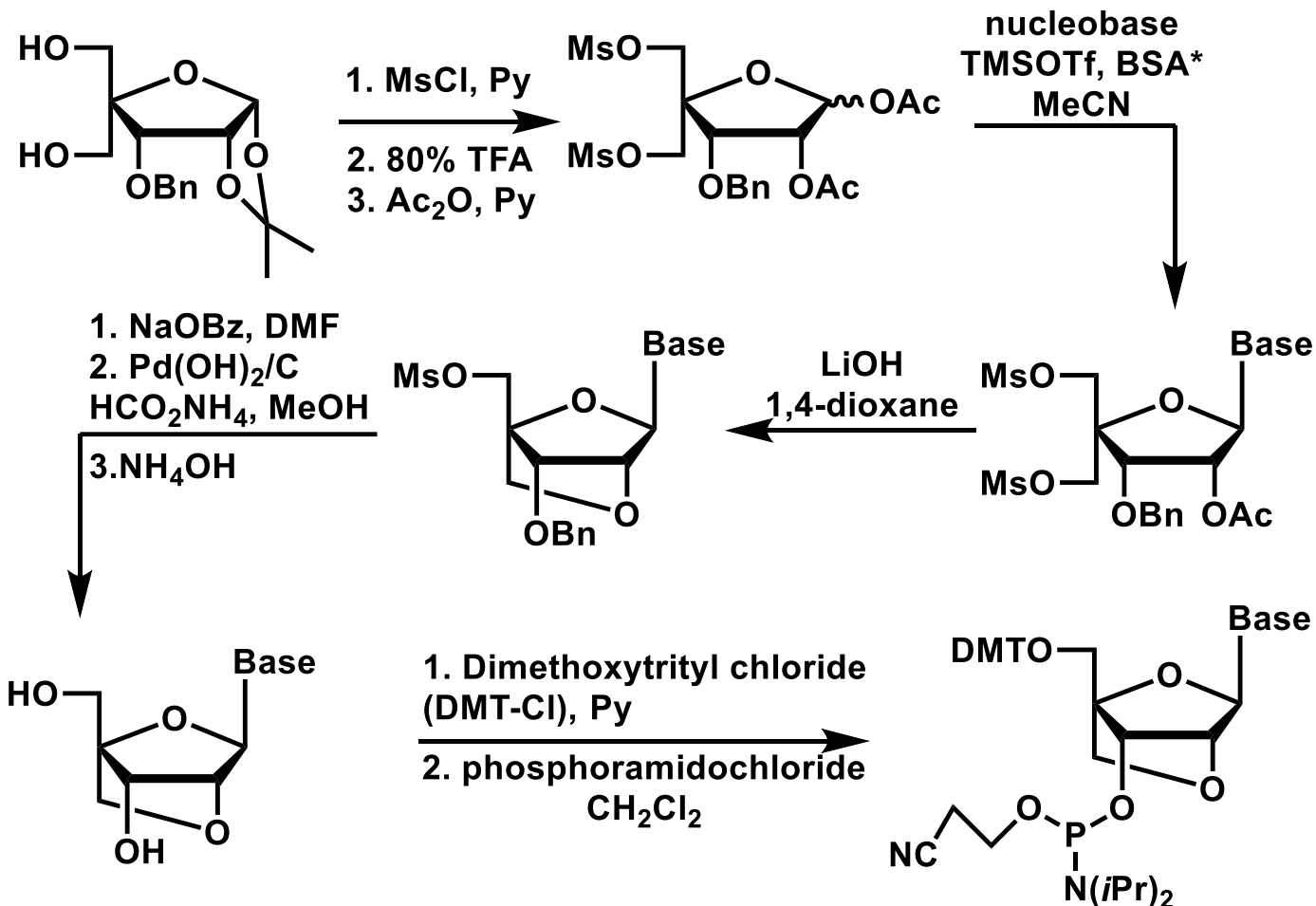
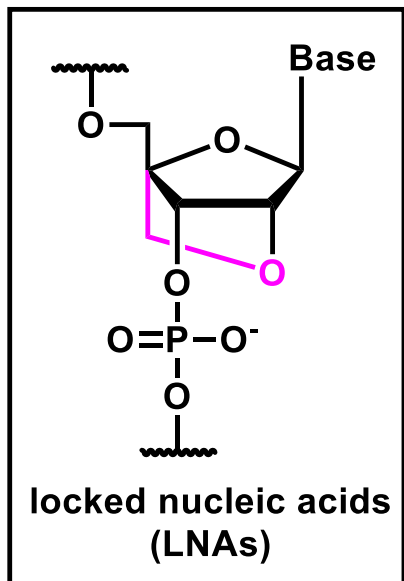
2'-*O*-methyl nucleotide



2'-*O*-methoxyethyl

- ✓ Only the (*S*)-P phosphorothioate diastereomer is nuclease resistant
- ✓ 2'-*O*-alkylation modification improves nuclease resistance and binding affinity

3rd generation-1 (LNAs)

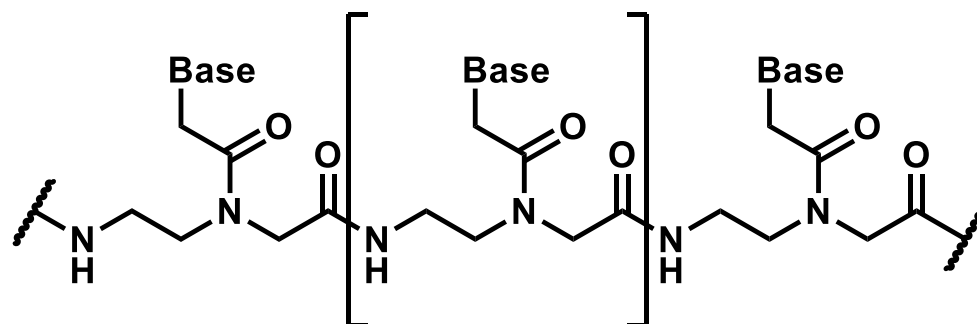


ASOs = antisense oligonucleotide

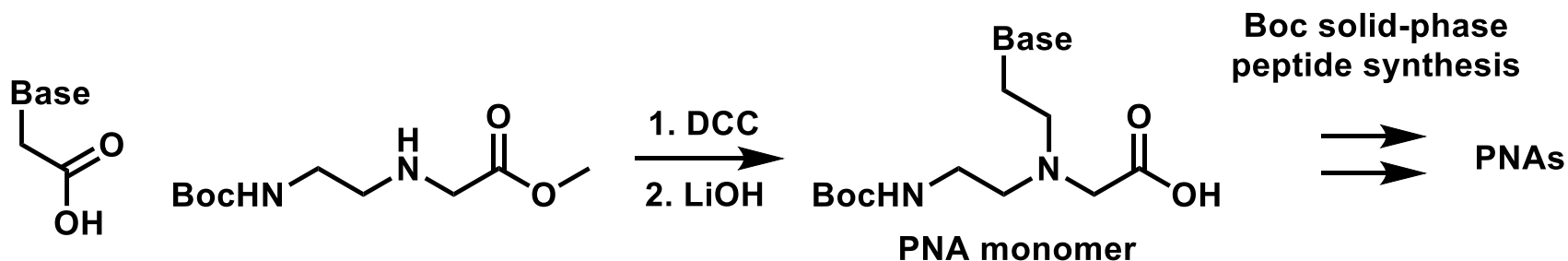
BSA* = *N,O*-bis(trimethylsilyl)acetamide

- ✓ LNAs improve sequence-specificity and stability of ASOs
- ✓ LNAs shows resistance to degradation by 3'-exonucleases

3rd generation-2 (Peptide nucleic acid)

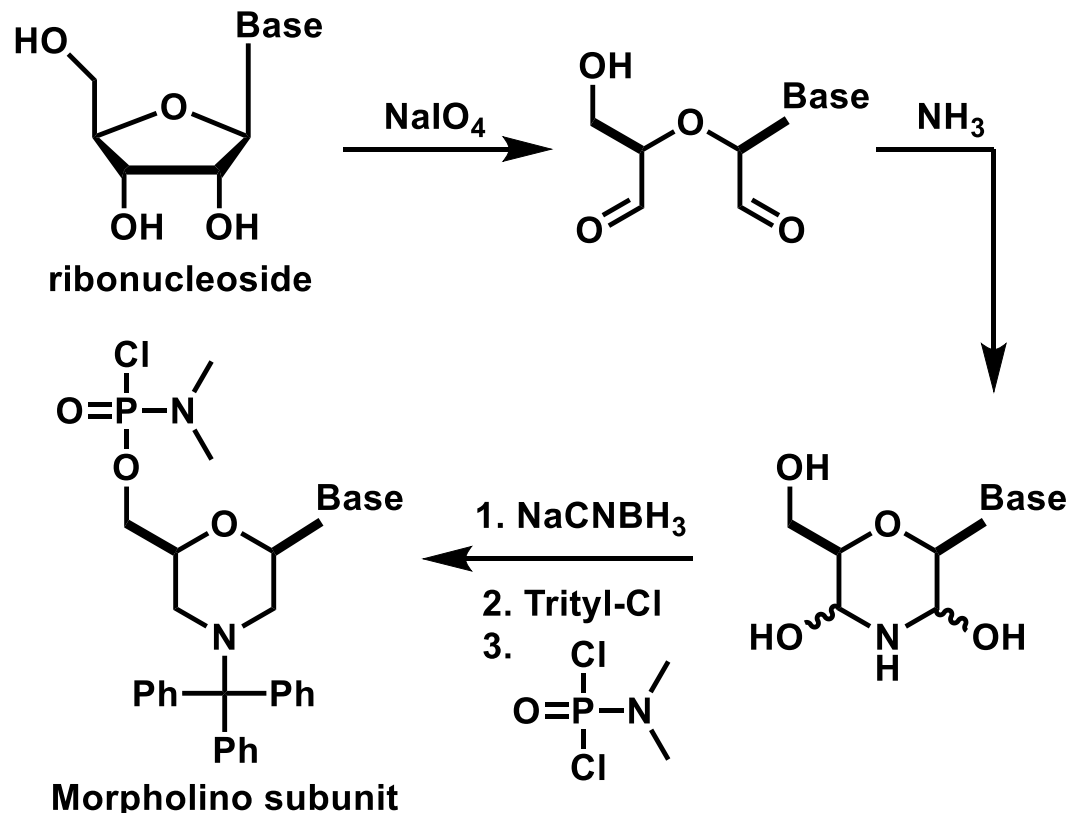
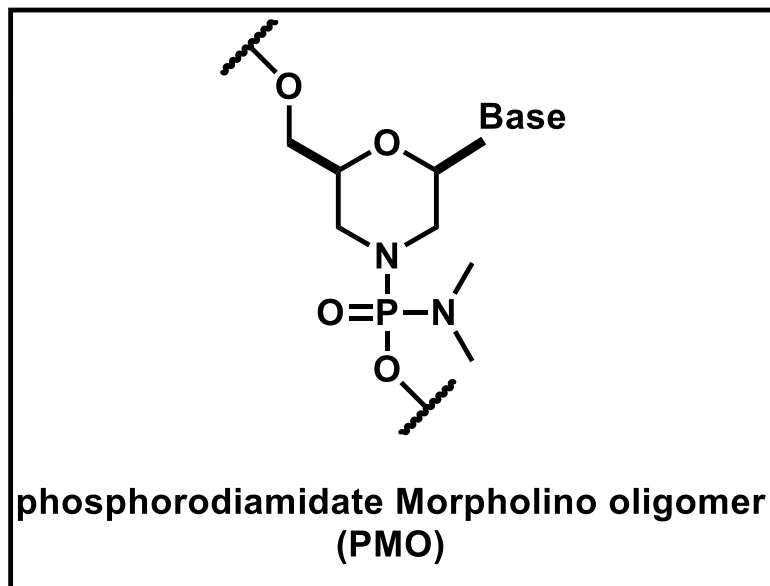


peptide nucleic acid (PNA)



- ✓ No negative charge to bind DNA more strongly than DNA
- ✓ High resistance to proteases and nucleases
- ✓ Stability in a wide range of pH

3rd generation-3 (Morpholino oligomer)



