

# Voltage-sensitive Probes

## Transmembrane potential

1. Electrophysiology

2. Ca<sup>2+</sup>-sensitive probes

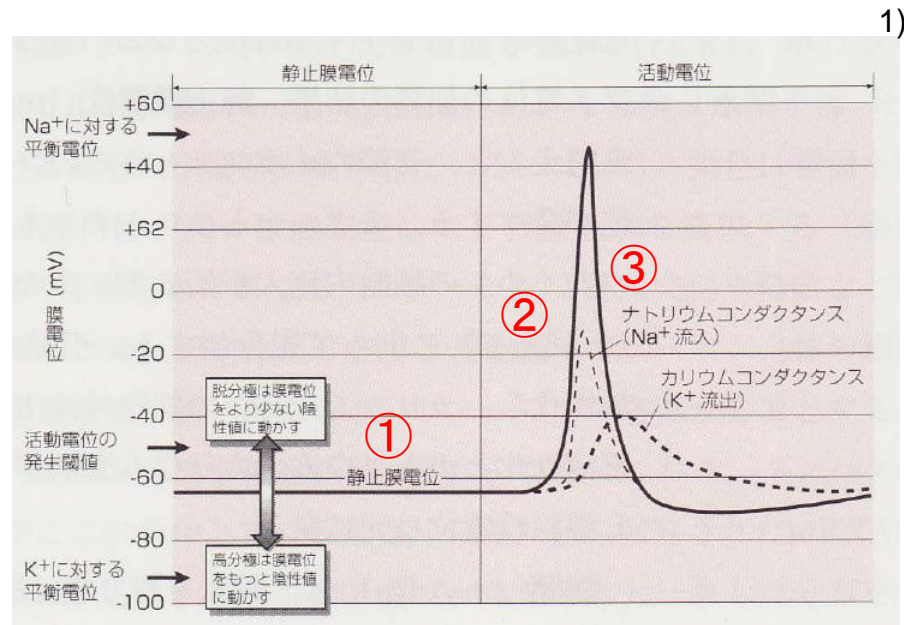
3. Voltage-sensitive probes

3-1. Electrochromic

3-2. FRET

3-3. PeT (main paper)

# Transmembrane potential -active potential-



① Polarization (different ion concentrations among inside and outside the membrane)

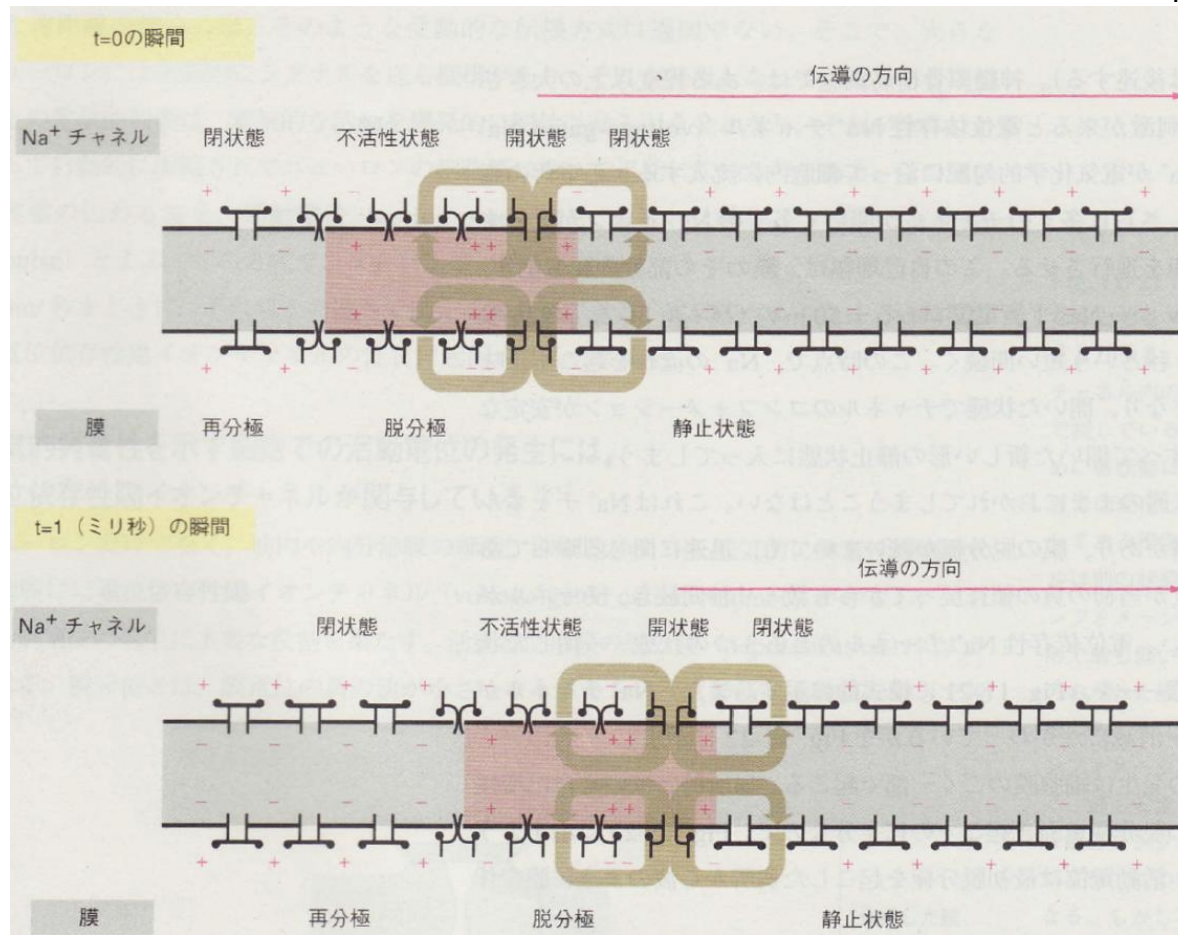
↓ Stimulation

② Depolarization (Na<sup>+</sup> influx)

↓

③ Polarization (K<sup>+</sup> efflux)

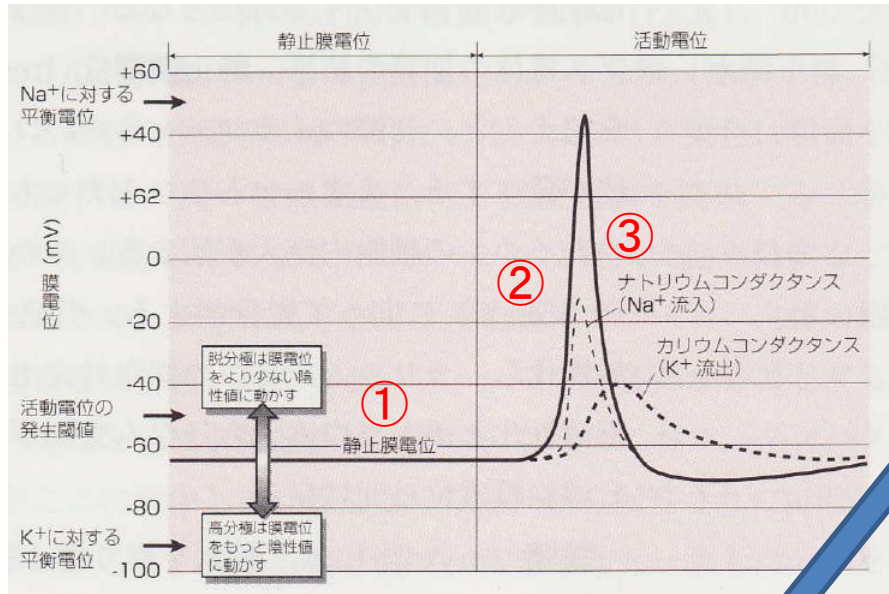
# Transmembrane potential -signal transmission- 1)



Signal transmission with action potential

# Transmembrane potential -detection-

1)



1. Electrophysiology

2. Ca<sup>2+</sup>-sensitive probes

3. Voltage-sensitive probes

① Polarization (different ion concentrations among inside and outside the membrane)

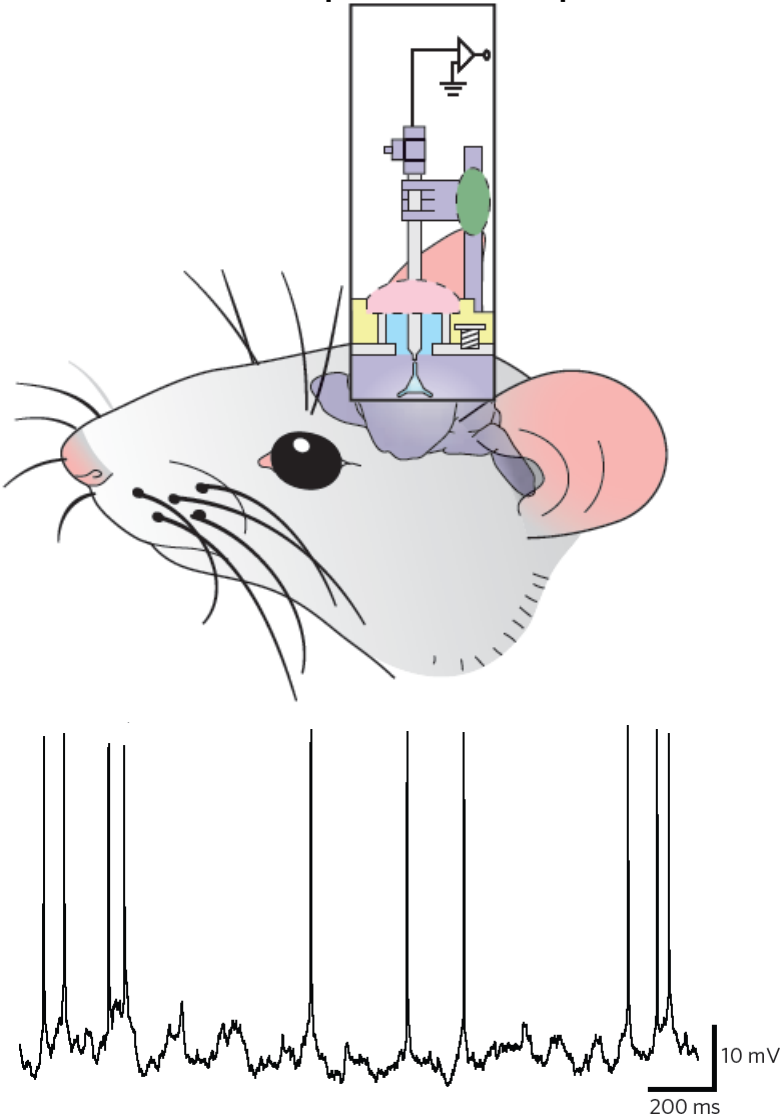
↓ Stimulation

② Depolarization (Na<sup>+</sup> influx) → Voltage sensitive Ca<sup>2+</sup> channel (Ca<sup>2+</sup> influx)

③ Polarization (K<sup>+</sup> efflux)

# 1. Electrophysiology

Illustration of patch-clamp<sup>2)</sup>



Galvani first reported the direct stimulation of nerves over 200 years ago.<sup>1)</sup>

## advantage

Electrical activity is recorded directly.  
Time resolution.  
High sensitivity.

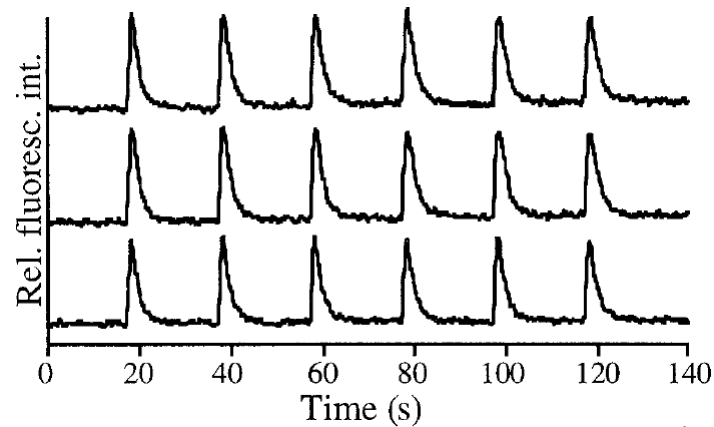
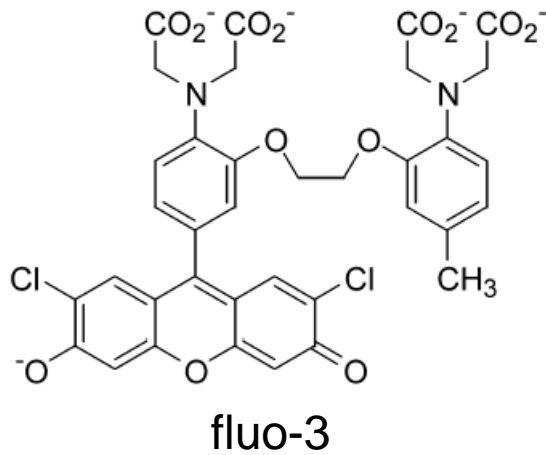
## disadvantage

Physical contact with the tissue is necessary.  
Spatial resolution.

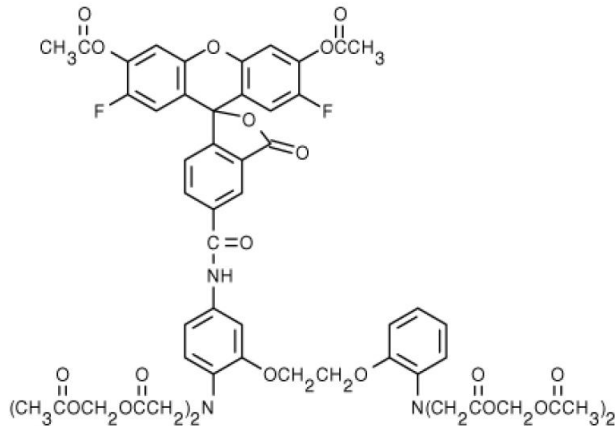
(1) Galvani, L. *Bonon. Sci. Art. Inst. Acad.* **1791**, 7, 364-415.

(2) Scanziani, M.; Häusser Michael *Nature* **2009**, 461, 930-939.

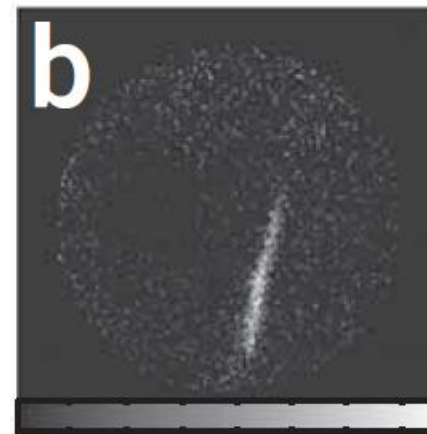
## 2. $\text{Ca}^{2+}$ -sensitive probes



Fluorescence change of fluo-3.<sup>1)</sup>



Oregon Green<sup>®</sup>488 BAPTA-1, AM



Imaging of  $\text{Ca}^{2+}$  spiking.<sup>2)</sup>

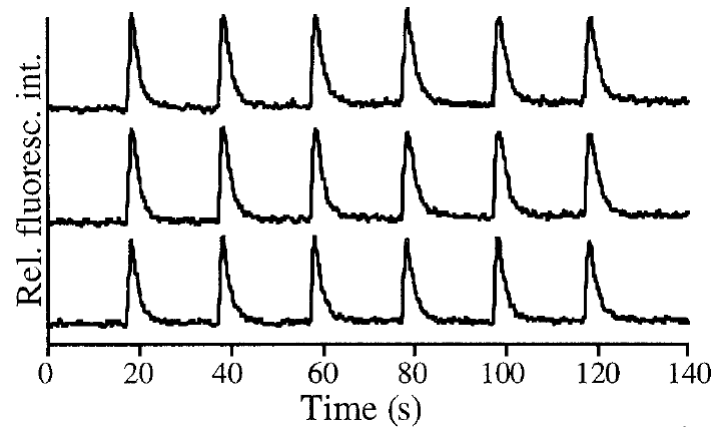
(1) Jacobs, J. M.; Meyer, T. *J. Neurosci.* **1997**, *17*, 4129-4135.

(2) Flusberg, B. A.; Nimmerjahn, A.; Cocker, E. D.; Mukamel, E. A.; Barretto, R. P. J.; Ko, T. H.; Burns, L. D.; Jung, J. C.; Schnitzer, M. J. *Nature Methods* **2008**, *5*, 935-938.

## 2. Ca<sup>2+</sup>-sensitive probes

### advantage

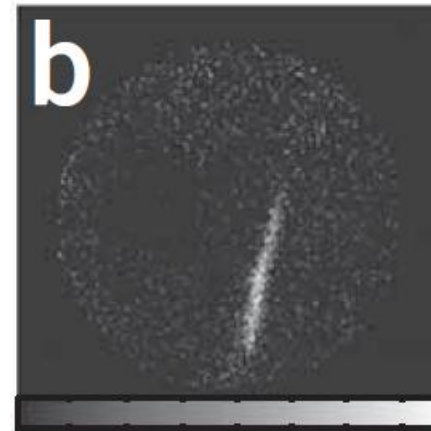
Spatial resolution.  
High sensitivity.



Fluorescence change of fluo-3.<sup>1)</sup>

### disadvantage

Electrical activity is **not** recorded directly.  
Slow time course of the intracellular calcium signal (time resolution).



Imaging of Ca<sup>2+</sup> spiking.<sup>2)</sup>

(1) Jacobs, J. M.; Meyer, T. *J. Neurosci.* **1997**, *17*, 4129-4135.

(2) Flusberg, B. A.; Nimmerjahn, A.; Cocker, E. D.; Mukamel, E. A.; Barretto, R. P. J.; Ko, T. H.; Burns, L. D.; Jung, J. C.; Schnitzer, M. J. *Nature Methods* **2008**, *5*, 935-938.



# Short summary

## 1. Electrophysiology

### advantage

Electrical activity is recorded directly.  
Time resolution.  
High sensitivity.