- ✓ Morpholinos do not degrade their target RNA molecules
- ✓ Acting by "steric blocking", binding to a target RNA
- ✓ Morpholinos are not recognized by cellular proteins
- ✓ Up to 18% of Morpholinos appear to induce nontarget-related phenotypes

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Small molecule-based approach with informatics



Prof. Matthew D. Disney

B.S., Chemistry, University of Maryland, College Park, 1997

M.S., Chemistry, University of Rochester, 1999

Ph.D., Biophysical Chemistry, University of Rochester, 2003 (Prof. Edwin L. Turner)

2002-2005 Postdoctoral Fellow, Swiss Federal Institute of Technology Zurich (ETH)

(Prof. Peter H. Seeberger)

2005-2010 Assistant Professor (Principle Investigator), University at Buffalo, New York

2011- Professor, Department of Chemistry, Graduate Program Faculty Member,

Kellogg School of Science and Engineering (THE SCRIPPS FLORIDA)

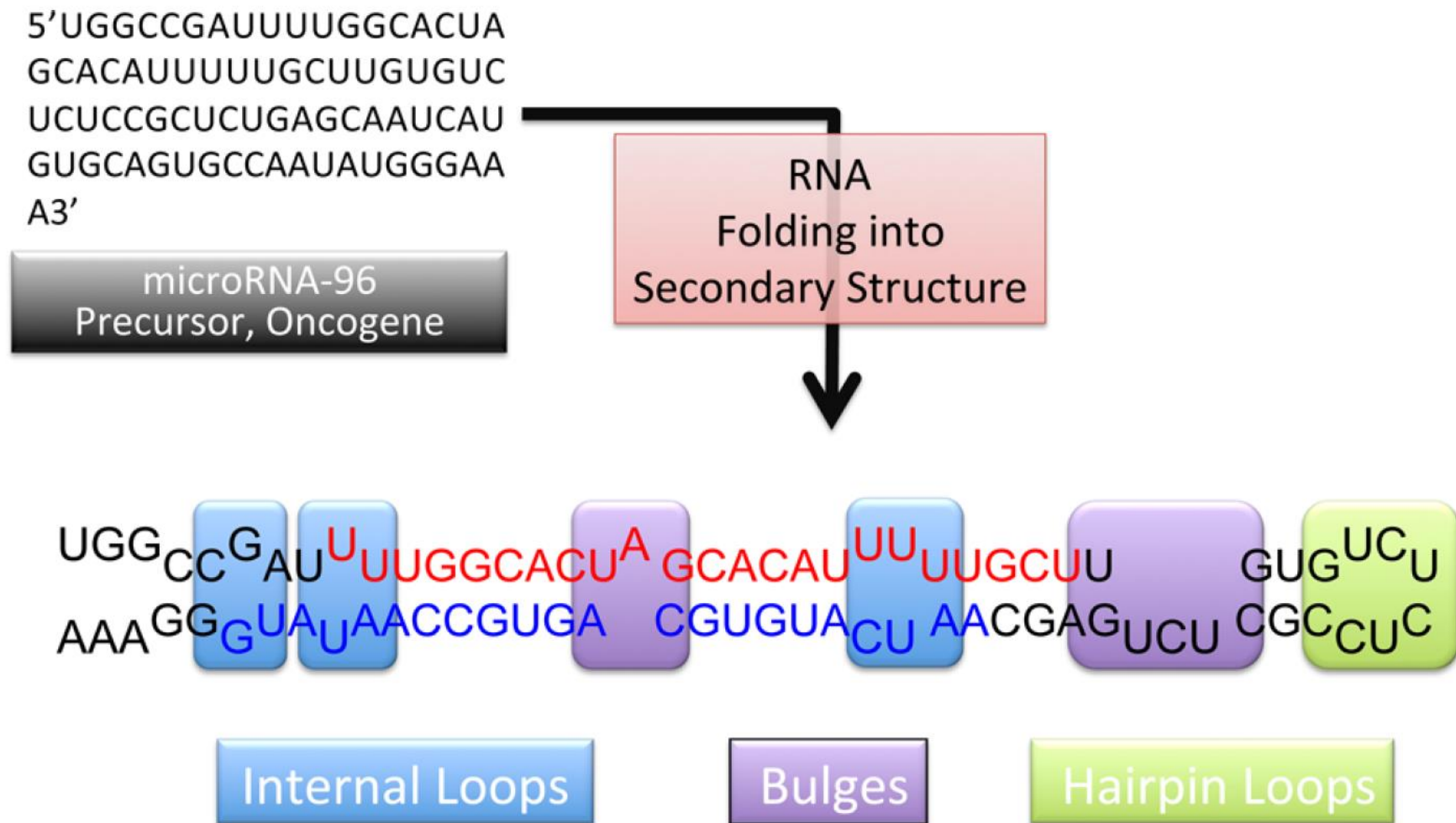
RNA is a highly desirable target for small molecule modulators

- ✓ Only 15% of all proteins are 'druggable' with small molecule, whereas 85% are not
- ✓ RNAs encode proteins are only a small portion (1-2%) and RNAs encode 'undruggable' proteins may be 'druggable'

Is RNA 'druggable' with small molecules?

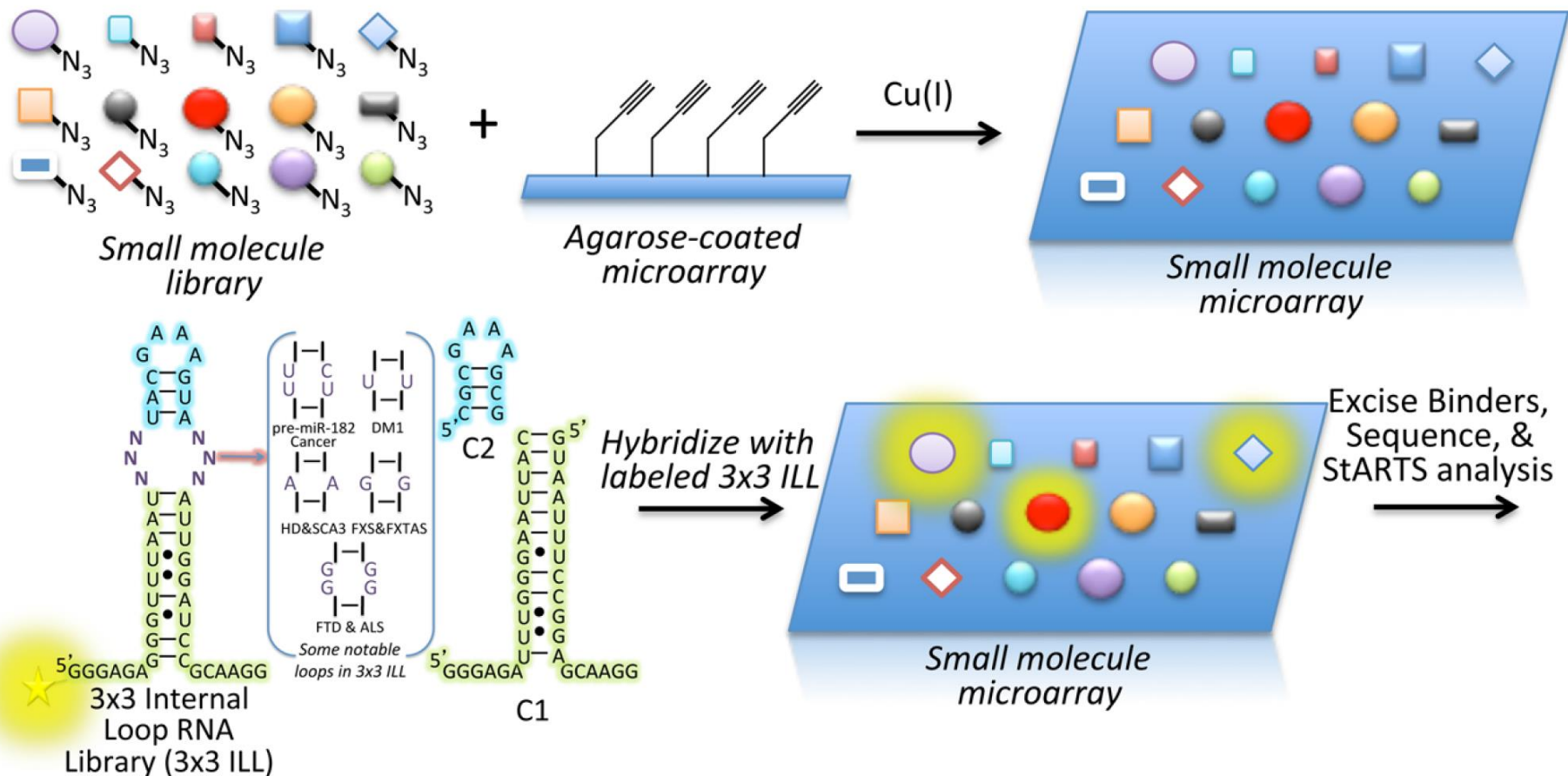
- ✓ No understanding about the RNA secondary structural motifs which can be binding site of small molecules
- ✓ Few small molecule elicit their effects by modulating RNA yet

Targeting noncanonical structures in RNAs



✓ These secondary structures can be deduced from sequences

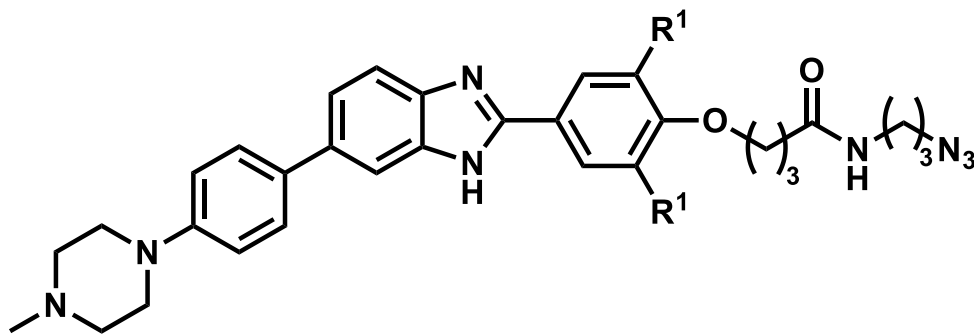
Two-dimensional combinatorial screening (2DCS)



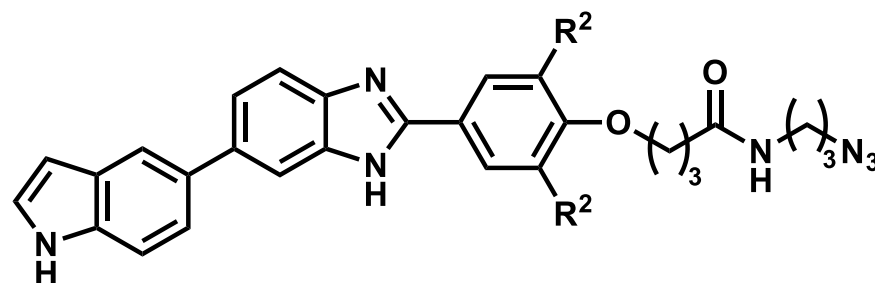
- ✓ A library vs library method to probe the small molecule-RNA motif bindings
- ✓ Resulting data comprise a database of binding partners

2DCS of an RNA-focused small molecule library

■ An RNA-focused small molecule library

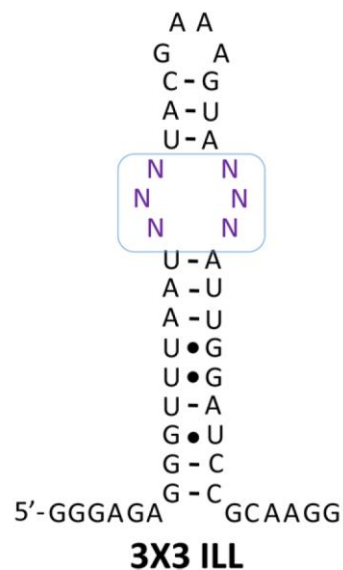


R¹ = H (1), Me (2), *i*-Pr (3), *t*-Bu (4)