### disadvantage

Physical contact with the tissue is necessary.  
Spatial resolution.

## 2. Ca<sup>2+</sup>-sensitive probes

### advantage

Spatial resolution.  
High sensitivity.

### disadvantage

Electrical activity is **not** recorded directly.  
Slow time course of the intracellular calcium signal (time resolution).

## 3. Voltage-sensitive probes

### advantage

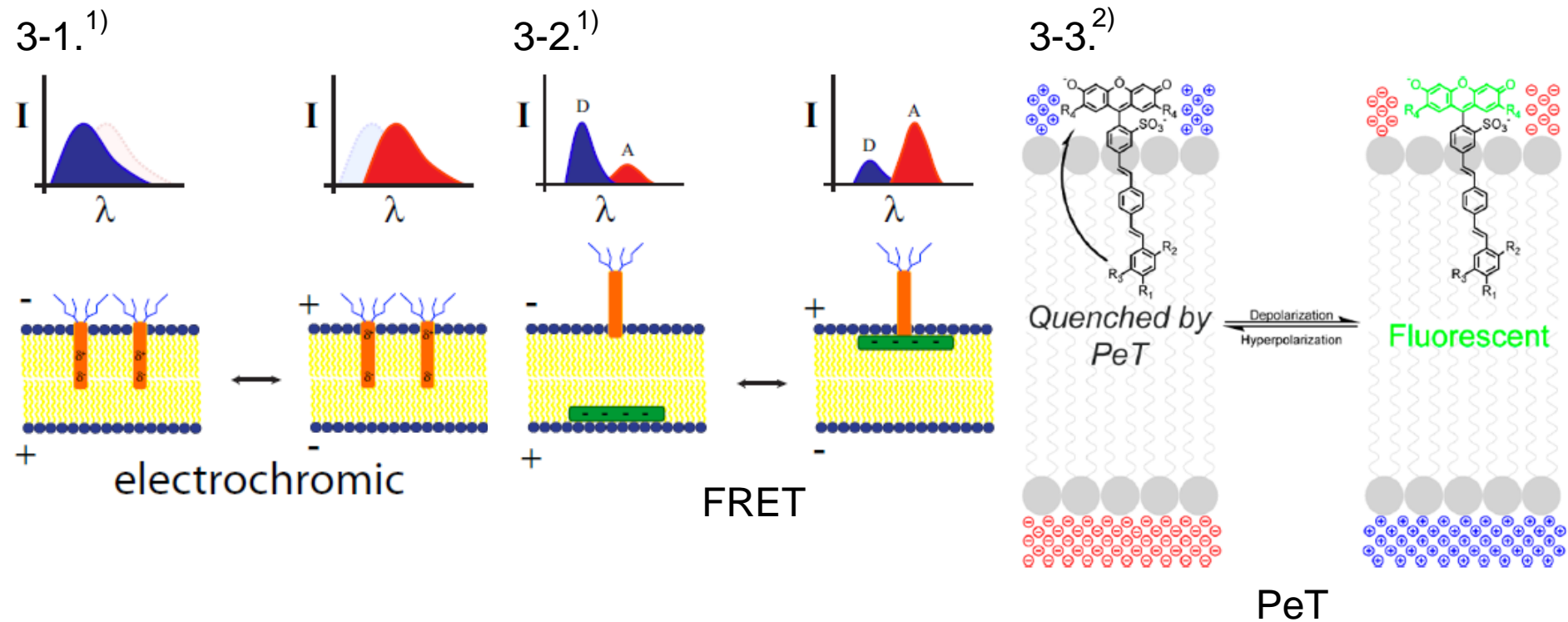
Electrical activity is recorded directly.

Time resolution?

Spatial resolution?

Sensitivity? (electron -> photon)

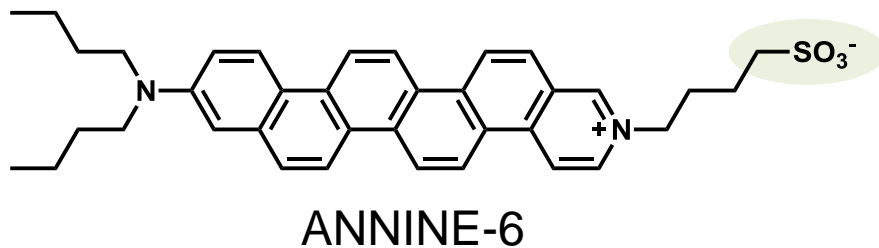
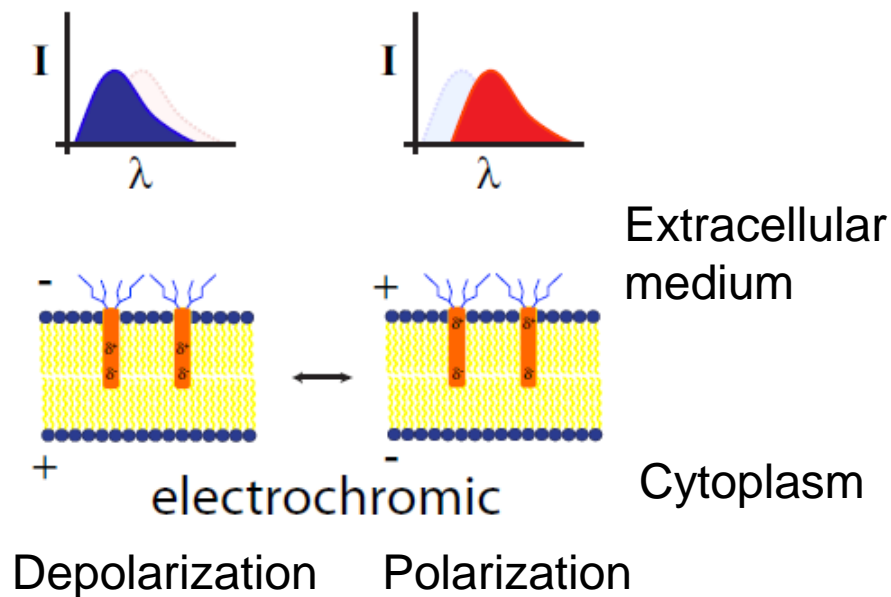
### 3. Voltage-sensitive probes



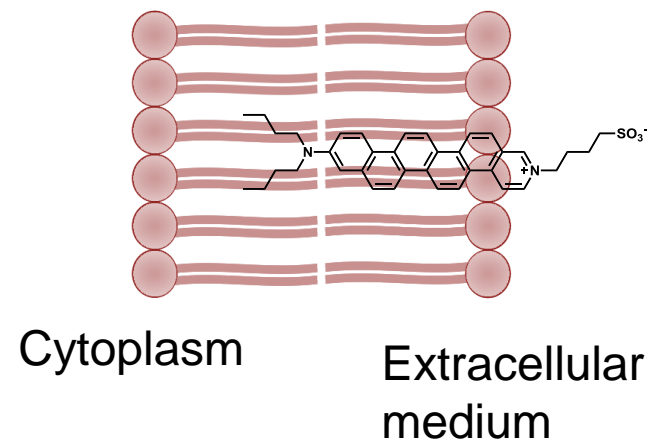
(1) Peterka, D. S.; Takahashi, H.; Yuste, R. *Neuron* **2011**, *69*, 9-21.

(2) Woodford, C. R.; Frady, E. P.; Smith, R. S.; Morey, B.; Canzi, G.; Palida, S. F.; Araneda, R. C.; Kristan, Jr., W. B.; Kubiak, C. P.; Miller, E. W.; Tsien, R. Y. *J. Am. Chem. Soc.* **2015**, *137*, 1817-1824.

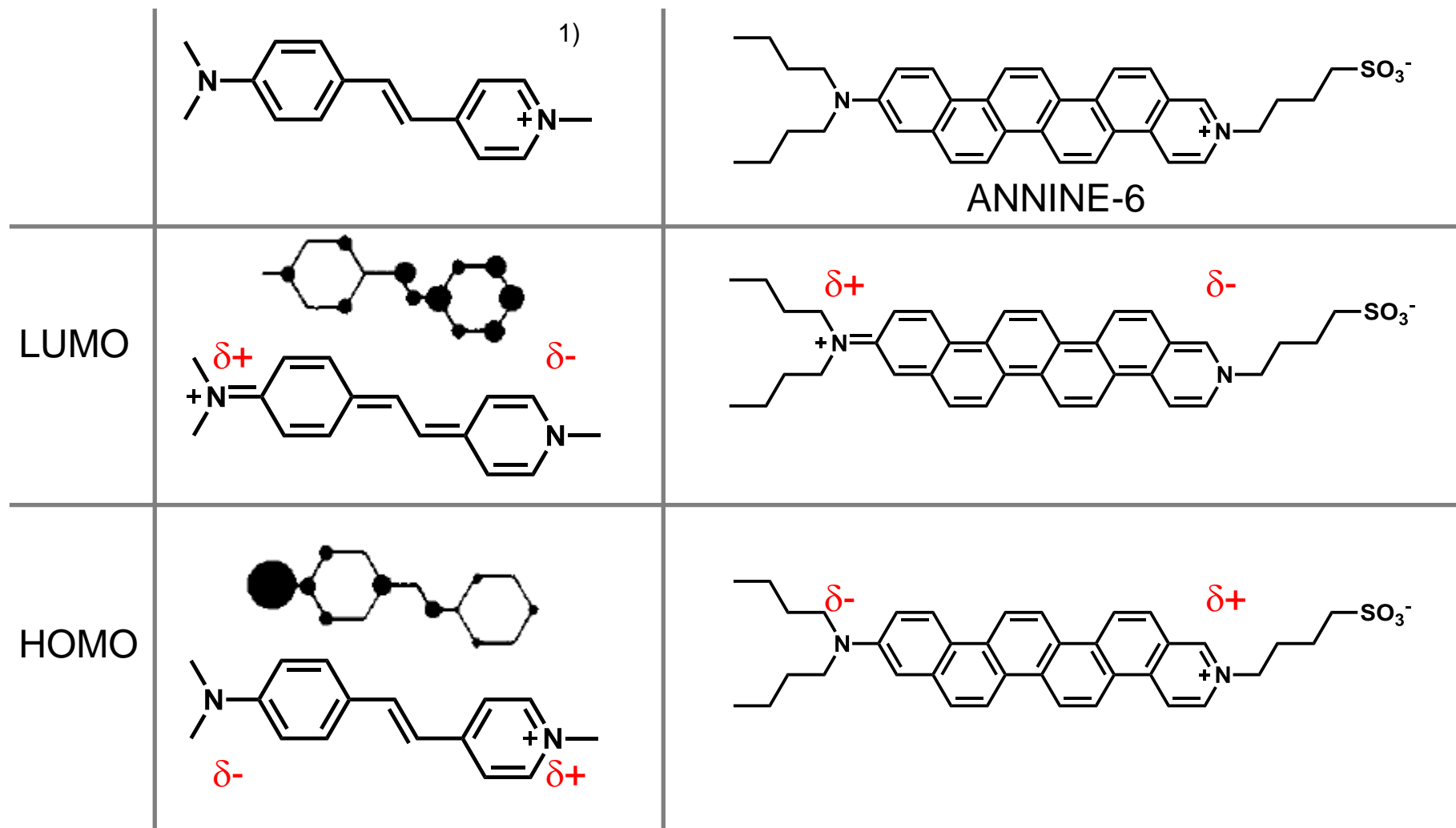
### 3-1. Electrochromic -ANNINE-6-



Prevent internalization.  
Determine direction of the molecule.