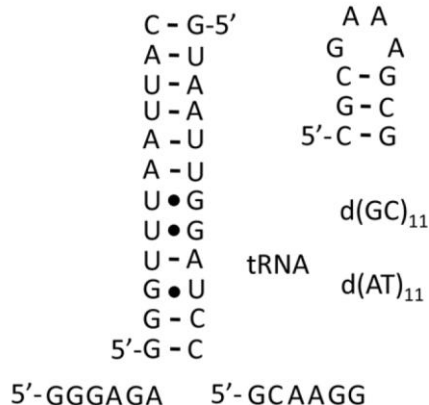


R² = H (5), Me (6), *i*-Pr (7), *t*-Bu (8)

■ An 3 × 3 nucleotide internal loop library (3 × 3 ILL)



Competitor Oligonucleotides



DNA Hairpin

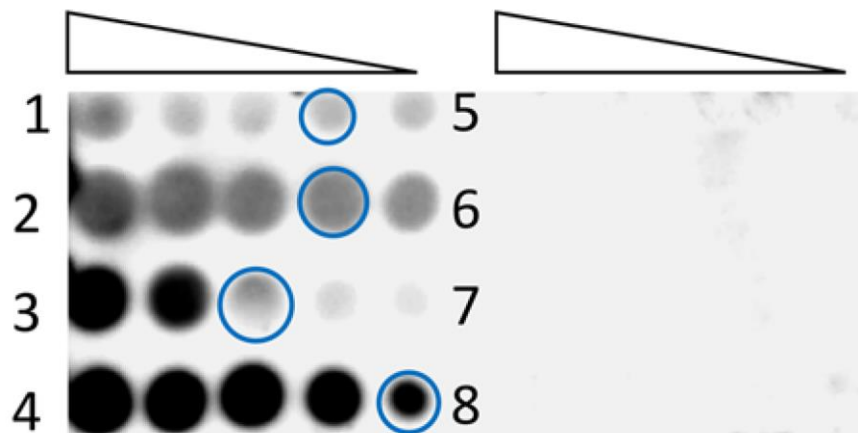


- ✓ ³²P-labeled
- ✓ 4⁶ = 4096 members

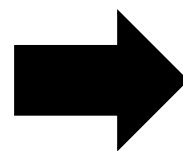
8 × 4096 = 32768 interactions were investigated

Results of 2DCS using 3 × 3 ILL

Small Molecule Loading

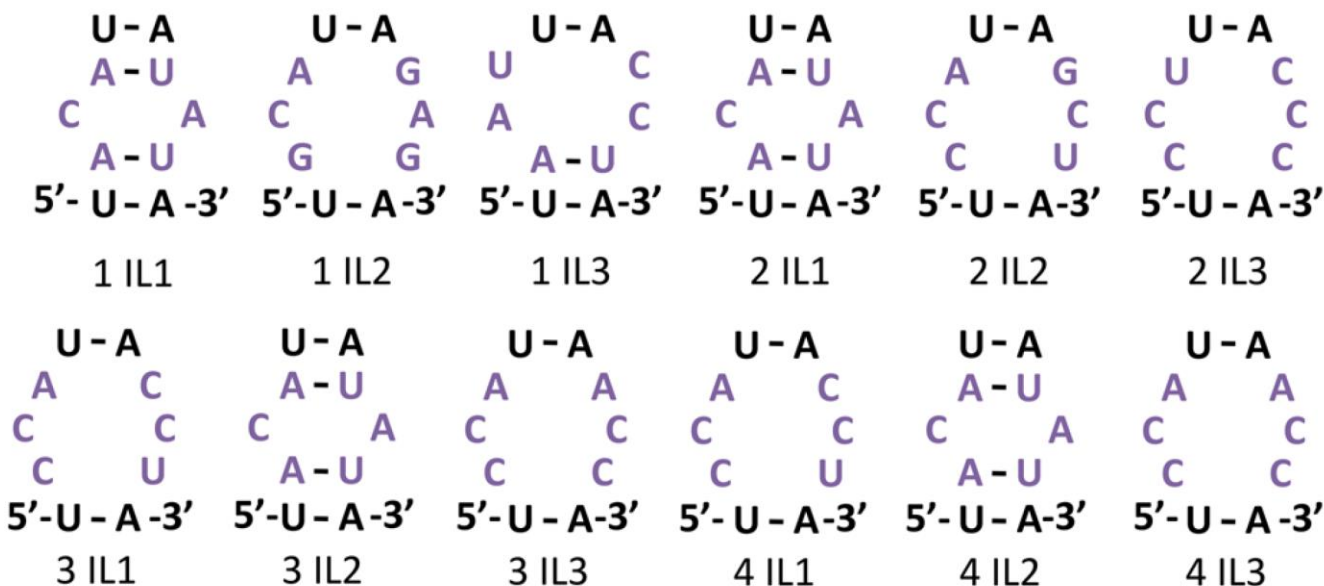


Immobilized molecules:
840, 560, 370, 250, 170
(picomoles)



*Harvested and
identified by RNA-seq*

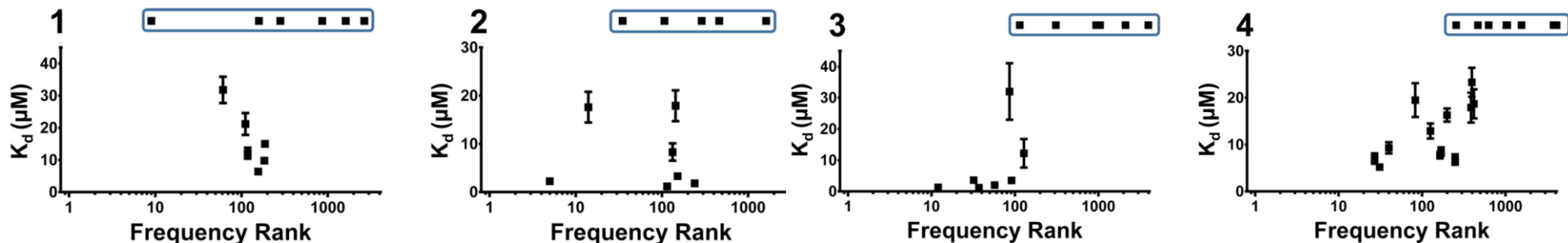
■ The top three selected RNA motif binding to 1-4



*Binding affinities
(K_d s) of the
identified pairs
were measured*

Investigation into global analysis of the interactions

■ Plots of frequency rank vs experimentally determined K_d



Due to the biases that occur during transcription and sequencing?

■ High Throughput Structure-Activity Relationship (HiT-StARTS) (Introduction of Z_{obs} rank to estimate the affinity accurately)

Pooled population comparison (Z-test)

$$(1) \phi = \frac{n_1 p_1 + n_2 p_2}{n_1 + n_2}$$

✓ n_1 is the size of population 1 (number of reads for a selected RNA)

✓ n_2 is the size of population 2 (number of reads for the same RNA from sequencing of the starting library)

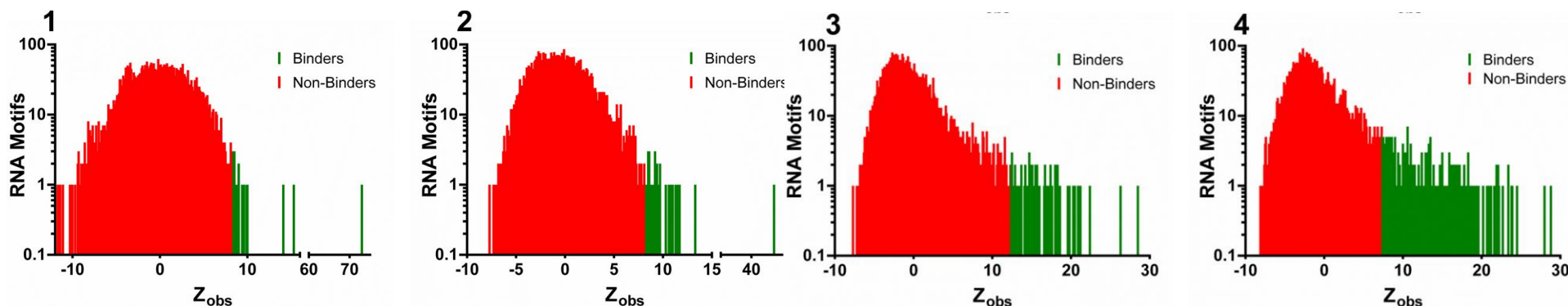
$$(2) Z_{\text{obs}} = \frac{p_1 - p_2}{\sqrt{\phi(1 - \phi)\left(\frac{1}{n_1} + \frac{1}{n_2}\right)}}$$

✓ p_1 is the observed proportion of population 1

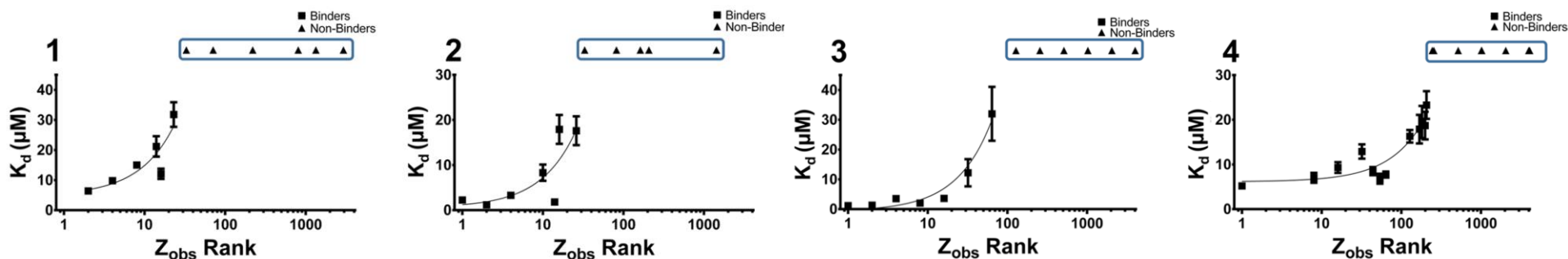
✓ p_2 is the observed proportion of population 2

Results of HiT-StART

■ Plots of frequency of RNA motifs as a function of Z_{obs}



■ Plots of Z_{obs} rank vs experimentally determined K_d

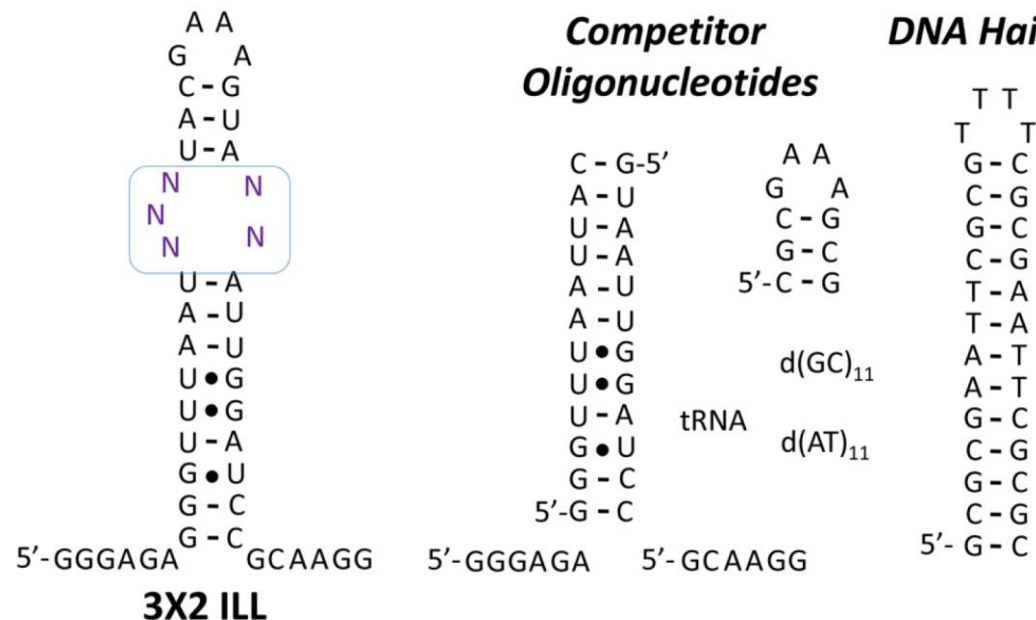


Excellent correlations were found between two values

- ✓ HiTStART analysis enables to estimate binding affinities rapidly and accurately
- ✓ Z_{obs} values can be normalized to *fitness score* (the most statistically significant RNA binder = 100% fitness)