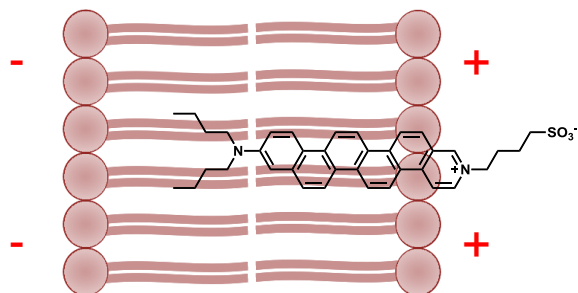


## 3-1. Electrochromic -HOMO, LUMO-



# 3-1. Electrochromic -energy shift-

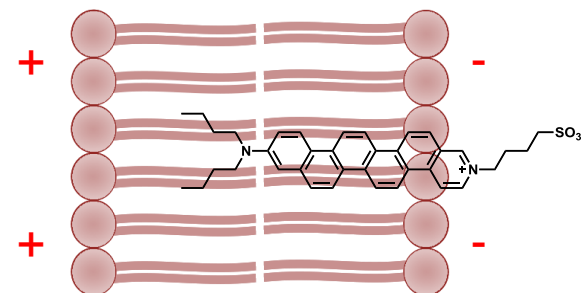
## Polarization



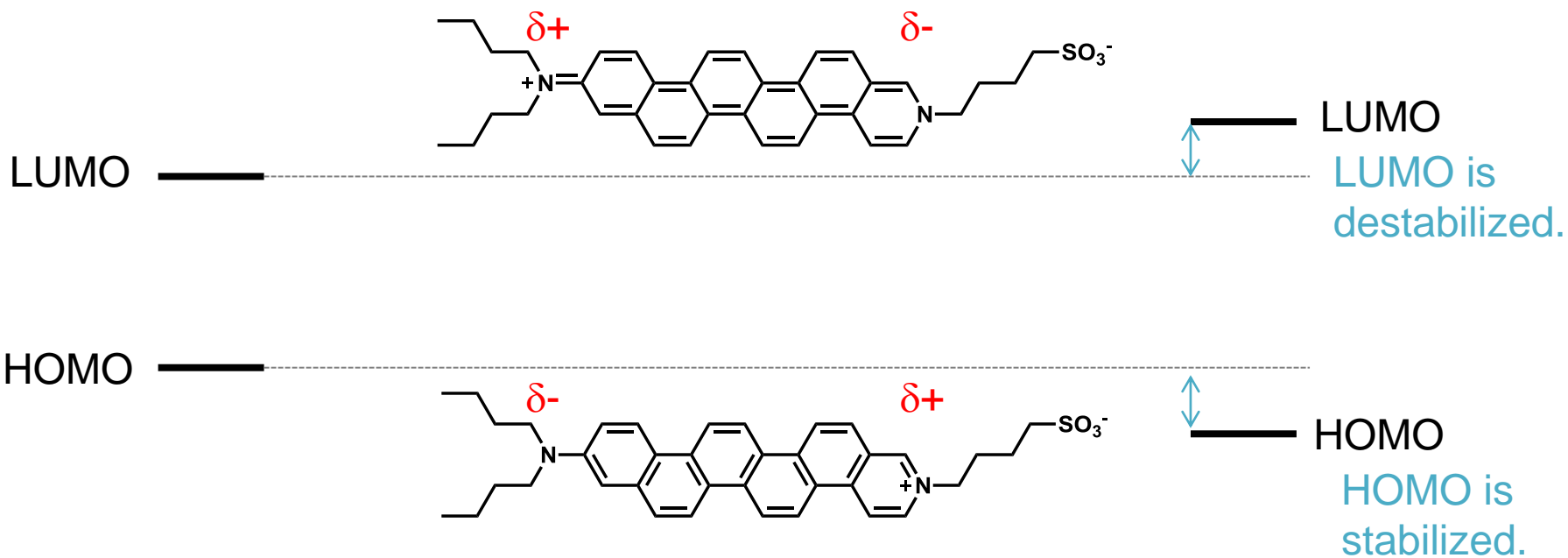
Cytoplasm

Extracellular  
medium

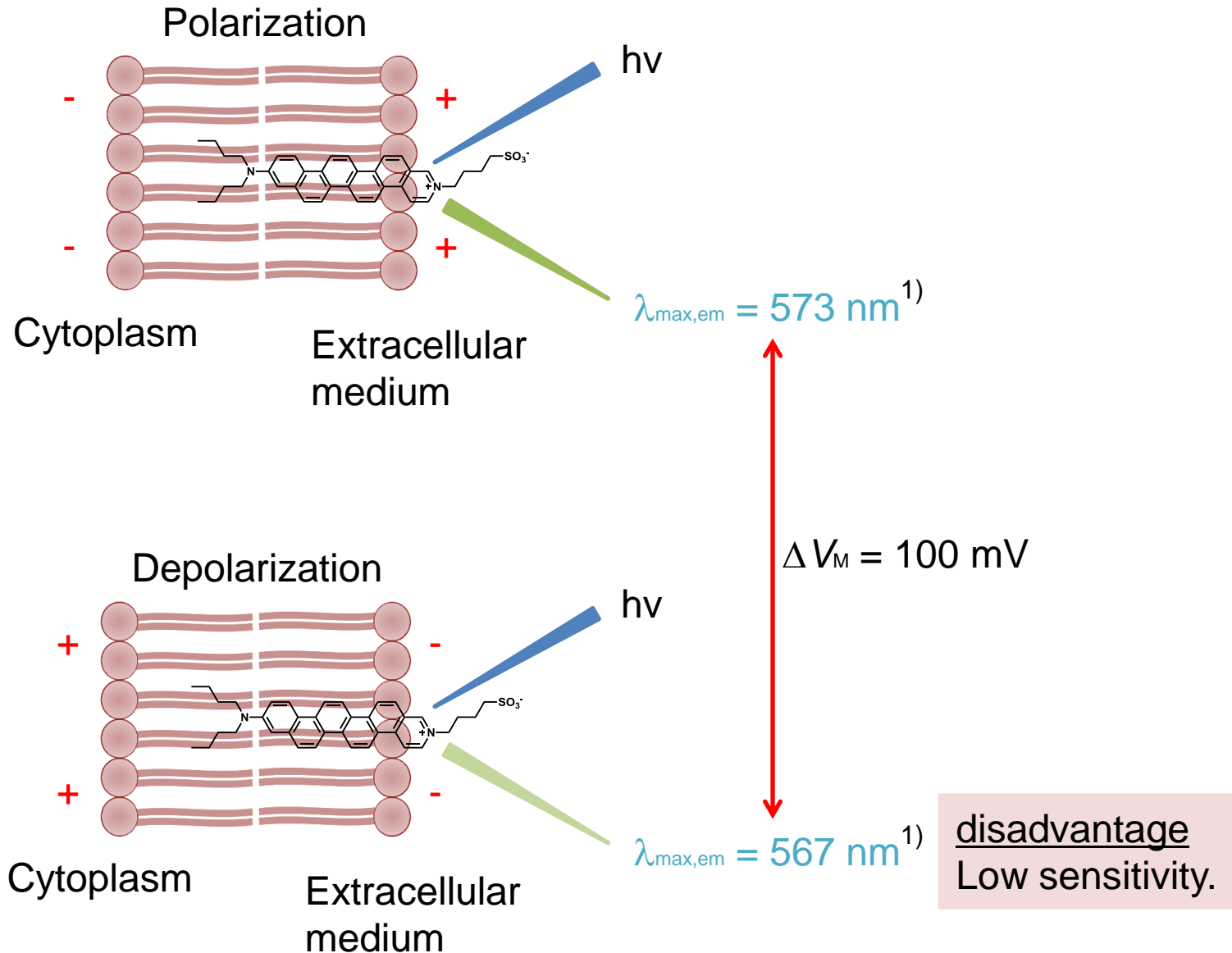
## Depolarization



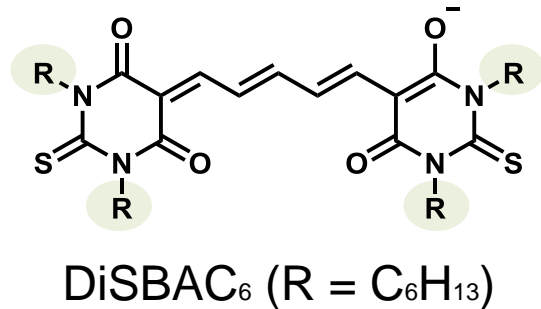
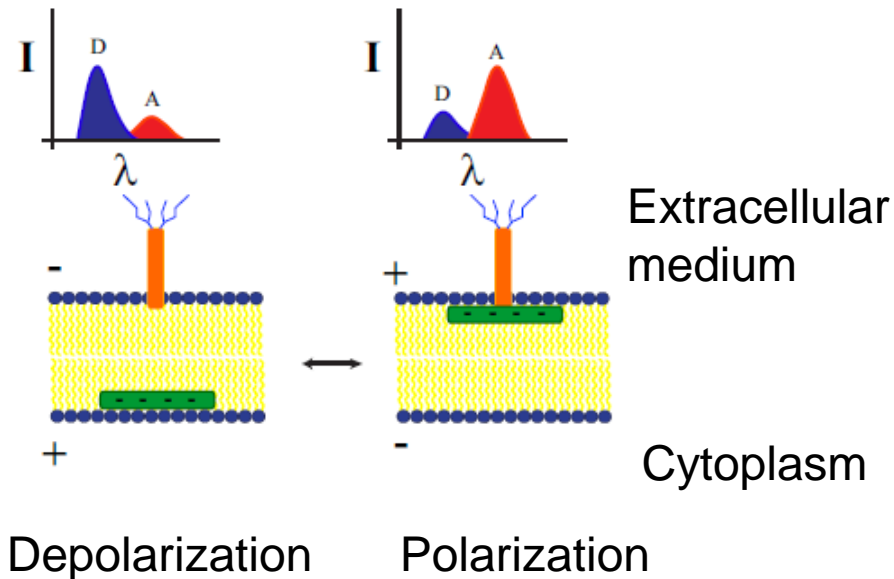
Cytoplasm

Extracellular  
medium

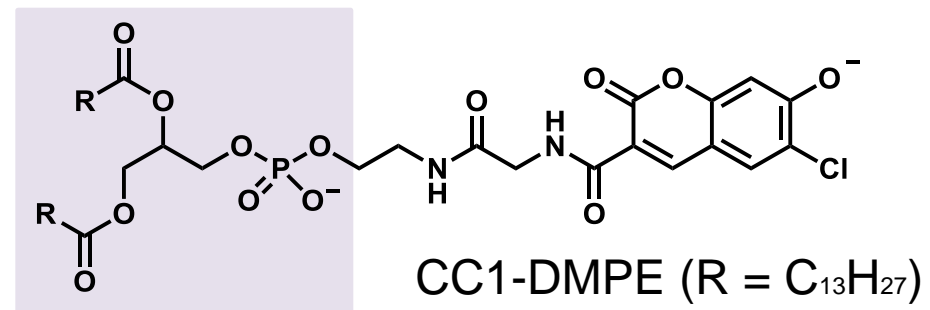
### 3-1. Electrochromic -disadvantage-



(1) Kuhn, B.; Fromherz, P. *J. Phys. Chem. B* **2003**, *107*, 7903-7913.

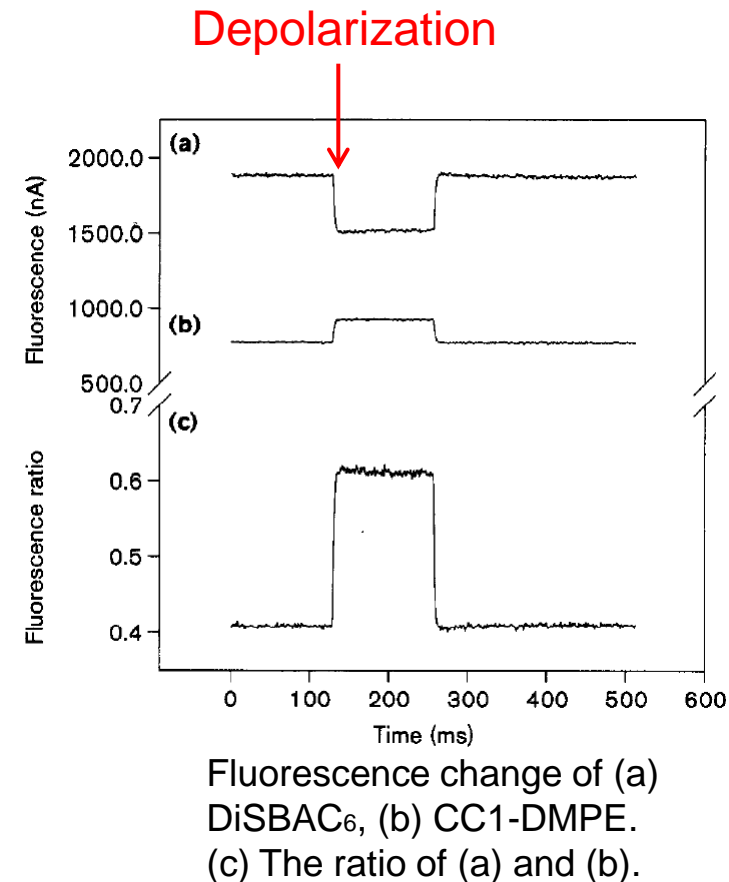
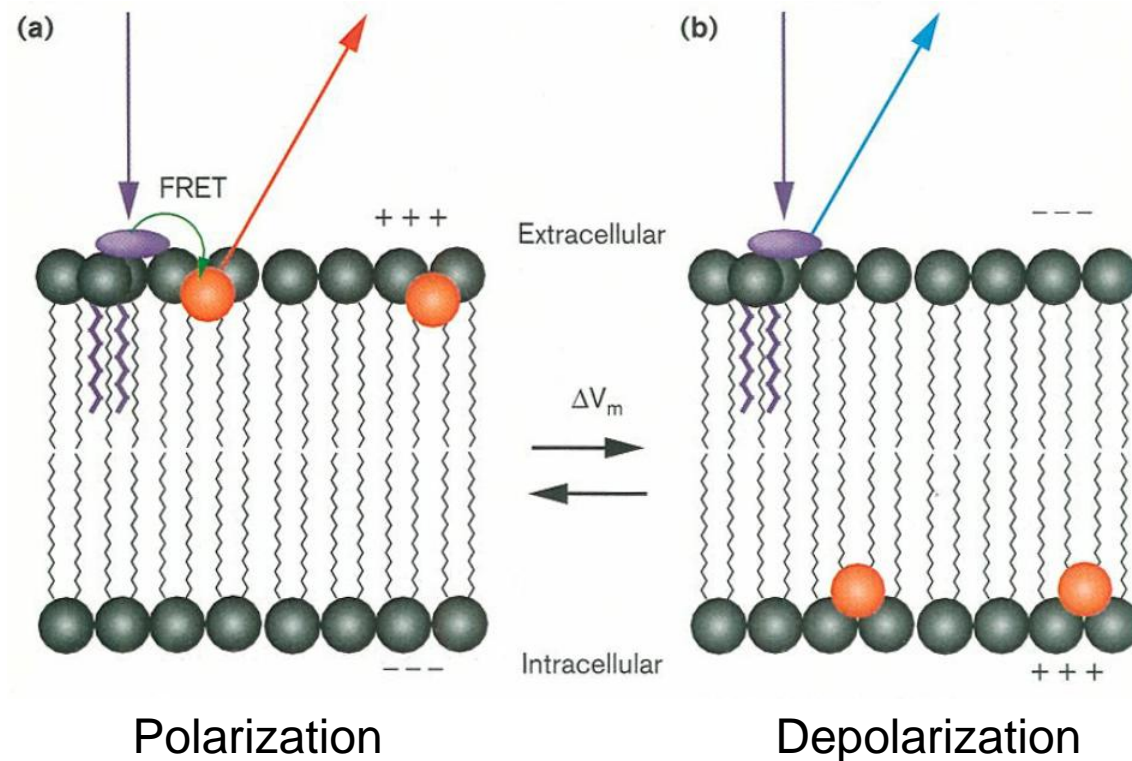
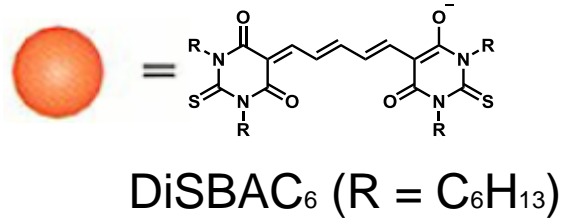
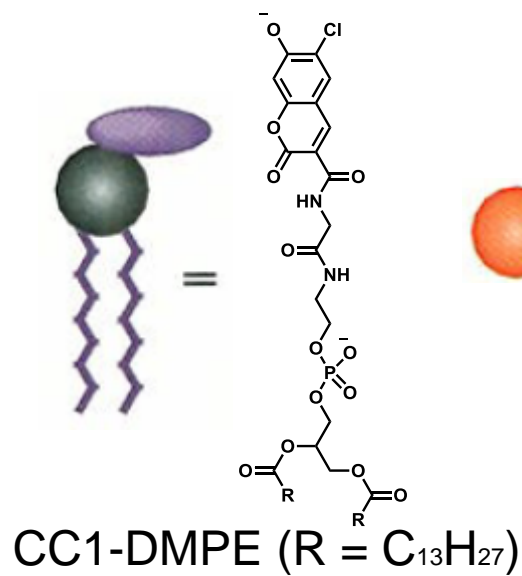
3-2. FRET -DisBAC<sub>6</sub> and CC1-DMPE-