HiT-StART applied to other RNA motif libraries

■ An 3×2 nucleotide internal loop library (3×2 ILL)

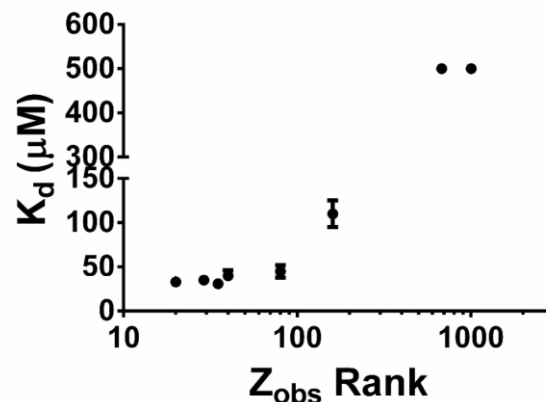


- ✓ ^{32}P -labeled
- ✓ $4^5 = 1024$ members
- ✓ vs the same small molecule library (1-8)

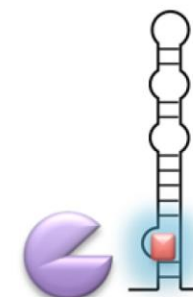
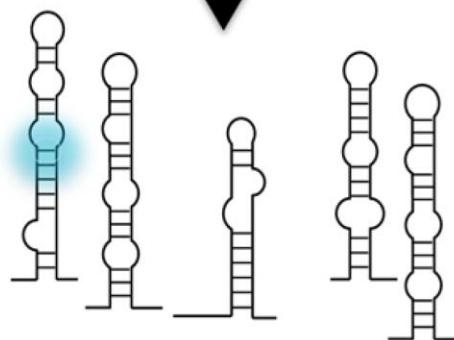
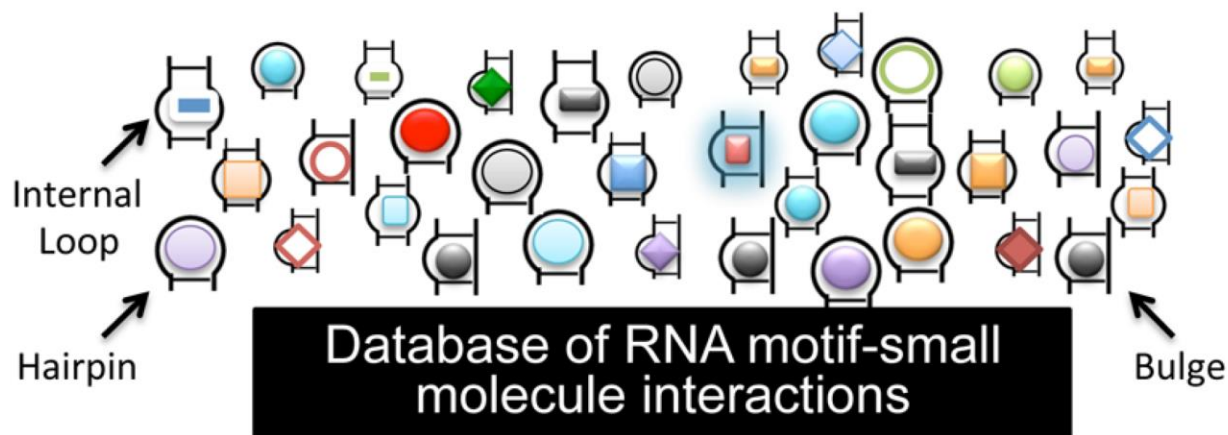
$8 \times 1024 = 8192$
interactions were investigated

- ✓ Only compounds **1-4** bind members of the RNA-library
- ✓ The interactions were well estimated by Z_{obs} value (\rightarrow)

3×2 ILL and compound **4** interactions



InfoRNA



Disease-causing
RNAs

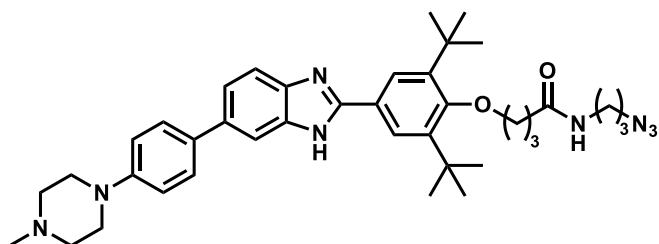
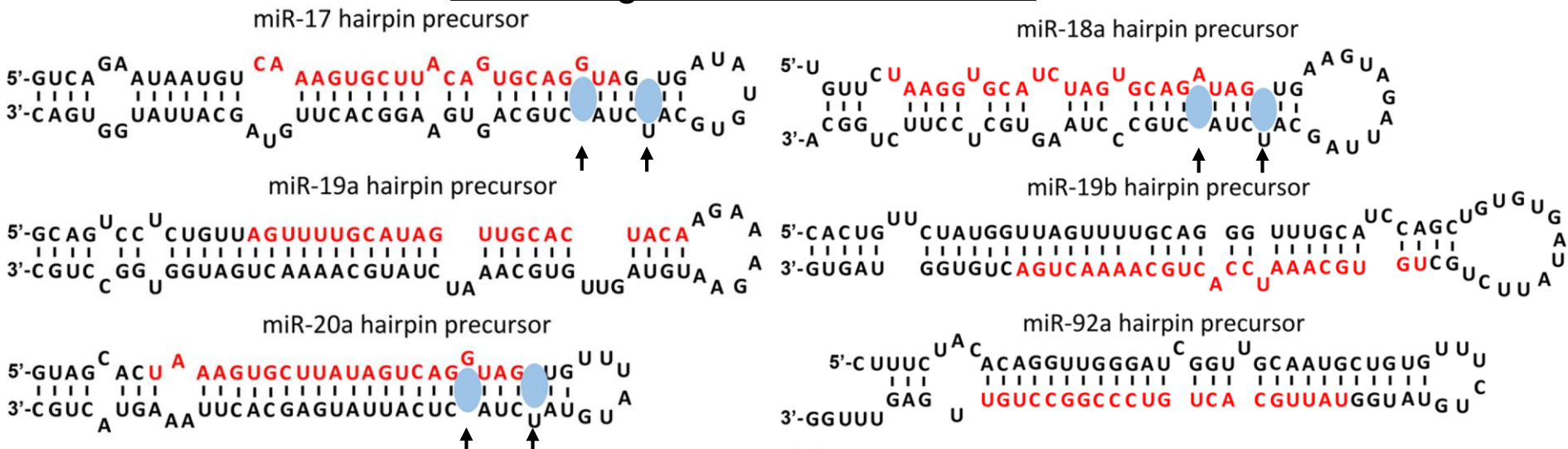
RNA Secondary
Structures

Small Molecule
Inhibition

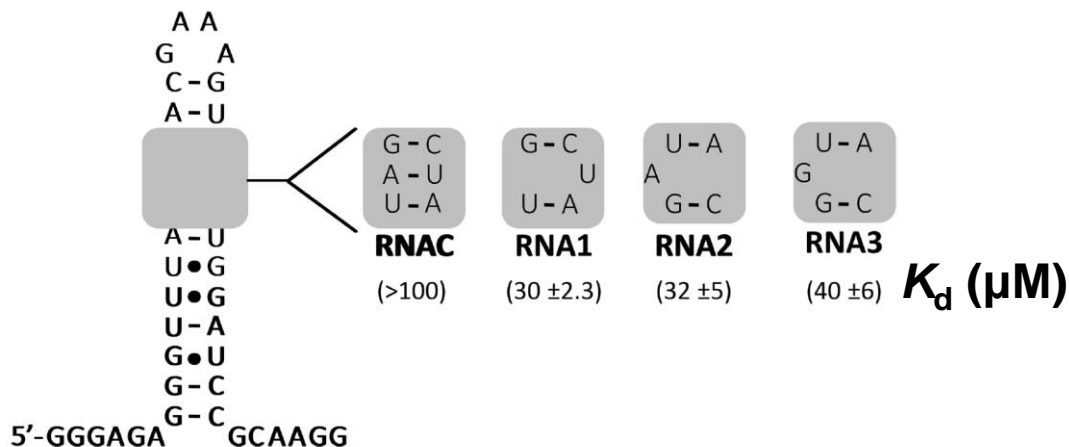
Identification of biologically important RNAs that can be targeted (InfoRNA analysis)

- Analysis focusing on miRNA associated with disease that have targetable motifs in Dicer/Drosha processing site

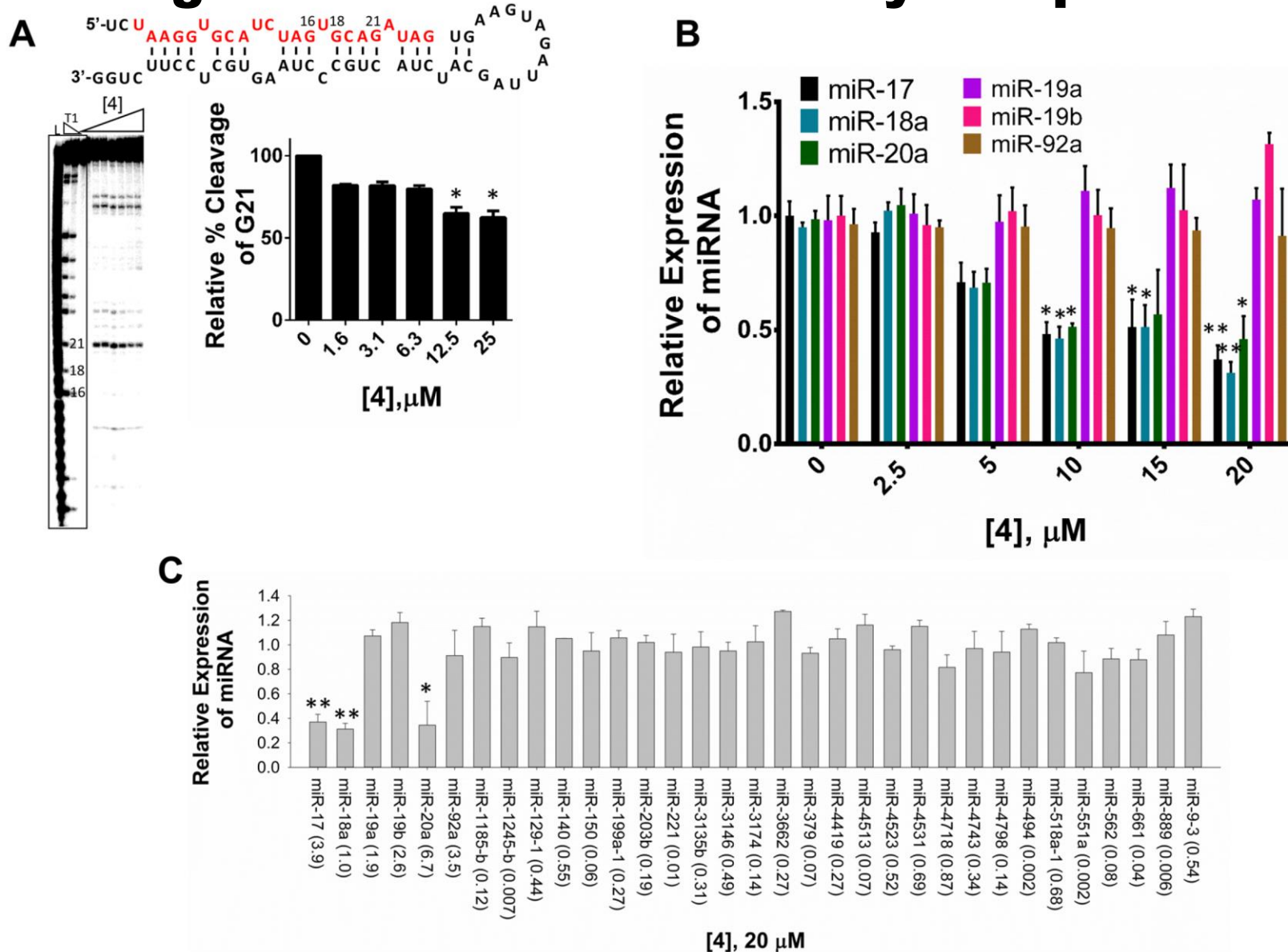
The oncogenic miR-17-92 cluster *Red = mature miRNA