Increasing lipophilicity.  
Localization to the membrane.



Anchor to the cell membrane.  
Determine direction of the molecule.

## 3-2. FRET -explanation-





## 3-2. FRET -advantage and disadvantage-

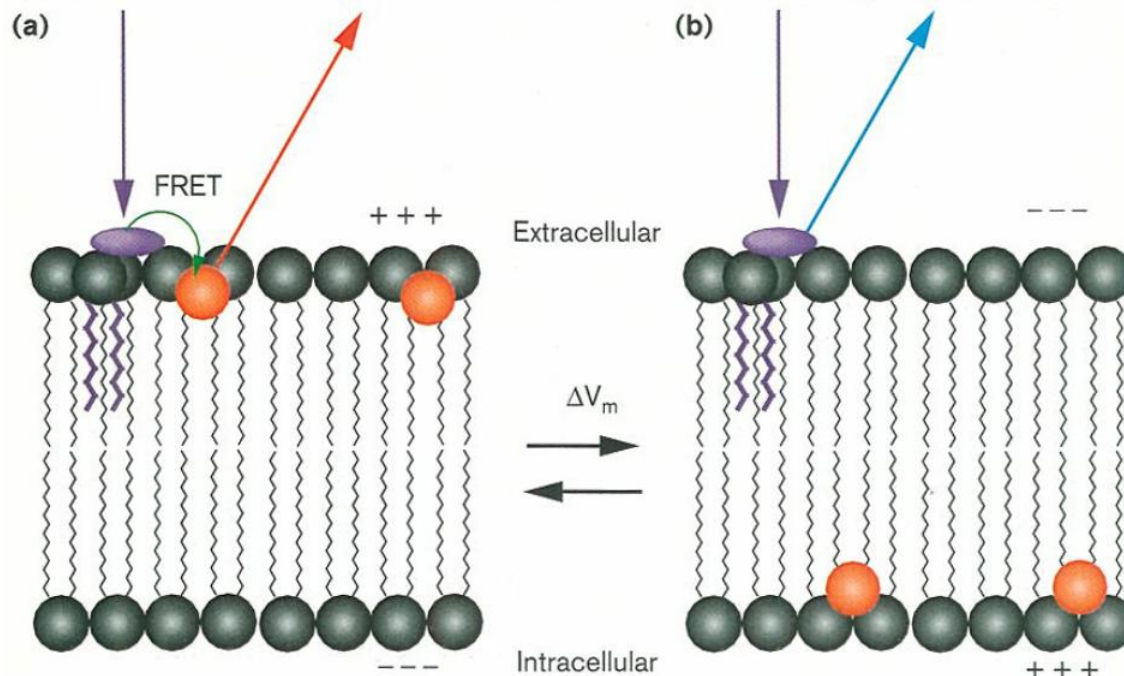
### advantage

High sensitivity.  
Spatial resolution.

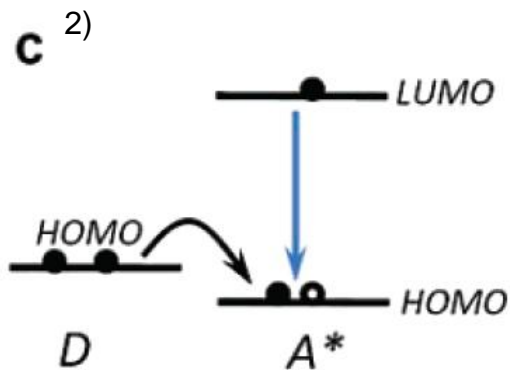
### disadvantage

Slow translocation of mobile charges (time resolution).

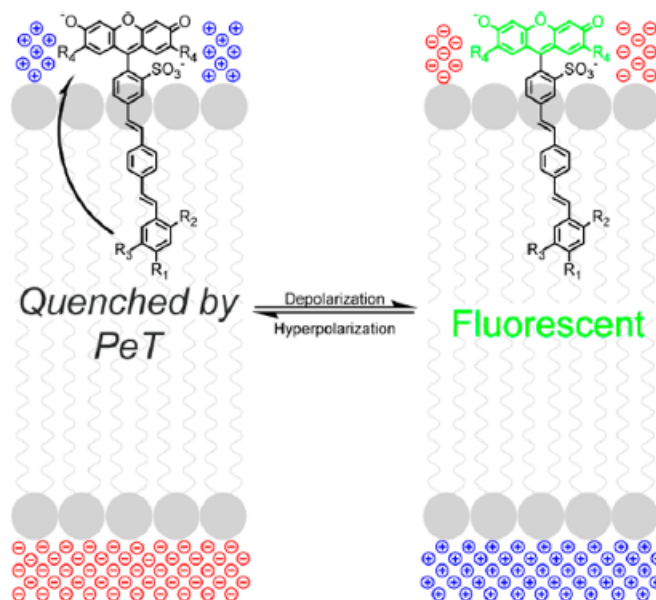
Changing the membrane capacitance.



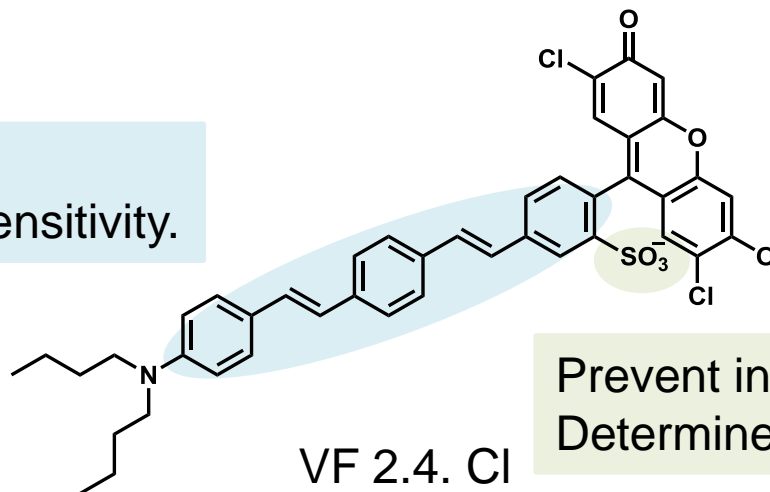
## 3-3. PeT -VF 2.4. Cl-



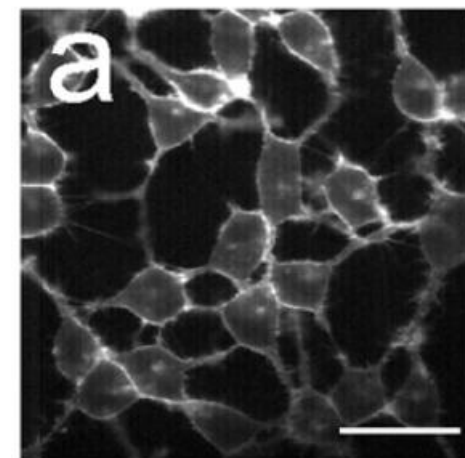
Mechanism of PeT.