100% fitness: 5'G_U/3'CUA
 91% fitness: 5'GAU/3'C_A
 78% fitness: 5'GGU/3'C_A

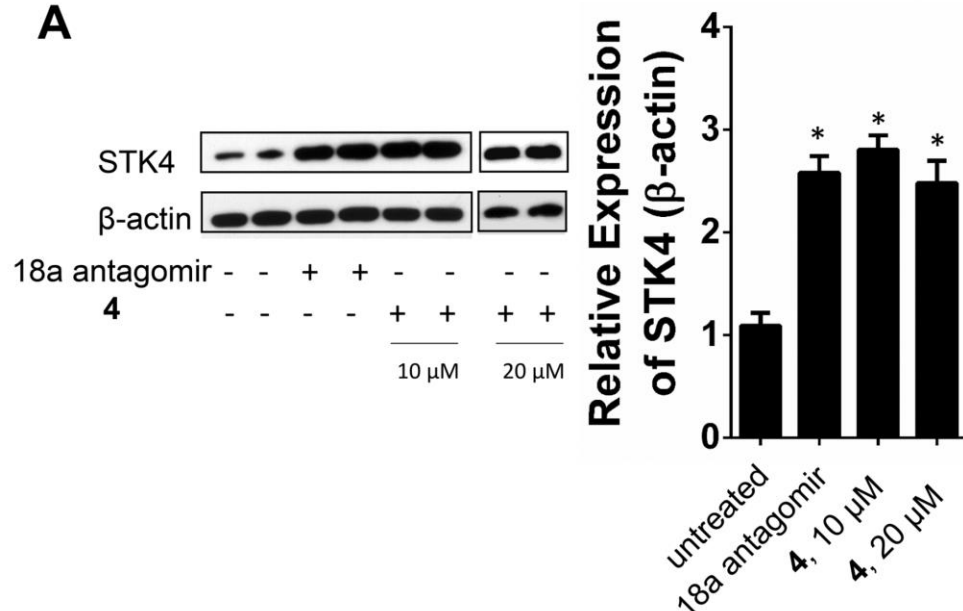


Inhibition of processing miRNA in the oncogenic miR-17-92 cluster by compound 4

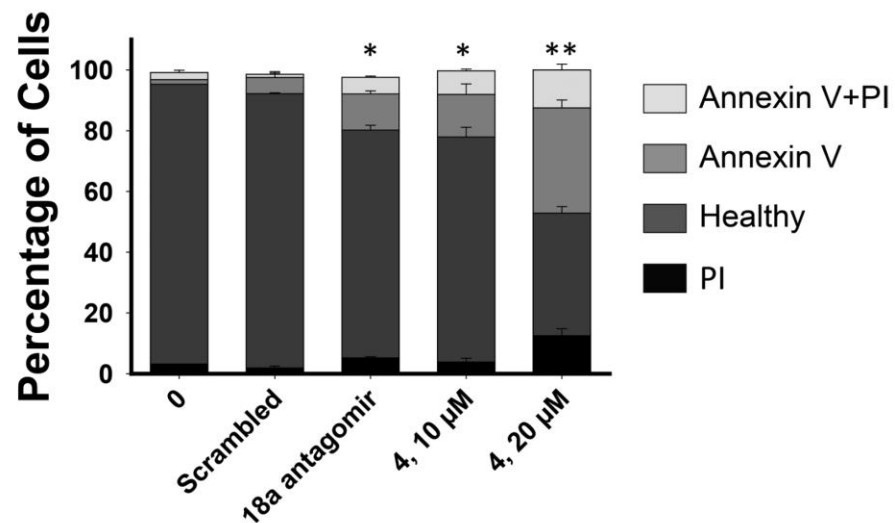


Downstream protein analysis and apoptosis detection

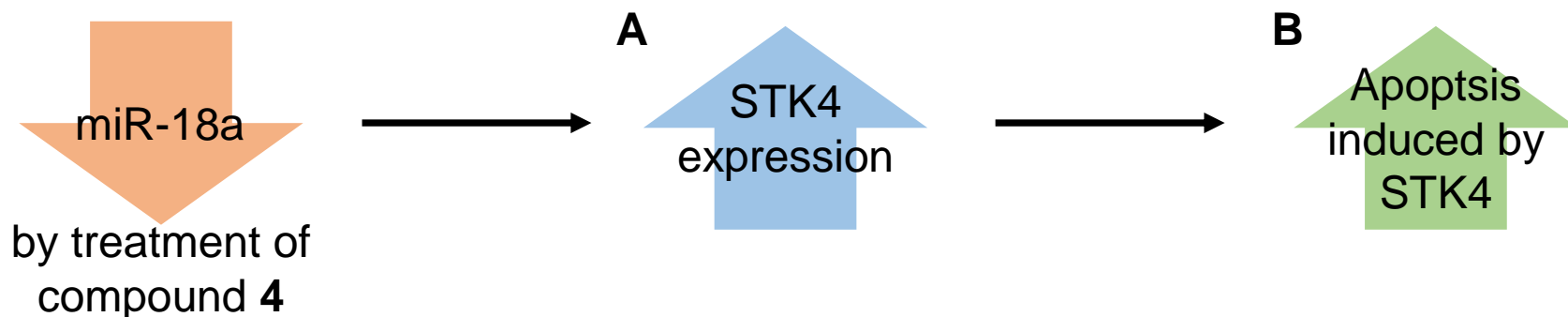
A



B

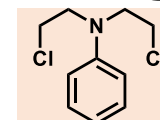
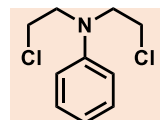


- ✓ Expression of serine/threonine protein kinase 4 (STK4) is repressed by miR-18a
- ✓ STK4 is a tumor suppressor in prostate cancer cells

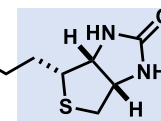
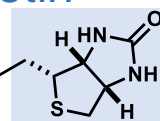


Chem-CLIP to study small molecule engagement

Cross-linking
module

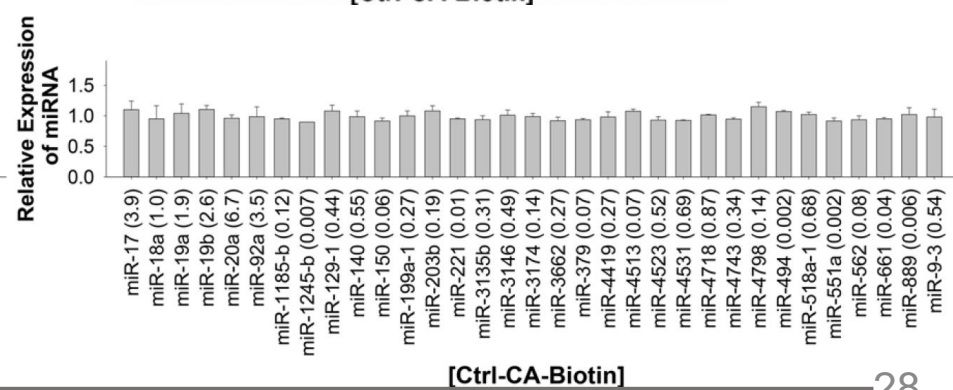
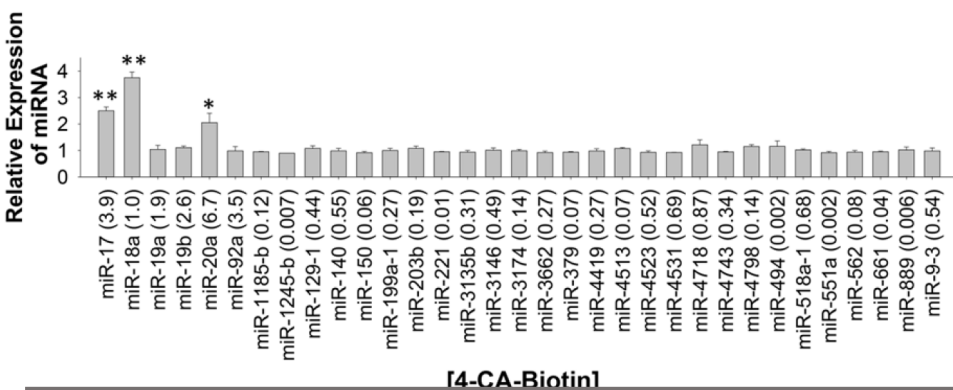
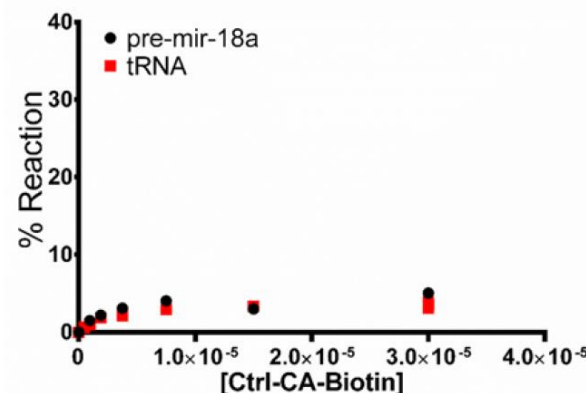
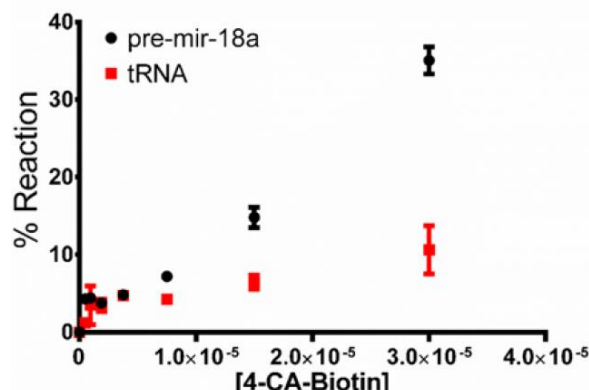


biotin

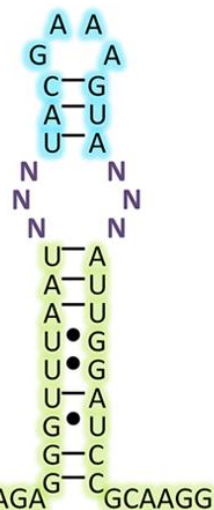


4-CA-biotin

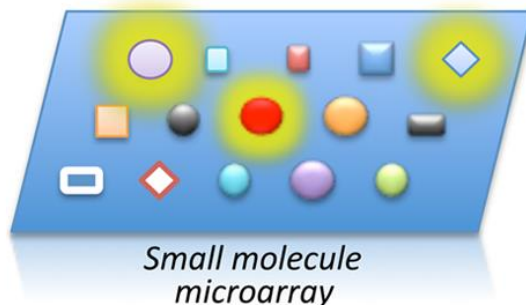
Ctrl-CA-biotin



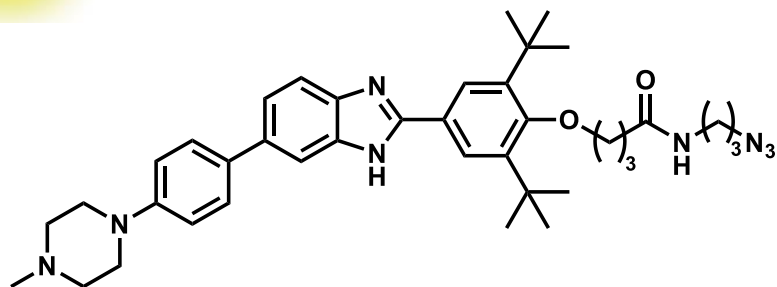
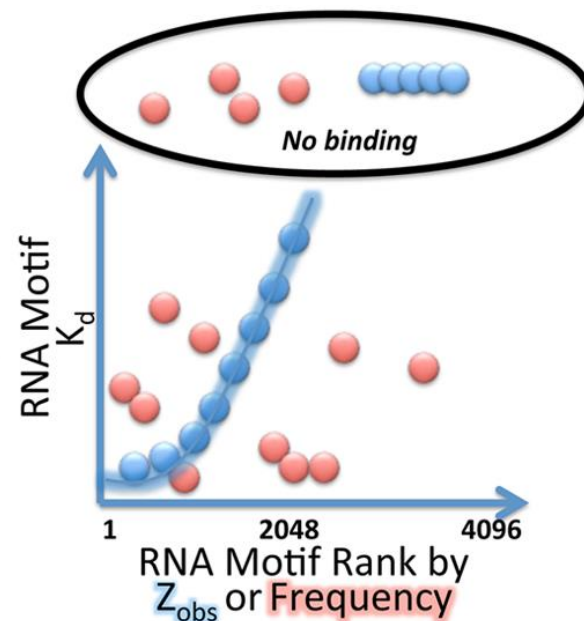
Short summary