Long molecular wire.  
Increasing voltage-sensitivity.



Prevent internalization.  
Determine direction of the molecule.



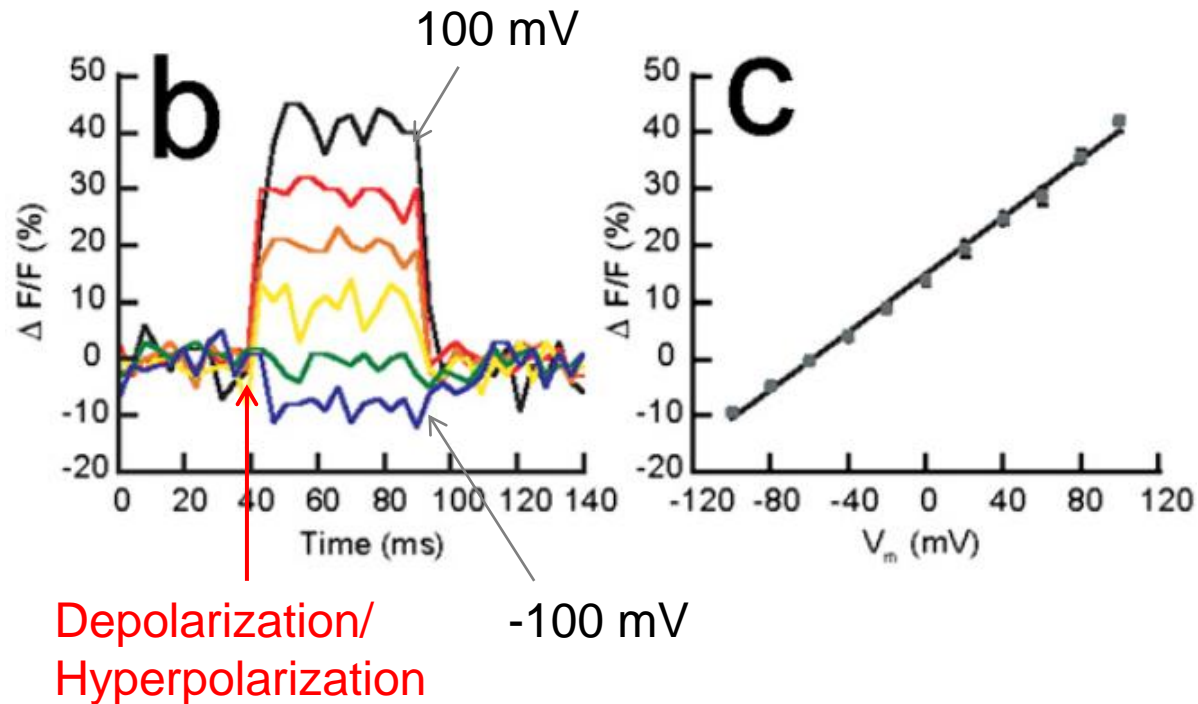
HEK 293 cells stained  
with 2  $\mu$ M VF 2.4. Cl

(1) Miller, E. W.; Lin, J. Y.; Frady, E. P.; Steinbach, P. A.; Kristan, Jr. W. B.; Tsien, R. Y. *P Natl. Acad. Sci. U.S.A.* **2012**, *109*, 2114-2119.

(2) Li, L-s. *Nano Lett.* **2007**, *7*, 2981-2986.

### 3-3. PeT -voltage sensitivity-

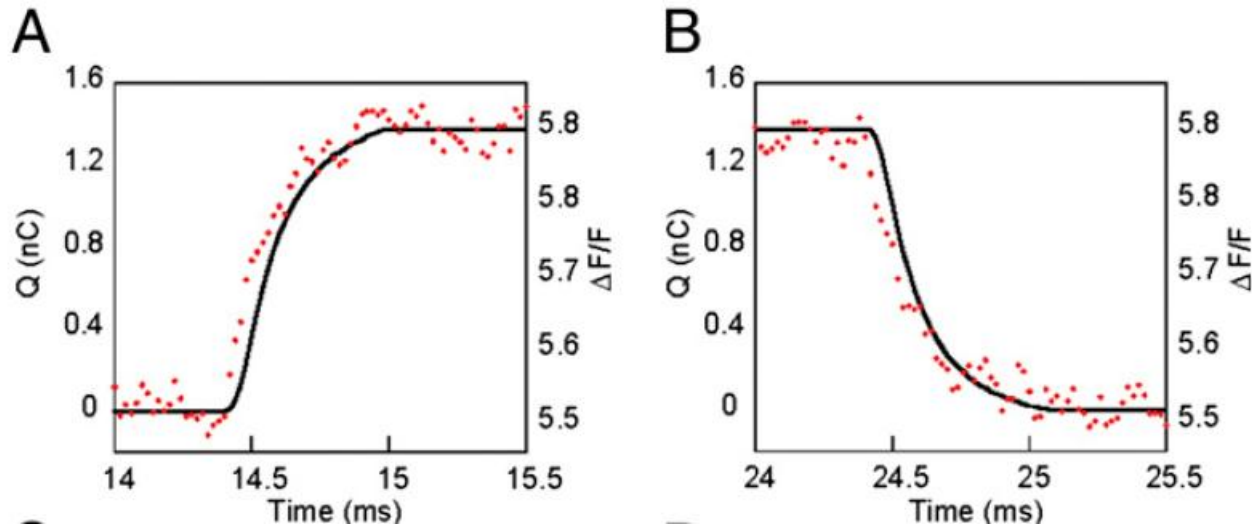
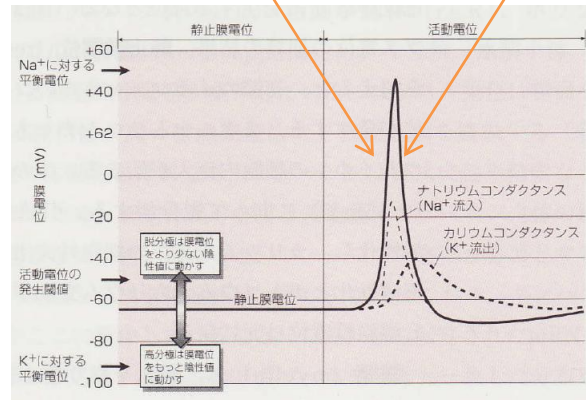
Voltage-sensitivity



- (b) Fractional changes in VF 2.4. Cl fluorescence during a series of voltage steps to +100 or -100 mV from a holding potential of -60 mV.
- (c) Fractional changes in VF 2.4. Cl fluorescence from (b) plotted against membrane potential .

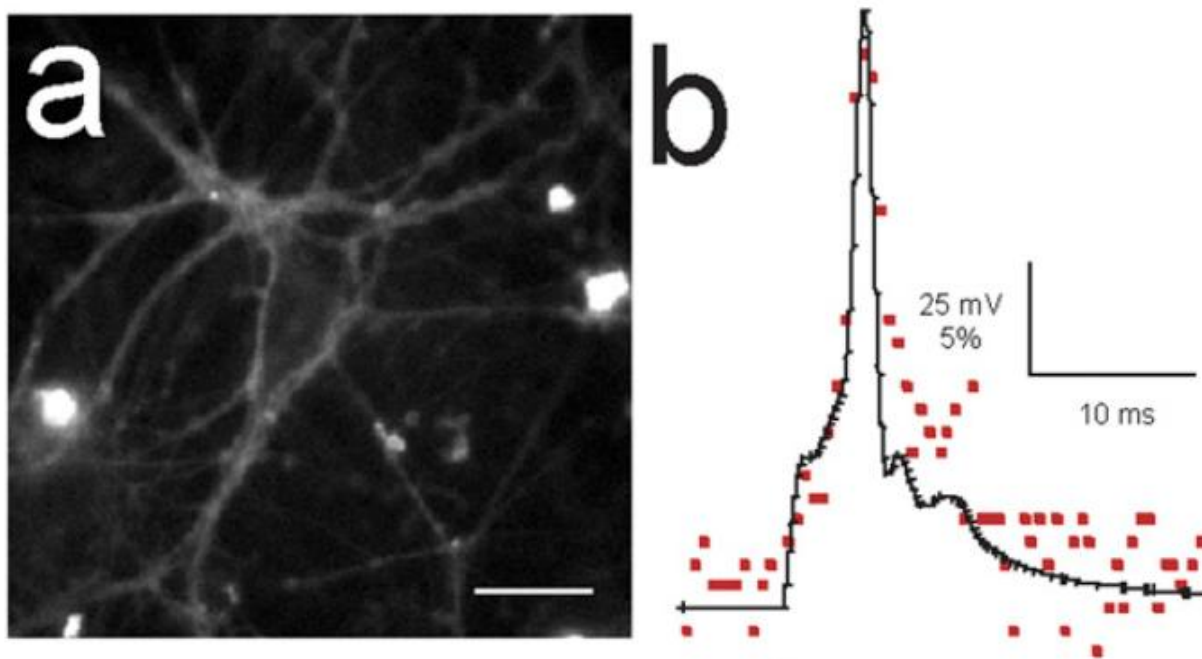
## Kinetics of signals (time resolution)

## 3-3. PeT -time resolution- Rising edge Falling edge



(A) Rising edge and (B) Falling edge of a 100-mV depolarizing step from -60 mV in HEK cells stained with VF 2.4. Cl.

Black, solid trace is the integrated current measured electrophysiologically.  
Red points are the optical recording.



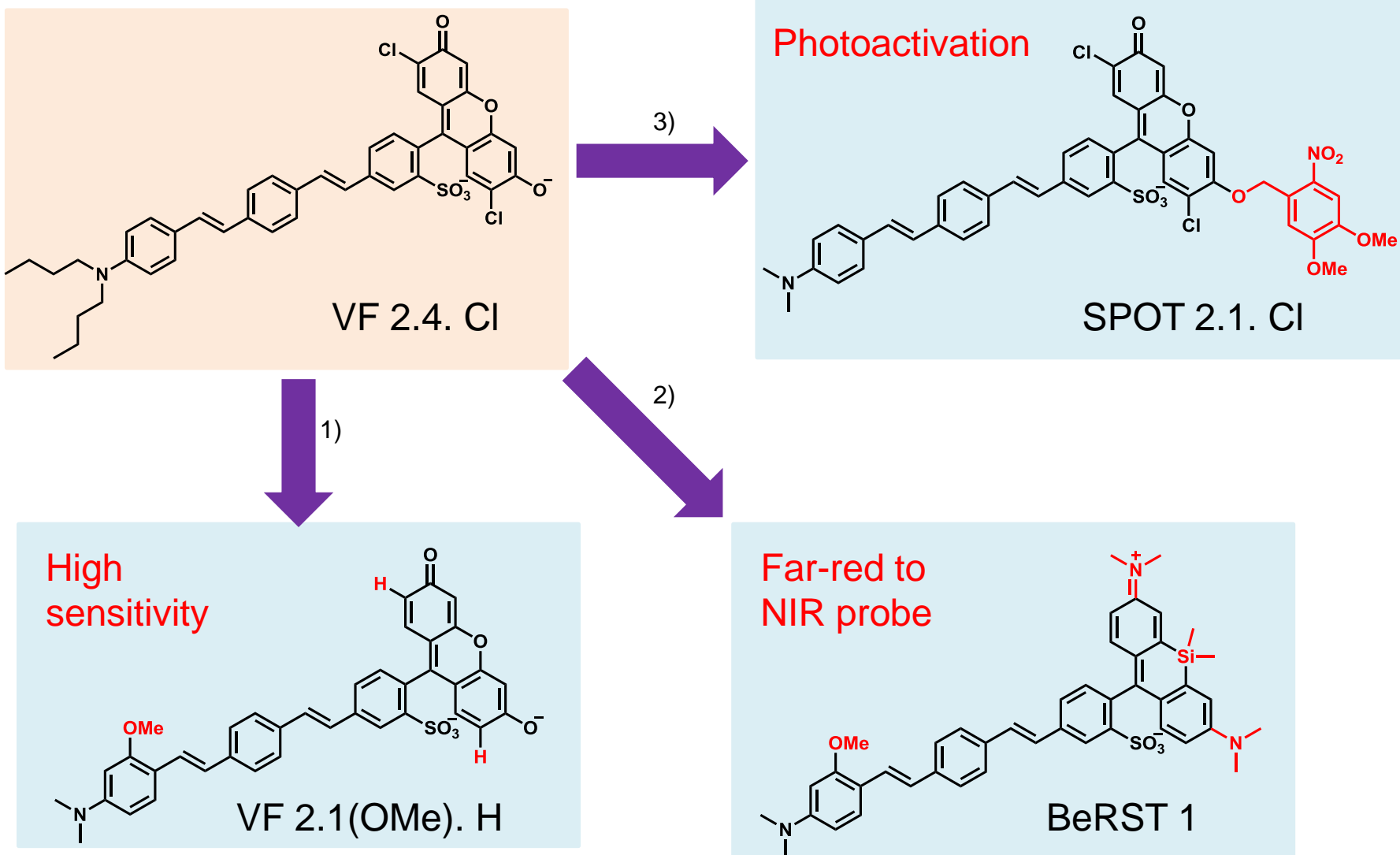
(A) Rat hippocampal neurons stained with 2  $\mu$ M VF 2.4. Cl.

(B) VF 2.4. Cl can detect evoked action potentials in rat hippocampal neurons in single trials.

The black trace is the recorded electrophysiology signal.

Individual points represent the optical signal from VF 2.4. Cl.

# PeT-based probes



(1) Woodford, C. R.; Frady, E. P.; Smith, R. S.; Morey, B.; Canzi, G.; Palida, S. F.; Araneda, R. C.; Kristan, Jr. W. B.; Kubiak, C. P.; Miller, E. W. Tsien, R. Y. *J. Am. Chem. Soc.* **2015**, *137*, 1817-1824.

(2) Huang, Y-L.; Walker, A. S.; Miller, E. W. *J. Am. Chem. Soc.* **2015**, *137*, 10767-10776.

(3) Grenier, V.; Walker, A. S.; Miller, E. W. *J. Am. Chem. Soc.* **2015**, *137*, 10894-10897.

# Summary

## 3. Voltage-sensitive probes

### advantage

Electrical activity is recorded directly.

Time resolution?

Spatial resolution?

Sensitivity? (electron -> photon)

### 3-1. Electrochromic

#### disadvantage

Low sensitivity.

### 3-2. FRET

#### advantage

High sensitivity.  
Spatial resolution.

#### disadvantage

Slow translocation of mobile charges (time resolution).  
Changing the membrane capacitance.

### 3-3. PeT

#### advantage

Electrical activity is recorded directly.  
Time resolution.  
Spatial resolution.  
High sensitivity.

#### Future perspective

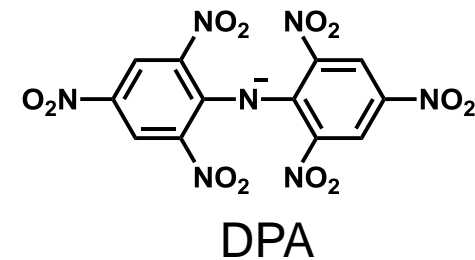
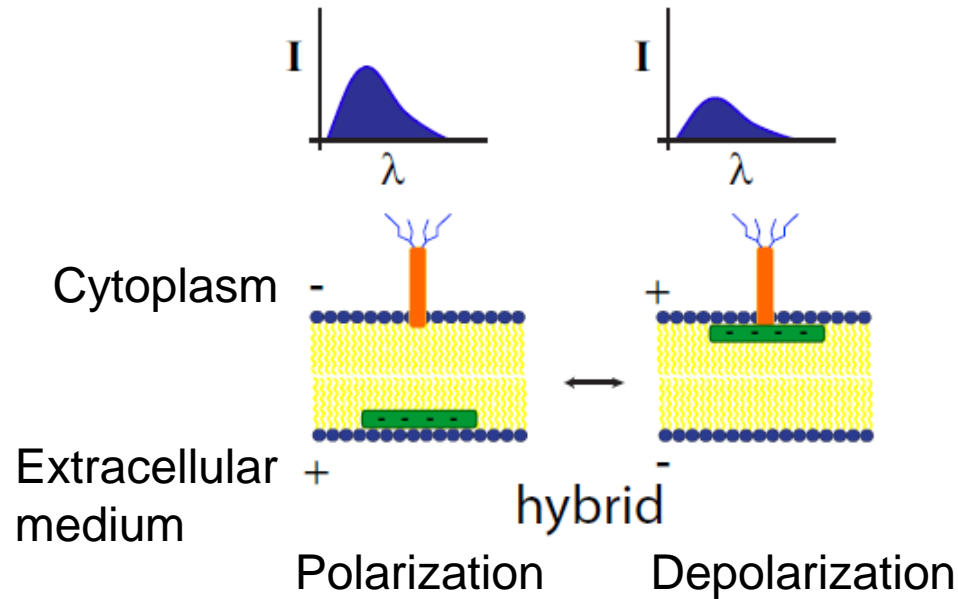
Selective targeting

Genetic expression of PeT-based pH<sup>-1</sup> sensitive protein sensor was reported.



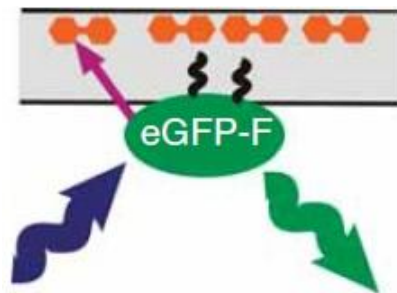


# A1. FRET + Genetic Expression

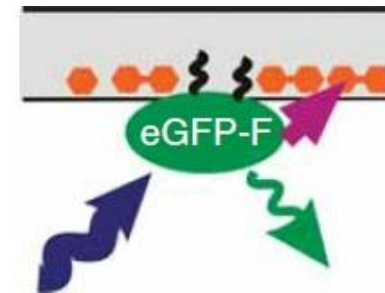


Extracellular medium

Cytoplasm



Polarization



Depolarization

## A2. PeT based pH-sensitive probe