2DCS

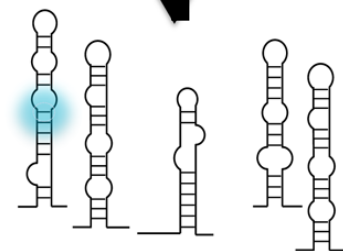


Excise Binders,
Sequence, &
HiT-StARTS
analysis



compound 4

infoRNA



RNA Secondary Structures

**Selective inhibition
of biogenesis of
miR-17, 18a, 20a**

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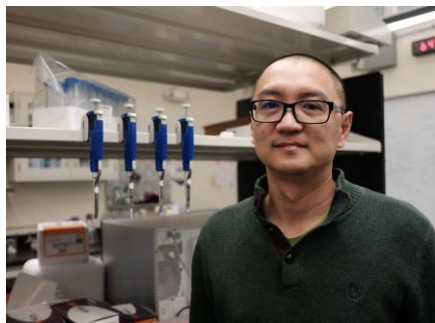
Rational Design of Small Molecule Targeting Noncoding RNAs
(Velagapudi, S. P. et al. *ACS Cent. Sci.* **2017**, 3, 205)

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Bifunctional strategy to inhibit miRNA biogenesis

Dr. Fu-Sen Liang



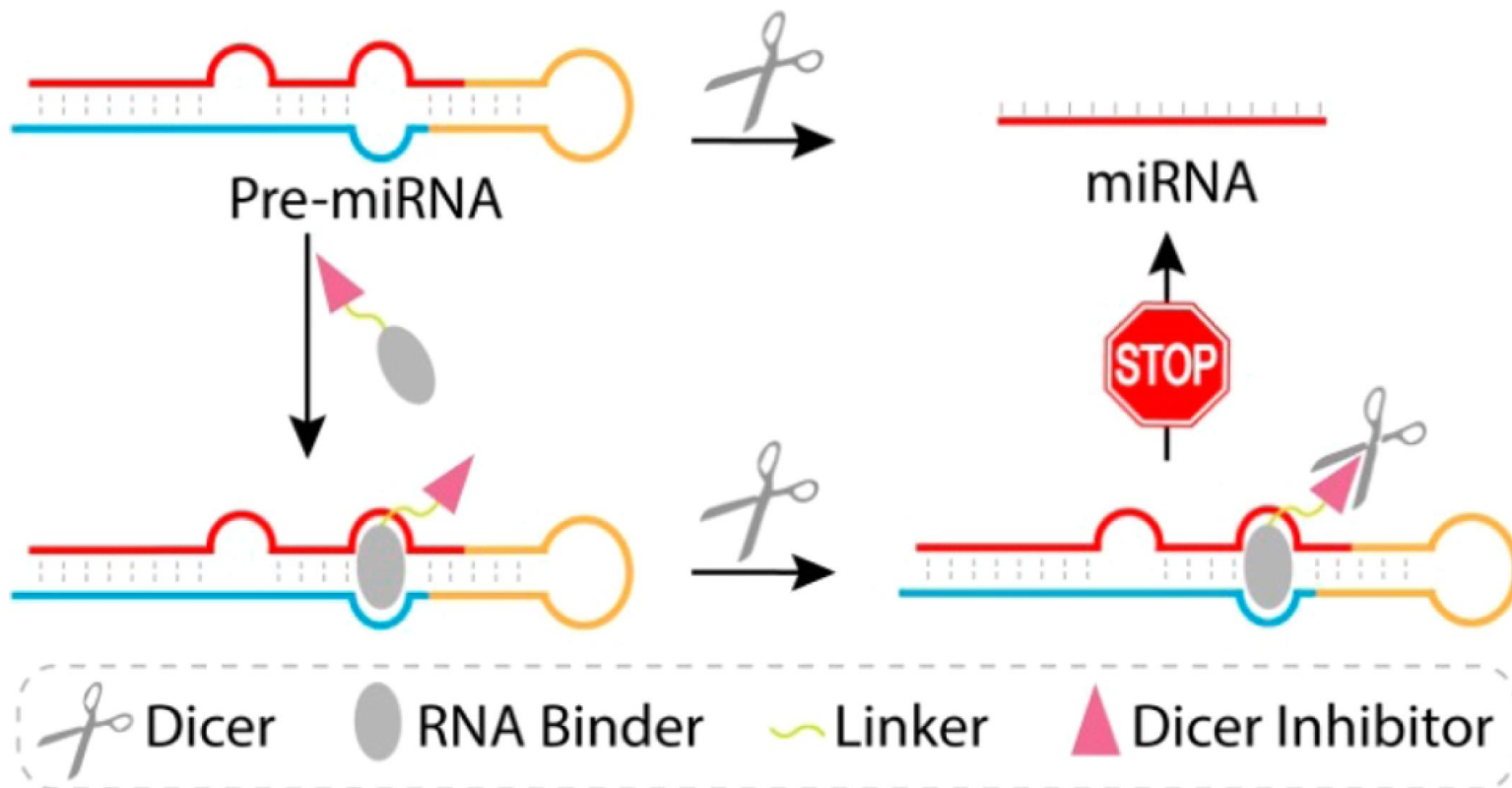
1994 BS in Chemistry, National Taiwan University

1996 MS in Organic Chemistry, National Chiao Tung University

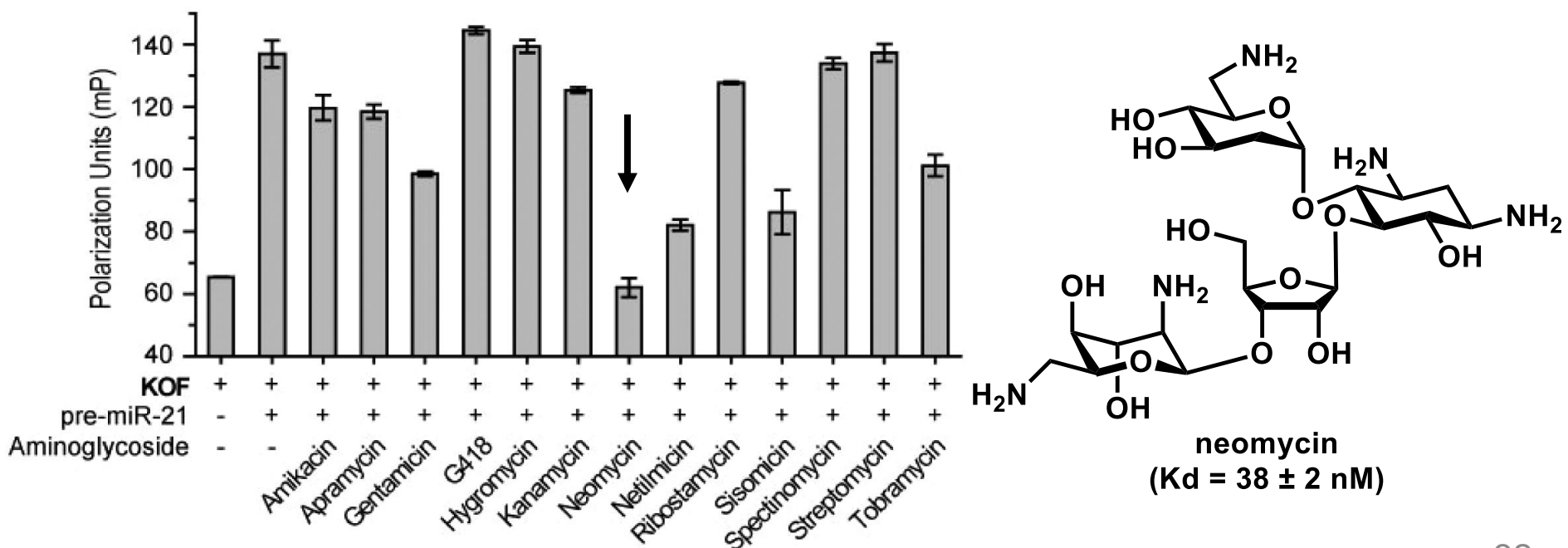
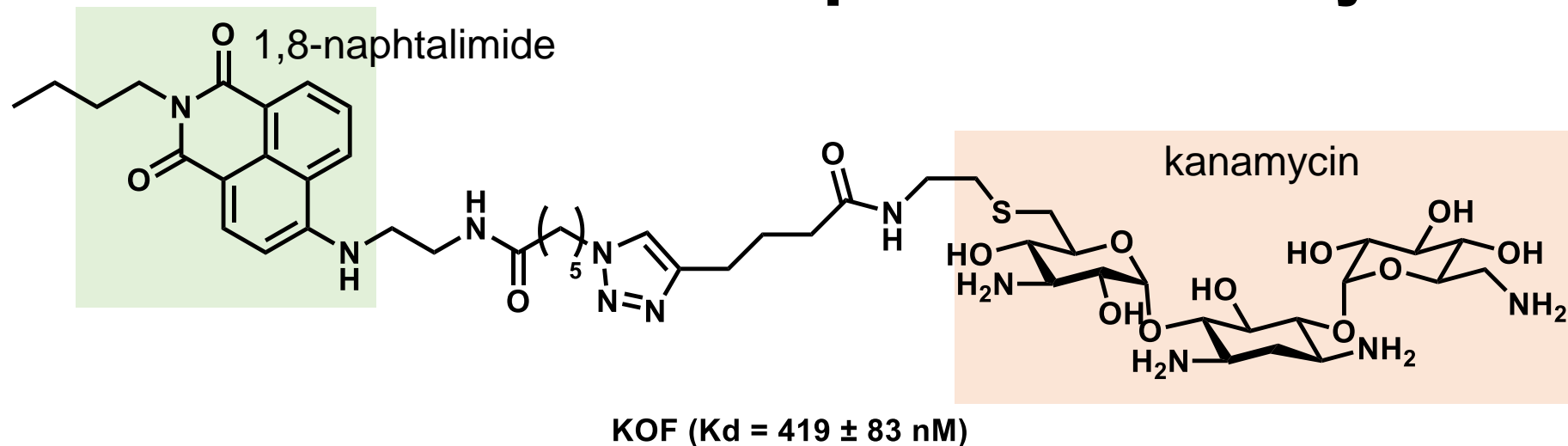
2005 Ph.D in Bioorganic Chemistry, The Scripps Research Institute, La Jolla, CA
(Prof. Chi-Huey Wong)

2005-2011 Postdoc in Chemical Biology, Stanford Medical School (Gerald R. Crabtree)

2012- Assistant professor (independent research group), University of New Mexico



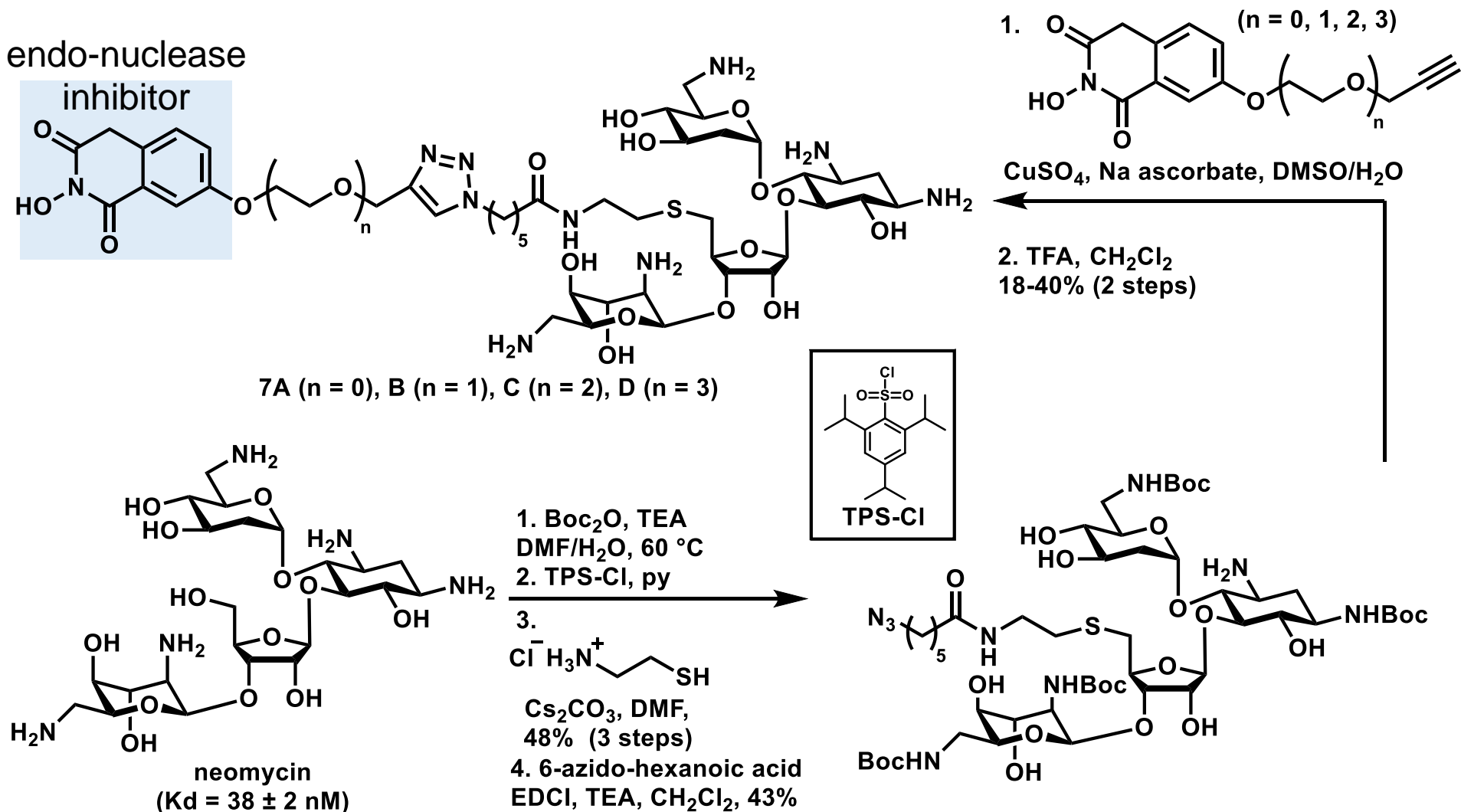
Screening of aminoglycosides binding to pre-miR-21 with a fluorescence polarization assay



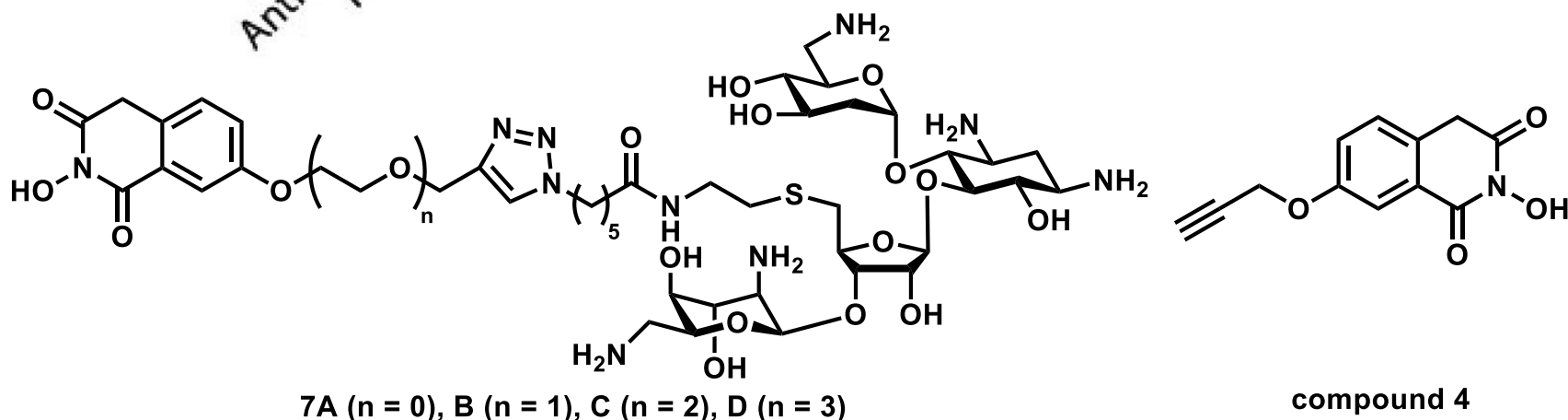
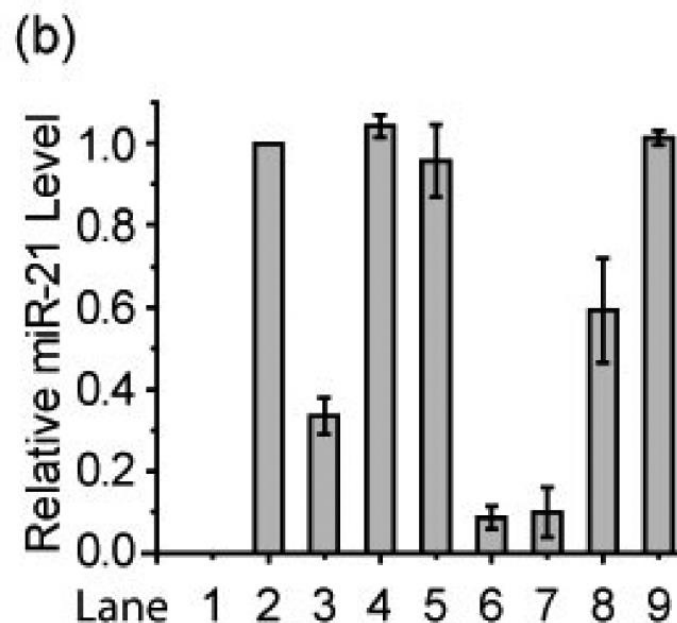
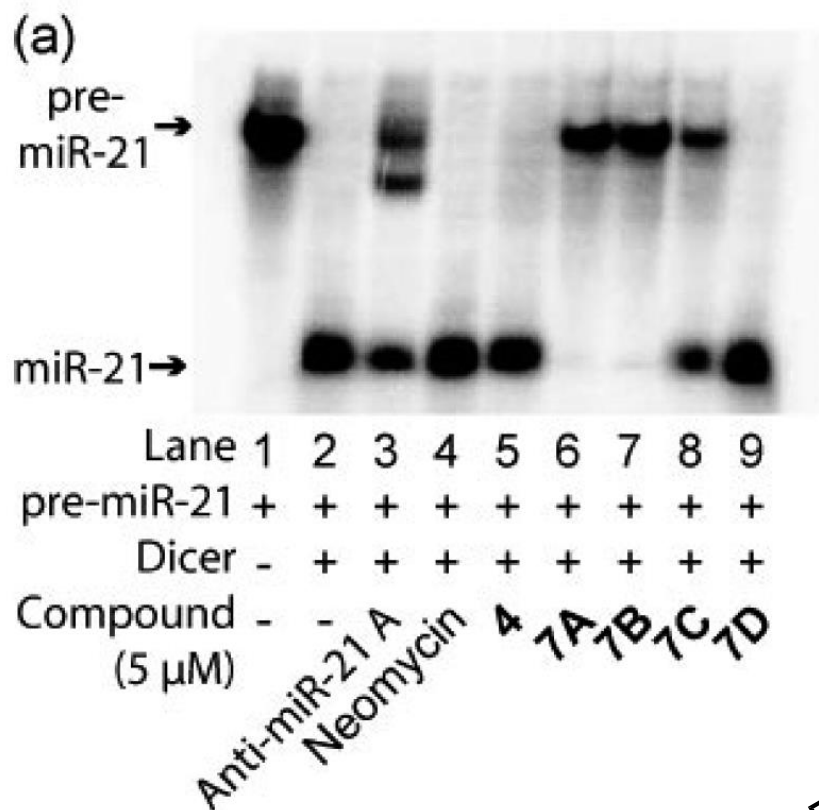
Design and synthesis of bifunctional small molecules

endo-nuclease

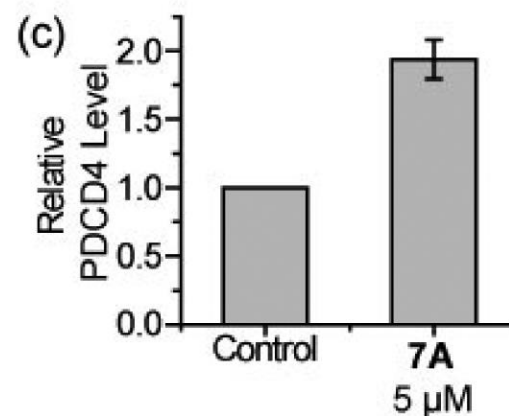
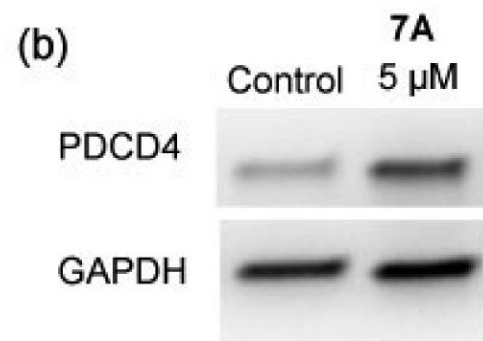
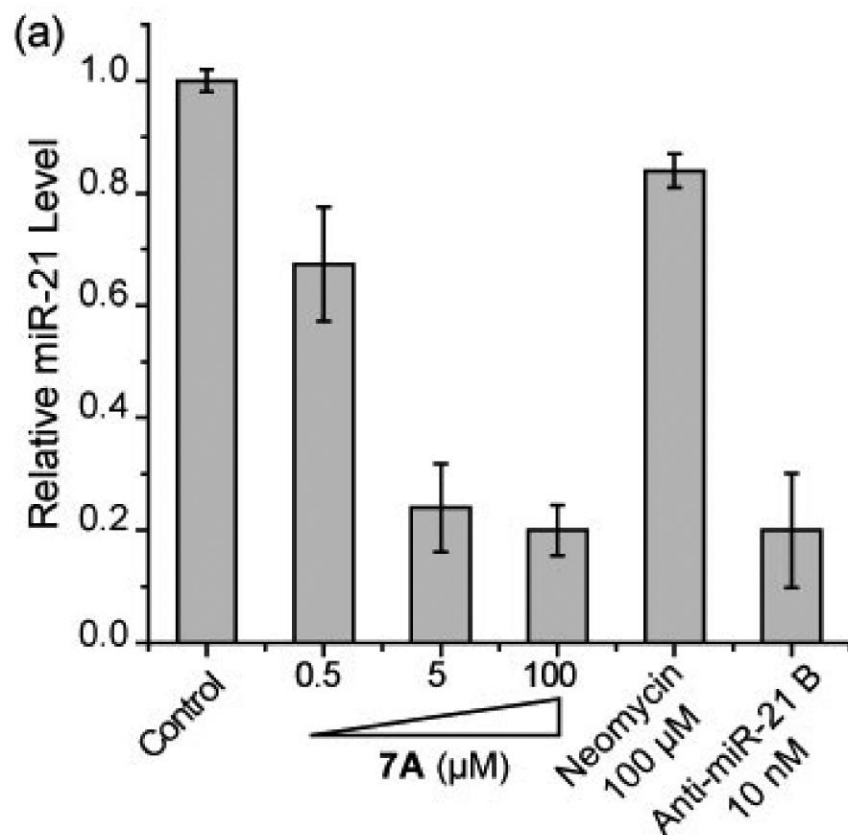
inhibitor



In vitro Dicer-mediated pre-miR-21 cleavage



***In cell* analysis of mature miR-21 expression levels and the downstream protein (PDCD4)**



miR-21
by treatment of
compound **7A**

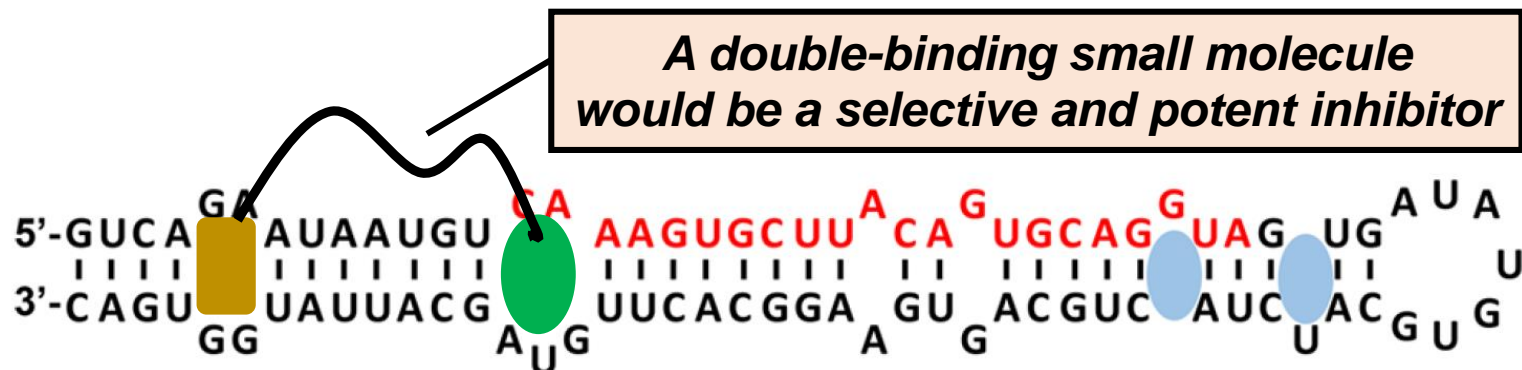


PDCD4
expression

PDCD4: Programmed
Cell Death Protein 4

Summary

- miRNAs are desirable therapeutic target
 - ✓ Modified antisense oligonucleotides with drug delivery systems are the main stream of strategies
 - ✓ The small molecule-based approach is a new field and has a great potential
- Problems and a proposal in small molecule-based approach
 - ✓ Small molecules seem to have a low affinity (up to 100 μ M) to RNAs (maybe due to the small binding site on RNAs)
 - ✓ Off-target effects have to be well considered
 - ✓ Can combination of the two topics make the affinity and the selectivity much higher?



Appendix-1

Table 2 | **Selected list of miRNA therapeutics in clinical trials**

Name (company)	Therapeutic agent	Delivery system	Target diseases	Trial details	ClinicalTrials.gov identifier
<i>miRNA-based therapeutics</i>					
Mirvirasen (Santaris Pharma A/S and Hoffmann-La Roche)	AntimiR-122	LNA-modified antisense inhibitor	Hepatitis C (chronic infections included)	Single-centre phase I, completed	NCT01646489
				Multicentre phase II, completed	NCT01200420
				Multicentre phase II, ongoing	NCT01872936
				Single-centre phase II, ongoing	NCT02031133
				Single-centre phase II, ongoing	NCT02508090
RG-101 (Regulus Therapeutics)	AntimiR-122	GalNAc-conjugated antimiR	Chronic hepatitis C	Phase I, completed	–
				Multiple phase II, ongoing	–
RG-125/ AZD4076 (Regulus Therapeutics)	AntimiR-103/107	GalNAc-conjugated antimiR	Patients with type 2 diabetes and non-alcoholic fatty liver diseases	Single-centre phase I, ongoing	NCT02612662
				Single-centre phase I/IIa, ongoing	NCT02826525
MRG-106 (miRagen Therapeutics)	AntimiR-155	LNA-modified antisense inhibitor	Cutaneous T cell lymphoma and mycosis fungoides	Multicentre phase I, ongoing	NCT02580552
MRG-201 (miRagen Therapeutics)	miR-29 mimic	Cholesterol-conjugated miRNA duplex	Scleroderma	Single-centre phase I, ongoing	NCT02603224
MesomiR-1 (EnGeneIC)	miR-16 mimic	EnGeneIC delivery vehicle	Mesothelioma, non-small cell lung cancer	Multi-centre Phase I, ongoing	NCT02369198
MRX34 (Mirna Therapeutics)	miR-34 mimic	LNPs (Smarticles)	Multiple solid tumours	Multicentre phase I, terminated	NCT01829971

DOPC, 1,2 dioleoyl-sn glycerol-3 phosphatidylcholine; eIF, eukaryotic initiation factor; GalNAc, N-acetyl-D-galactosamine; HBV, hepatitis B virus; LNA, locked nucleic acid; LNPs, lipid nanoparticles; miRNA, microRNA; PEI, polyethylenimine; RSV, respiratory syncytial virus.

Appendix-2: synthesis of compound 4 in main topic

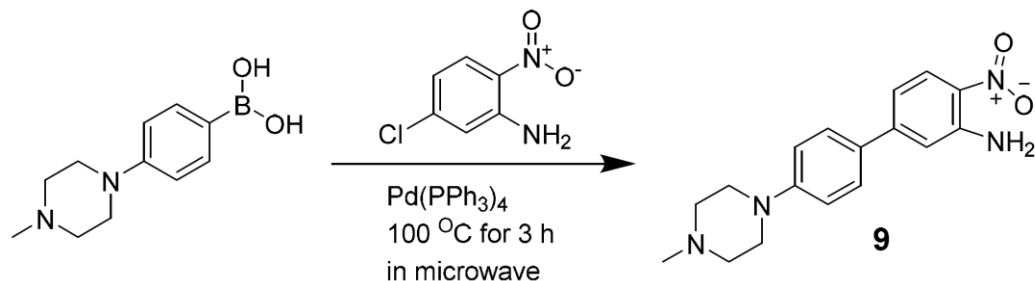


Figure S-1: Synthetic scheme of 4'-[4-(4-methylpiperazin-1-yl)-4-nitro-1,1'-biphenyl]-3-amine (**9**)

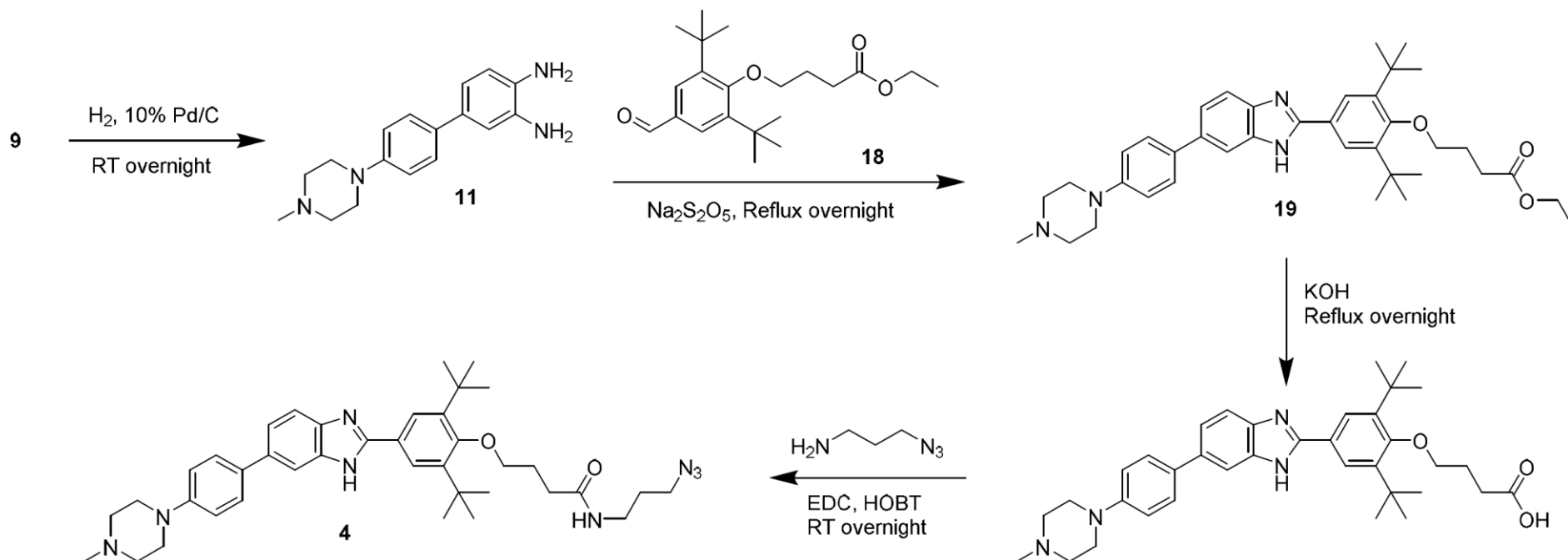
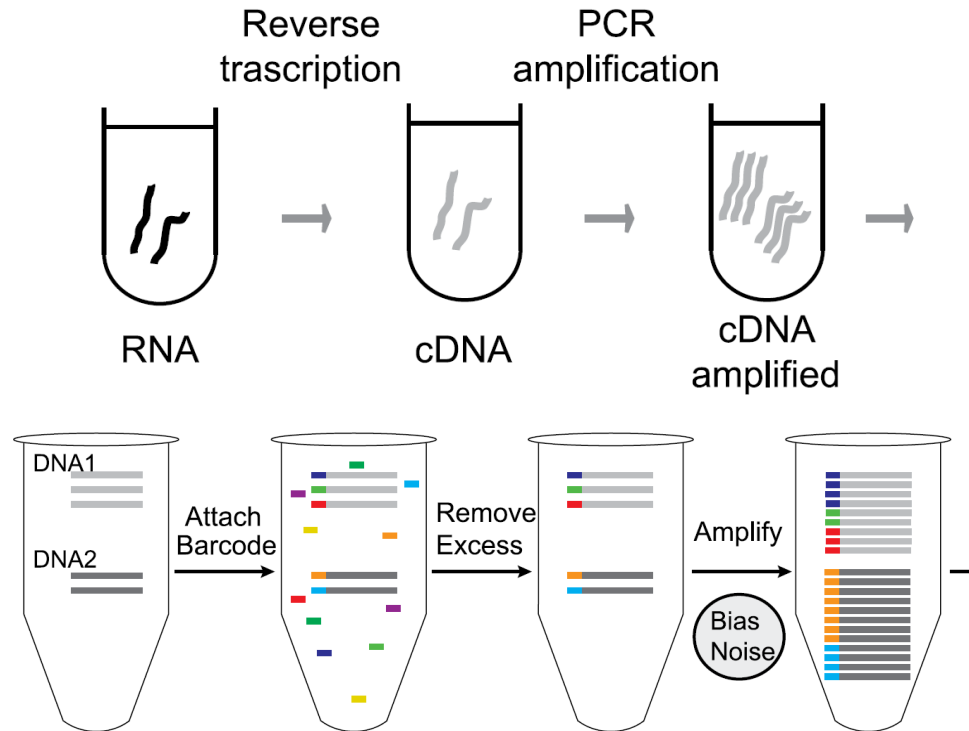


Figure S-22: Synthetic scheme of **4**

Appendix-3: quantitative RNA-seq



	DNA1	DNA2	Ratio
Number of Molecules in Original Sample	3	2	3 : 2
Number of Sequence Reads (Conventional Counting)	9	12	3 : 4
Number of Molecules Detected by Counting Unique Barcodes (Digital Counting)	3	2	3 : 2

Massively Parallel Sequencing →